

REFERENCES

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- Abdelhak, R. and Soraya, B. (2018). Phytochemical characterization, antiinflammatory and anti-ulcer activity of a spontaneous succulent *Delosperma reseii*. *Universal Journal of Agricultural Research*. 6(3): 113-117.
- Abdel-Karim, O.H., Abo-Shady, A.M., Gheda, S.F. and Ismail, G.A. (2019). In vitro potential activity of some seaweed as antioxidants and inhibitors of diabetic enzymes. *Food Science and Technology (Brazil)*. 40(3): 681-691.
- Abdou Bouba, A., Ponka, R., Augustin, G., Njintang Yanou, N., AbulHamd El-Sayed M, Montet, D., Scher, J. and Mbofung, C.M. (2016). Amino acid and fatty acid profile of twenty wild plants used as spices in Cameroon. *American Journal of Food Science and Technology*. 4 (2): 29-37.
- Abdulaziz, A.A., Dapar, A.L.G., Manting, M.M.M., Torres, M.A.J., Aranas, A.T., Mindo, R.A.R., Cabrido, C.K. and Demayo, C.G. (2019). Qualitative evaluation of the antimicrobial, antioxidant and medicinally important phytochemical constituents of the ethanolic extracts of the leaves of *Gliricidia sepium* (Jacq.) Walp. *Pharmacophore*. 10(4): 72-83.
- Aberoumand, A. and Deokule, S.S. (2009). Determination of Elements Profile of Some Wild Edible Plants. *Food Analytical Methods*. 2(2): 116- 119.
- Abifarín, T.O., Otunola, G.A. and Afolayan, A.J. (2020). Nutritional composition and antinutrient content of *Heteromorpha arborescens* (Spreng.) Cham. & Schltdl. leaves: An underutilized wild vegetable. *Food Science and Nutrition*. 9(1): 172-179.
- Abu-Rabia, A. (2005). Urinary diseases and ethnobotany among pastoral nomads in the Middle East. *Journal of Ethnobiology*. 1(4): 1-3.
- Acharyaa, S.N. and Patelb, J.J. (2016). Quality assessment and phytochemical analysis of *Clerodendrum serratum* roots. *International Journal of Phytopharmacy*. 6(3): 51-57.
- Achi, N.K., Onyeabo, C., Ekeleme-Egedigwe, C.A. and Onyeonula, J.C. (2017). Phytochemical, proximate analysis, vitamin and mineral composition of aqueous extract of *Ficus capensis* leaves in South Eastern Nigeria. *Journal of Applied Pharmaceutical Science*. 7(3): 117-22.
- Adam, O.A.O., Mohmmmed, R.S.A. and Hussein, S.M.A. (2019). The Effect of extraction method and solvents on yield and antioxidant activity of certain Sudanese medicinal plant extracts. *The Journal of Phytopharmacology*. 8(5): 248-252.

- Adamu, E., Asfaw, Z., Demissew, S. and Baye, K. (2022). Antioxidant Activity and Anti-Nutritional Factors of Selected Wild Edible Plants Collected from Northeastern Ethiopia. *Foods*. 11(2291): 1-12.
- Adane, T., Shimelis, A., Negussie, R., Tilahun, B. and Haki, G.D. (2013). Effect of processing method on the Proximate composition, mineral content and antinutritional factors of Taro (*Colocasia esculenta*, L.) growth in Ethiopia. *African Journal of Food, Agriculture, Nutrition and Development*. 13(5): 7383-7396.
- Adebayo, A.H., Tan, N.H., Akindahunsi, A. A., Zeng, G. Z. and Zhang., Y. M. (2010). Anticancer and antiradical scavenging activity of *Ageratum conyzoides* L. (Asteraceae). *Pharmacognosy Magazine*. 6 (21): 62-66.
- Adedapo, A.A., Jimoh, F.O., Koduru, S., Afolayan, A.J. and Masika, P.J. (2008). Antibacterial and antioxidant properties of the methanol extracts of the leaves and stems of *Calpurnia aurea*. *BMC Complementary and Alternative Medicine*. 8(53): 1-58.
- Adhikary, P. and Barman, D. (2014). The Indigenous medicinal knowledge of the Rabha Tribe: A Case study in the Chirang District of Assam, North-East India. *Life Science Leaflets*. 52(48): 94-97.
- Agarwal, Y.S. and Ghosh, B. (1985). *Drug Plants of India (Root Drugs)*. Kalyani Publishers, New Delhi.
- Agrawal, A. (1995). Indigenous and scientific knowledge: Some critical comments. *IK Monitor*. 3(3): 1-9.
- Agrawal, A. (2008). Dismantling the divide between indigenous and scientific knowledge. *Development and Change*. 26(3): 413-439.
- Ahmadiani, A., Hosseiny, J., Semnianian, S., Javan, M., Saeedi, F., Kamalinejad, M. and Saremi, S. (2000). Anti-nociceptive and Anti-inflammatory Effects of *Eleagnus angustifolia* Fruit Extract. *Journal of Ethnopharmacology*. 72(1-2): 287-292.
- Ahmed, A.S., Khafagy, E.S., Mohammed, F.A., Almansour, K. and Lila, A.S.A. (2022). Screening of Apoptosis Pathway-Mediated Anti-Proliferative Activity of the Phytochemical Compound Furanodienone against Human Non-Small Lung Cancer A-549 Cells. *Life*. 12(1): 1-16.
- Ahmed, V.U. and Alam, N. (1995). New antifungal bithienylacetylenes from *Blumea obliqua*. *Journal of Natural Product*. 58(9): 1426-1429.
- Ahmed, F.A., Mohamed, M.A., P, Abdel-Aziem, A. and El-Azab, M.M. (2017). Proximate composition, amino acids and sugar contents of leaves and stems of

Ficus cordata thunb. Subsp. *Salicifolia* (VAHL). *International Journal of Innovative Science, Engineering & Technology*. 4(11): 25-33.

Ainsworth, E.A. and Gillespie, K.M. (2007). Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin Ciocalteu reagent. *Nature Protocols*. 2(4): 875-877.

Akbar, M., Ali, U., Khalil, T., Iqbal, M.S., Amin, A., Naeem, R., Nazir, A., Waqas, H.M., Aslam, Z., Jafri, F.I., Aslam, N. and Chohan, S.A. (2020). *Cornus macrophylla*, the antibacterial activity of organic leaf extracts and the characterization of the more lipophilic components by GC/MS. *Molecules*. 25(10): 1-11.

Akindahunsi, A.A. and Salawu, S.O. (2005). Phytochemical screening and nutrient-antinutrient composition of selected tropical green vegetables. *African Journal of Biotechnology*. 4(6): 497-501.

Akintimehin, E., Onoagbe, I. and Abu, O. (2022). Proximate Composition, Qualitative Phytochemical Screening and In-Vitro Antioxidant Potential of *Triumfetta rhomboidea* Leaves Extracts. *Trends in Sciences*. 19(24): 1-10.

Akpabio, U.D. and Ikpe, E.E. (2013). Proximate composition and nutrient analysis of *Aneilema aequinoctiale* leaves. *Asian Journal of Plant Science and Research*. 3(2): 55-61.

Akpuaka, A., Ekwenchi, M.M., Dashak, D.A. and Dildar, A. (2013). Biological activities of characterized isolated of n-hexane extract of *Azadirachta indica* a juss (Neem) leaves. *Newyork Science Journal*. 6(6): 119-124.

Akubugwo, I.E., Obasi, N.A., Chinyere, G.C. and Ugbo, A.E. (2007). Nutritional and chemical value of *Amaranthus hybridus* L. leaves from Afikpo, Nigeria. *African Journal of Biotechnology*. 6(24): 2833-9.

Al-Abd, N.M., Mohamed, Z.N., Mansor, M., Azhar, F., Hasan, M.S. and Kassim, M. (2015). Antioxidant, antibacterial activity, and phytochemical characterization of *Melaleuca cajuputi* extract. *BMC Complementary Alternative Medicine*. 13(1): 1-13.

Alagbe, J.O., Adeoye, A. and Oluwatobi, A.O. (2020). Proximate and Mineral Analysis of *Delonix regia* Leaves and Roots. *Annals of Nutrition & Food Science*. 4(1): 104.

Alam, A.H.M.K., Ali, A.S.M.R., Haque, M.A., Sarker, J., Nasrin, M.S., Rahman, M.M., Tareq, A.M., Khan, Z., Rashid, M., Sadik, M.G. and Tsukahara, T. (2021).

Antiproliferative and antioxidant potentials of bioactive edible vegetable fraction of *Achyranthes ferruginea* Roxb. in cancer cell line. *Food Science and Nutrition*. 9(7): 3777-3805.

Alamery, S.F. and Algaraawi, N.I. (2020). Phytochemical profile and antifungal activity of stems and leaves methanol extracts from the *Juncus Maritimus* Linn. Juncaceae Family against some dermatophytes fungi. *AIP Conference Proceedings*. 2290 (1): 1-17.

Alegria, E.C.B.A., Ribeiro, A.P.C., Mendes, M., Ferraria, A.M., Rego, A.M.B. and Pombeiro, A.J.L. (2018). Effect of Phenolic Compounds on the Synthesis of Gold Nanoparticles and Its Catalytic Activity in the Reduction of Nitro Compounds. *Nanomaterials*. 8(5): 1-18.

Aletan, U.I. and Kwazo, H.A. (2019). Analysis of the proximate composition, anti-nutrients and mineral content of *Maerua crassifolia* leaves. *Nigerian Journal of Basic and Applied Science*. 27(1): 89-96.

Alexander, H.J. and Rosy, B.A. (2022). Profiling of Biologically Active Compounds from *Sansevieria Cylindrica* Bojer Ex Hook. using GCMS. *International Journal of Innovative Research in Science, Engineering and Technology*. 11(1): 527-531.

Ali, A., Waris, Z., Iqbal, Y., Hussain, A., Shafqatullah, Khan, A.A. and Khan, M.W. (2018). Proximate composition, phytochemical analysis and antioxidant capacity of *Aloe vera*, *Cannabis sativa* and *Mentha longifolia*. *Pure and Applied Biology*. 7(3): 1122-1130.

Altundag, H. and Tuzen, M. (2011). Comparison of dry wet and microwave digestion methods for the multi element determination in some dried fruit samples by ICP-OES. *Food and Chemical Toxicology*. 49(11): 2800-2807.

Amalarasi, L.E. and Jothi, G.J. (2019). BCL2 Mediated Apoptotic Induction Potential of the DNOP Isolated from *Pachygone ovata* (POIR.) Miers Ex Hook. F.& Thomson in MCF7 (Human Breast Carcinoma). *International Research Journal of Pharmacy*. 10(3): 190-194.

Amensour, M., Sendra, E., Perez-Alvarez, J.A., Skali-Senhaji, N., Abrini, J. and Fernandez-Lopez, J. (2010). Antioxidant activity and chemical content of methanol and ethanol extracts from leaves of rockrose (*Cistus ladaniferus*). *Plant Foods for Human Nutrition*. 65(2): 170-178.

- Amorin, D. A., Borba, H. R., Carauta, J. P., Lopes, D. and Kaplan, M. A. (1999). Anthelmintic activity of the latex of Ficus species. *Journal of Ethnopharmacology*. 64(3): 255-258.
- Andualem, B. and Gessesse, A. (2014). Proximate composition, mineral content and antinutritional factors of Brebra (*Millettia ferruginea*) seed flour as well as physicochemical characterization of its seed oil. *Springer Plus*. 3(1): 298.
- Ang, H. and Lee, K. (2005). Analysis of mercury in Malaysian herbal preparations: a peer review. *Biomedical Science*. 4(1): 31-6.
- Anila, L., Vijayalakshmi, N.R. and Tian, C. (2000). Beneficial effect of flavonoids from *Sesamum indicum*, *Emblica officinalis* and *Momordicacharantia*. *Phytotherapy Research*. 14(8): 592-595.
- Anis, M., Sharma, M.P. and Iqbal, M. (2000). Herbal ethnomedicine of the Gwalior forest division in Madhya Pradesh, India. *Pharmaceutical Biology*. 38(4): 241-253.
- Anwar, F., Sultana, B. and Muhammad, A. (2009). Effect of Extraction Solvent/Technique on the Antioxidant Activity of Selected Medicinal Plant Extracts. *Molecules*. 14(6): 2167-2180.
- Anyasor, G.N., Onajobi, F.D., Osilesi, O. and Adebawo, O. (2014). Proximate composition, mineral content and in vitro antioxidant activity of leaf and stem of *Costus afer* (Ginger lily). *Journal of Intercultural Ethnopharmacology*. 3(3): 128-134.
- AOAC (2005). Official methods of analysis. (18th Edn). Association of Official Analytical Chemists, Washington DC. pp. 935.14 and 992.24.
- Arabshahi-Delouee, S. and Urooj, A. (2007). Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L) leaves. *Food Chemistry*. 102(4): 1233-1240.
- Arakaki, D. G., Candido, C. J., Silva, A. F. D., Guimaraes, R. D. C. A. and Hiane, D. A. (2016). In vitro and in vivo antioxidant activity of the pulp of *Jatoba-do-cerrado*. *Food Science and Technology (Campinas)*. 36(1): 166-170.
- Armania, N., Yazan, L., Musa, S.N., Ismail, I.S., Foo, J.B., Chan, K.W., Noreen, H., Hisyam, A.H., Zulfahmi, S. and Ismail, M. (2013). *Dillenia suffruticosa* exhibited antioxidant and cytotoxic activity through induction of apoptosis and G2/M cell cycle arrest. *Journal of Ethnopharmacology*. 146(2): 525-535.
- Artuso, A. (2002). Bioprospecting, benefit sharing and biotechnological capacity building. *World Development*. 30(8): 1355-1368.

- Arun, K.K.V. and Paridhavi, M. (2012). An ethnobotanical phytochemical and pharmacological utilization of widely distributed species *Zanthoxylum*: A comprehensive overview. *International Journal of Pharmaceutical Innovation*. 2(2): 5-7.
- Arunachalam, K. and Parimelazhagan, T. (2014). Evaluation of Phenolic Content, Antioxidant Activity and Nutritional Composition of *Cordia evolution* (Clarke) Gamble. *International Journal of Food Properties*. 17(1): 226-238.
- Arunachalam, K.D., Dowlath, M.J.H., Karuppanan. S.K., Rayan, D.G., Mohamed, S.K. and Subramanian, S. (2020). Effect of solvents on phytochemical composition and antioxidant activity of *Cardiospermum halicacabum* (L.) extracts. *Pharmacognosy Journal*. 12(6): 1241-1251.
- Arya, O.P., Pandey, A. and Sama, P.K. (2016). Ethnobotany and nutritional importance of four selected medicinal plants from Eastern Himalaya, Arunachal Pradesh. *Journal of Medicinal Plants Studies*. 5(1): 45-49.
- Asemave, K. and Ode, S.S. (2018). Proximate and antinutritional analyses of Seed, Pulp and Peel of *Aframomum angustifolium* (Sonn) K. Schum. *Organic and Medical Chemistry*. 7(4): 1-4.
- Asghar, S.F. and Choudahry, M.I. (2011). Gas chromatography-mass spectrometry (GC-MS) analysis of petroleum ether extract (oil) and bio-assays of crude extract of *Iris germanica*. *International Journal of Genetics and Molecular Biology*. 3(7):95-100.
- Asl, M.N. and Hosseinzadeh, H. (2008). Review of pharmacological effects of *Glycyrrhiza* sp. and its bioactive compounds". *Phytotherapy Research*. 22 (6): 709-24.
- Awad, B. and Fink, C.S. (2000). Phytosterols as anticancer dietary components: evidence and mechanism of action. *Journal of Nutrition*. 130(9): 2127-2130.
- Ayam, V.S. and Gogoi, P. (2018). Evaluation of antioxidant activity of the selected frequently used traditional wild medicinal plants from Lakhimpur, Assam (India). *Pleione*. 12(2): 187-192.
- Ayangla, N.W., Singh, N. and Kumar, A. (2016). Phytochemical analysis of plant species of genus *Zanthoxylum*. *International Journal of Medicine and Pharmaceutical Science*. 6(1): 1-8.

- Ayyanar, M. and Ignacimuthub, S. (2011). Ethnobotanical survey of medicinal plants commonly used by Kani tribals in Tirunelveli hills of Western Ghats, India. *Journal of Ethnopharmacology*. 134(2): 851-864.
- Bag, G.C., Devi, P.G. and Bhaigyabati, T. (2015). Assessment of total flavonoid content and antioxidant activity of methanolic rhizome extract of three *Hedychium* species of Manipur Valley. *International Journal of Pharmaceutical Sciences Review and Research*. 30(1): 154-159.
- Bahramsoltani, R., Mohammed, H.F. and Rahimi, R. (2014). Medicinal plants and their natural components as future drugs for the treatment of burn wounds: an integrative review. *Archives of Dermatological Research*. 306 (7): 601-617.
- Bahuguna, A., Khan, I., Bajpai, V.K. and Kang, S.C. (2017). MTT assay to evaluate the cytotoxic potential of a drug. *Bangladesh Journal Pharmacology*. 12(2): 115-118.
- Baker, E.A. (1982). Chemistry and Morphology of Plant Epicuticular Waxes. In: Cutler, D.F., Alvin, K.L. and Price, C.E., Eds., *The Plant Cuticle*, Academic Press, London, 139-166.
- Bakhari, N.A., Othman, C.N., Jusoh, N.A.M., Othman, M.I. and Mohamed, A.S. (2020). Chemical compositions and antiproliferative activity of *Clinacanthus nutans* extract on Human Cervical Cancer Cell lines. *ASM Science Journal*. 13(6): 7-13.
- Bakari, S., Daoud, A., Felhi, S., Smaoui, S., Gharsallah, N. and Kadri, A. (2017). Proximate analysis, mineral composition, phytochemical contents, antioxidant and antimicrobial activities and GC-MS investigation of various solvent extracts of cactus cladode. *Food Science and Technology*. 37(2): 286-293.
- Banerjee, S.K. and Bonde, C. (2011). Total phenolic content and antioxidant activity of extracts of *Bridelia retusa* Spreng Bark: Impact of dielectric constant and geographical location. *Journal of Medicinal Plants Research*. 5(5): 817-822.
- Bangash, J.A., Arif, M., Khan, F., Amin-Ur-Rahman. and Hussain, I. (2011). Proximate composition, minerals and vitamins content of selected vegetables grown in Peshawar. *Journal of the Chemical Society of Pakistan*. 33(1): 118-122.
- Banik, B., Kakoti, S., Saikia, M. and Das, J. (2018). Evaluation of nutritional parameters in medicinal plants of Assam. *Current Trends in Pharmaceutical Research*. 5(1): 34-43.
- Bano, I. and Deora, G.S. (2020). Preliminary Phytochemical Screening and GC-MS analysis for identification of bioactive compounds from *Abutilon fruticosum* Guill

and Perr. A Rare and Endemic plant of Indian Thar Desert. *International Journal of Pharmaceutical Science and Research*. 11(6): 2671-2679.

Barbosa, A.P.de.O., Silverira, G.de.O., Menezes, I.A.C.de., Neto, J.M.R., Bitencurt, J.L.C., Estavam, C.D.S., Lima, A.do.C.B.de., Thomazzi, S.M., Guimaraes, A.G., Jr, .L.J.Q. and Santos, M.R.V.dos. (2013). Antidiabetic effect of the *Chrysobalanus icaco* L. aqueous extract in rats. *Journal of Medicinal Food*. 16(6): 538-543.

Barbouchi, M., Elamrani, K., El Idrissi, M. and Choukrad, M. (2018). A comparative study on phytochemical screening, quantification of phenolic contents and antioxidant properties of different solvent extracts from various parts of *Pistacia lentiscus* L. *Journal of King Saud University-Science*. 32(1): 302-306.

Barman, D., Nath, D., Basumatary, K., Rabha, D., Rao, S., Borah, D. and Boruah, D.C. (2022). Histological and phytochemical analysis of four ethnomedicinal plants used to cure skin diseases. *International Journal of Pharmaceutical Sciences and Research*. 14(1): 416-422.

Baruah, C.C., Bora, M., Saikia, B.N., Hazarika, M. and Barua, I.C. (2015). Quantitative analysis of proximate and mineral composition of a few important medicinal plants of North East Region. *International Journal of Applied Biology and Pharmaceutical Technology*. 6(3): 188-193.

Baruah, H., Sarma, J., Mohan, P., Khargharia, S., Bora, P., Deka, P., Bora, D.P., Hussian, J. and Tamuly, S. (2021). Evaluation of cytotoxicity of *Azadirachta indica* A. Juss., *Curcuma longa* L. and *Ocimum sanctum* L. in BHK-21 cell line. *Annals of Phytomedicine*. 10(2): 182-186.

Basumatary, S. and Narzary, H. (2017). Nutritional Value, phytochemicals and antioxidant property of six wild edible plants consumed by the Bodos of North East India. *Mediterranean Journal of Nutrition and Metabolism*. 10(2): 259-271.

Basumatary, S.K., Ahmed, M. and Deka, S.K. (2004). Some medicinal plant leaves used by Bodo (tribal) people of Goalpara district, Assam. *Natural Product Radiance*. 3(2): 88-90.

Batiha, G.E.S., Beshbishy, A.M., Alkazmi, L., Adeyemi, O.S., Nadwa, E., Rashwan, E., Amany, E.M. and Igarashi, I. (2020). Gas chromatography-mass spectrometry analysis, phytochemical screening and anti-protozoal effects of the methanolic *Viola tricolor* and acetonc *Laurus nobilis* extracts. *BMC Complementary Medicine and Therapies*. 20(87): 1-14.

- Becker, C.D. and Ghimire, K. (2003). Synergy between traditional ecological knowledge and conservation science supports forest preservation in Ecuador. *Conservation Ecology*. 8(1): 1-11.
- Beegum, G.R.J., Beevy, S.S. and Sugunan, V.S. (2014). Nutritive and anti-nutritive properties of *Boerhavia diffusa* L. *Journal of Pharmacognosy and Phytochemistry*. 2(6): 147-151.
- Begum, I.F., Mohankumar, R., Jeevan, M. and Ramani, K. (2016). GCMS analysis of bio-active molecules derived from *Paracoccus pantotrophus* FMR19 and the antimicrobial activity against bacterial pathogens and MDROs. *Indian Journal of Microbiology*. 56(4): 426-432.
- Behbahani, M., Ali, A.M., Muse, R. and Mohd, N.B. (2007). Anti-oxidant and anti-inflammatory activities of leaves of *Barringtonia racemosa*. *Journal of Medicinal Plant Research*. 095-102.
- Bele, A.A., Jadhav, V.M. and Kadam, V.J. (2010). Potential of Tannins: A Review. *Asian Journal of Plant Sciences*. 9(4): 209-214.
- Belewu, M.A., Olatunde, O.A. and Giwa, T.A. (2009). Underutilized medicinal plants and spices: Chemical composition and phytochemical properties. *Journal of Medicinal Plants Research*. 3(12): 1099-1103.
- Benavente-Garcia, O. and Castillo, J. (2008). Update on uses and properties of citrus flavonoids: New findings in anticancer, cardiovascular, and anti-inflammatory activity. *Journal of Agricultural and Food Chemistry*. 56(15): 6185-205.
- Berdeaux, O., Voinot, L., Angioni, E., Juaneda, P. and Sebedio, J.L. (1998). A simple method of preparation of methyl trans-10, cis-12- and cis-9, trans-11-octadecadienoates from methyl linoleate. *Journal of the American Oil Chemists Society*. 75(12): 1749-1755.
- Berdowska, I., Zielinski, B., Fecka, I., Kulbacka, J., Saczko, J. and Gamian, A. (2013). Cytotoxic impact of phenolics from Lamiaceae species on human breast cancer cells. *Food Chemistry*. 141(2): 1313-1321.
- Bhaskaram, P. (2001). Immunobiology of mild nutrient deficiencies. *British Journal of Nutrition*. 85(2): 75-80.
- Bhattacharyya, R., Medhi, K.K. and Borkataki, S. (2019). Phytochemical analysis of *Drymaria cordata* (L.) Willd. Ex Schult. (Whole Plant) used by Tea Tribes of

Erstwhile Nagaon District of Assam, India. *International Journal of Pharmaceutical Sciences and Research*. 10(9): 4264-4269.

Bhatti, M.S., Ali, A., Ahmad, A., Saeed, A. and Malik, S.A. (2015). Antioxidant and phytochemical analysis of *Ranunculus arvensis* L. extracts. *BMC Research Notes*. 8(1): 279-286.

Bhau, B.S., Deka, M., Sarmah, D.K. and Medhi, K. (2014). High gene flow and genetic diversity in three economically important *Zanthoxylum* spp. of Upper Brahmaputra Valley Zone of NE India using molecular markers. *Meta Gene*. 2(C): 706-721.

Bhaumik, U.K., Kumar, D.A., Selvan, V.T., Saha, P., Gupta, M. and Mazumder, U.K. (2008). Antioxidant and free radical scavenging property of methanol extract of *Blumea lanceolaria* leaf in different in vitro models. *Pharmacologyonline*. 2: 74-89.

Bhujbal, S.S., Nanda, R.K., Deoda, R.S., Kumar, D., Kewatkar, S.M., More, L.S. and Patil, M.J. (2010). Structure elucidation of a flavonoid glycoside from the roots of *Clerodendrum serratum* (L.) Moon, Lamiaceae. *Brazilian Journal of Pharmacognosy*. 20(6): 1001-1002.

Bhuyan, M. (2021). A study on traditional medicinal herbs used by the ethnic people of Goalpara district of Assam, North east India. *Scientific Research Journal of India*. 5(1): 18-23.

Biffi, W.L., Moore, E.E., Moore, F.A. et al: (1996). Interleukin-6 delays neutrophil apoptosis via a mechanism involving platelet-activating factor. *Journal of Trauma*. 40(4): 575-78.

Bimakr, M., Rahman, R.A., Taip, F.S., Ganjloo, A., Salleh, L.M., Selamat, J. and Zaidul, I. (2011). Comparison of different extraction methods for the extraction of major bioactive flavonoid compounds from spearmint (*Mentha spicata* L.) leaves. *Food and Bioproducts Processing*. 89(1): 67-72.

Bird, J.K., Calder, P.C. and Eggersdorfer, M. (2018). The role of n-3 long chain polyunsaturated fatty acids in cardiovascular disease prevention, and interactions with statins. *Nutrients*. 10(6): 775. doi: 10.3390/nu10060775.

Birech, Z., Mwangi, P.W., Bukachi, F. and Mandela, K.M. (2017). Application of Raman spectroscopy in type 2 diabetes screening in blood using Leucine and isoleucine amino acids as biomarkers and in comparative anti-diabetic drugs efficacy studies. *Plos ONE*. 12(9): e0185130.

- Bishayee, A., Ahmed, S., Brankov, N. and Perloff, M. (2011). Triterpenoids as potential agents for the chemoprevention and therapy of breast cancer. *Frontiers in Bioscience*. 16(3): 980-986.
- Booth, G.M., Malmstrom, R.D., Kipp, E. and Paul, A. (2012). Cytotoxicity of Selected Medicinal and Nonmedicinal Plant Extracts to Microbial and Cervical Cancer Cells. *Journal of Biomedicine and Biotechnology*.1: 1-4.
- Bora, R., Khakhalary, S. and Dutta, T. (2019). Phytochemical profiling, assessment of total phenolic content, total flavonoid content and antioxidant activity of ethnomedicinal plant, *Meyna Spinosa* from Assam. *Asian Journal of Pharmaceutical and Clinical Research*. 12(11): 1-3.
- Borah, D., Kardong, D., Verma, A.K. and Upadhyaya, S. (2013). Phytochemical and cytotoxic properties of wild *Sarchoclamys pulcherrima* Goud from Assam, North Eastern India. *International Journal of Pharmacy and Pharmaceutical Science*. 3(4): 394-397.
- Borah, D., Mipun, P., Sarma, J., Mili, C. and Narah, D. (2021). Quantitative documentation of traditionally used medicinal plants and their significance to healthcare among the Mishing community of Northeast India. *Ecological Questions*. 32(4): 61-94.
- Borah, P.J. and Sarma, R. (2022). GCMS analysis and qualitative phytochemical screening of *Aristolochia assamica*, a newly discovered rare medicinal plant species of India. *Indian Journal of Natural Product and Resources*. 13(4): 552-558.
- Bordoloi, M., Dutta, D., Narzary, B., Saikia, S., Tamuli, K.J., Lhouvum, N., Nath, S., Sahoo, R.K., Gogoi, B., Saikia, S. and Bhattacharjya, N.K. (2022). Major chemical constituents from *Illicium griffithii* Hook. f. & Thoms of North East India and their cytotoxicity and antimicrobial activities. *Turkish Journal of Chemistry*. 46(5): 1468-1476.
- Borkataky, M., Kakoty, B. B. and Saikia, L.R. (2013). Proximate analysis and antimicrobial activity of *Eclipta Alba* (L.) Hassk.- A Traditionally Used Herb. *International Journal of Pharmacy and Pharmaceutical Sciences*. 5(1): 149-154.
- Bouaziz, A., Khennouf, S., Zarga, M. A., Abdalla, S., Baghiani, A. and Charef, N. (2015). Phytochemical analysis, hypotensive effect and antioxidant properties of *Myrtus communis* L. growing in Algeria. *Asian Pacific Journal of Tropical Biomedicine*. 5(1): 19-28.
- Boudjelal, F., Zitouni, A., Mathieu, F., Lebrihi, A. and Sabaou, N. (2011). Taxonomic study and partial characterization of antimicrobial compounds from a

moderately halophilic strain of the genus *Actinoalloteichus*. *Brazilian Journal of Microbiology*. 42(3): 835-845.

Bouic, P. J. (2001). The role of phytosterols and phytosterols in immune modulation: a review of the past 10 years. *Current Opinion Clinical Nutrition and Metabolic Care*. 4(6): 471-475.

Bowler, P.G., Duerden, B.I. and Armstrong, D.G. (2001). Wound microbiology and associated approaches to wound management. *Clinical Microbiological Reviews*. 14(2): 244-269.

Boyce, P.W. and Christy, L.P. (2004). Applied pharmacology for the veterinary technicians. 2nd ed. St. Luis, USA: WB Saunders Co. 126-127.

Brady, H.J.M. (2004). *Apoptosis Methods and Protocols*. Totowa, NJ, USA: Humana Press; 2004.

Brahma, J., Singh, B., Rethy, P. and Gajurel, P. (2014). Nutritional analysis of some selected wild edible species consumed by the Bodos Tribes of Kokrajhar District, BTC. Assam. *Asian Journal of Pharmaceutical and Clinical Research*. 7(3): 34-37.

Brown, T.J., Brainard, J., Song, F., Wang, X., Abdelhamid, A. and Hooper, L. (2019). Omega-3, omega-6 and total dietary polysaturated fat for prevention and treatment of type 2 diabetes mellitus: Systematic review and meta- analysis of randomized controlled trials. *British Medical Journal*. 366: 14697. doi: 10.1136/bmj.14697.

Budniak, L., Slobodianiuk, L., Marchyshyn, S. and Potishnyi, I. (2022). Determination of amino acids of plants from *Angelica L.* genus by HPLC method. *Pharmacia*. 69(2): 437-446.

Buenz, E. J., Johnson, H. E., Beekman, E. M., Motley, T. J. and Bauer, B. A. (2005). Bioprospecting Rumphius's Ambonese Herbal. *Journal of Ethnopharmacology*. 96(2): 57-70.

Buragohain, J., Konwar, B.K. and Bordoloi, M.J. (2011). Isolation of an antimicrobial compounds from the tender shoots of *Zanthoxylum*. *Der Pharmacia Sinicia*. 2(6): 149-152.

Byers, B.A. (1996). Understanding and influencing behaviors in conservation and natural resources management. African Biodiversity Series, No. 4, Biodiversity Support Program, 109 pp. A USAID-funded consortium of World Wildlife Fund, The Nature Conservancy and World Resources Institute.

- Cahyaningsih, R., Compton, L.J., Rahayu, S., Brehm, J.M. and Maxted, N. (2022). DNA Barcoding Medicinal Plant Species from Indonesia. *Plants*. 11(2): 1375.
- Calixto, J.B. (2005). Twenty five years of research on medicinal plants in Latin America: A personal review. *Journal of Ethnopharmacology*. 100(1-2): 131-134.
- Calder, P.C. (1999). Dietary fatty acids and the immune system. *Lipids*. 34(1): S137-S140.
- Canadanovic-Brunet, J., Cetkovic, G., Dilas, S., Tumbas, V., Bogdanovic, G., Mandic, A., Markov, S., Cvetkovic, D. and Canadanovic, V. (2008). Radical scavenging, Antibacterial and antiproliferative activities of *Melissa officinalis* L. extracts. *Journal of Medicinal Food*. 11(1): 133-143.
- Carmichael, J., Mitchell, J.B., DeGraff, W.G., Gamson, J., Gazdar, A.F., Johnson, B.E., Glatstein, E. and Minna, J.D. (1988). Chemosensitivity testing of human lung cancer cell lines using the MTT assay. *British Journal of Cancer*. 57(6): 540-47.
- Casuga, F.P., Castillo, A.L. and Corpuz, M.J.T. (2016). GC-MS analysis of bioactive compounds presents in different extracts of an endemic plant *Broussonetia luzonica* (Blanco) (Moraceae) leaves. *Asian Pacific Journal of Tropical Biomedicine*. 6(11): 957-961.
- Census of Assam, 2011. Economic Survey of Assam.
- Chakraborty, R., De, B., Devanna, N. and Sen, S. (2012). North East India an Ethnic storehouse of unexplored medicinal plants. *Journal of Natural Product and Plant Resource*. 2(1): 143-152.
- Chand, R.N., Gopalan, R.D. and Christi, K. (2018). Evaluation of antioxidant properties in thirteen Fijian medicinal plants used in Alzheimer's. *Free Radicals and Antioxidants*. 8(1): 11-7.
- Chandra, S. (2018). Fourier transform infrared (FT-IR) spectroscopic analysis of *Nicotiana plumbaginifolia* (Solanaceae). *Journal of Medicinal Plants Studies*. 7(1): 82-85.
- Chandra, S., Saklani, S., Mishra, A.P. and Rana, G. (2014). Nutritional, anti-nutritional profile and phytochemical screening of flowers of *Indigofera tinctoria* from Garhwal Himalaya. *International Journal of Herbal Medicine*. 1(5): 23-27.
- Chandrasekaran, M., Kannathasan, K. and Venkatesalu, V. (2008). Antimicrobial activity of fatty acid methyl esters of some members of Chenopodiaceae. *Zeitschrift Fur Naturforschung C- A Journal of Biosciences*. 63(5): 331-336.

- Chang, J.P., Chang, S., Yang, H., Chen, H., Chien, Y., Yang, B., Su, H. and Su, K. (2020). Omega-3 polyunsaturated fatty acids in cardiovascular diseases comorbid major depressive disorder-results from a randomized controlled trial. *Brain Behaviour and Immunity*. 85: 14-20.
- Chaouche, T.M., Haddouchi, F., Ksouri, R. and Bekkara, F.A. (2014). Evaluation of antioxidant activity of hydromethanolic extracts of some medicinal species from South Algeria. *Journal of the Chinese Medical Association*. 77(6): 302-307.
- Chase, M.W., Cowan, R.S., Hollingsworth, P.M., Berg, C.V.D., Madrinan, S., Petersen, G., Seberg, O., Jorgensen, T., Cameron, K.M., Carine, M., Pedersen, N., Hedderson, A.J.T., Conrad, F., Salazar, G.A., Richardson, J.E., Hollingsworth, M.L., Barraclough, T.G., Kelly, L. and Wilkinson, M. (2007). All proposal for a standardized protocol to barcode all land plants. *New Trends in Plant Systematic*. 56(2): 295-299.
- Chatterjee, S., Karmakar, A. Azmi, S.A. and Barik, A. (2017). Antibacterial activity of long-chain primary alcohols from *Solena amplexicaulis* Leaves. *Proceeding of the Zoological Society*. 71(4): 313-319.
- Chawngkunga, C., Medicinal Plants Used as Healing Agents by the Ethnic People of Mizoram, Aizawl. 2005: pp. 3-4.
- Chayee, C.C., Mau, J.L. and Wu, C.M (1996). Characteristics of the steam distilled oil and carbondioxide extract of *Zanthoxylum simulans* fruits. *Journal of Agriculture and Food Chemistry*. 44(4): 1096-1099.
- Chen, L., Wang, Y., Xu, Q. and Chen, S. (2018). Omega-3 fatty acids as a treatment for non-alcoholic fatty liver disease in children: A systematic review and meta-analysis of randomized controlled trials. *Clinical Nutrition*. 37(2): 516-521.
- Chen, M., Jin, H., Zhang, W., Yan, S. and Shen, Y. (2009). Chemical constituents of plants from the genus *Blumea*. *Chemistry and Biodiversity*. 6(6): 809-817.
- Chen, S., Yao, H., Han, J., Liu, C., Song, J., Shi, L., Zhu, Y., Ma, X., Gao, T., Pang, X., Luo, K., Li, Y., Jia, X., Lin, Y. and Leon, C. (2010). Validation of the ITS2 Region as a Novel DNA Barcode for Identifying Medicinal Plant Species. *PLoS ONE*. 5(1): e8613.
- Chen, W.P., Yang, X.Y., Hegeman, A.D., Gray, W.M. and Cohen, J.D. (2010). Microscale analysis of amino acids using gas chromatography-mass spectrometry after methyl chloroformate derivatization. *Journal of chromatography B, Analytical Technologies in the Biomedical and Life*. 878 (24): 2199-2208.

- Chen, Y., Chen, H., Huang, B., Chen, Y. and Chang, C. (2019). Polyphenol rich extracts from *Toona sinensis* bark and fruit ameliorate free fatty acid-induced lipogenesis through AMPK and LC3 pathways. *Journal of Clinical Medicine*. 8(10): 1664. doi: 10.3390/jcm8101664.
- Cheng, X., Su, X., Chen, X., Zhao, H., Bo, C., Xu, J., Bai, H. and Ning, K. (2014). Biological ingredient analysis of traditional Chinese medicine preparation based on high-throughput sequencing: the story for Liuwei Dihuang Wan. *Scientific reports*. 4(1): 1-12.
- Chetia, J. (2021). Nutritive value of some medicinally important species of Lamiaceae from Dibrugarh, Assam, India. *Ecology, Environment and Conservation*. 27(1): S168-S175.
- Chibueze, I. and Peters, D.E. (2022). Nutrient's composition of *Costus lucanusianus* stem. *GSC Biological and Pharmaceutical Science*. 18(02): 070-081.
- Chiou, S.J., Yen, J.H., Fang, C.L., Chen, H.L. and Lin, T.Y. (2007). Authentication of medicinal herbs using PCR-amplified ITS2 with specific primers. *Planta Medica*. 73(13):1421-1426.
- Cho, S.K., Ngoc, B.T., Yen, T.K.N., Moon, J.Y. and Ediriweera, M.K. (2020). Pentadecanoic Acid, an Odd-Chain Fatty Acid, Suppresses the Stemness of MCF-7/SC Human Breast Cancer Stem-Like Cells through JAK2/STAT3 Signaling. *Nutrients*. 12(6): 1-20.
- Choi, C.W., Kim, S.C., Hwang, S.S., Choi, B.K., Ahn, H.J., Lee, M.Y., Park, S.H. and Kim, S.K. (2002). Antioxidant activity and free radical scavenging capacity between Korean medicinal plants and flavonoids by assay-guided comparison. *Plant Science*. 163(6): 1161-1168.
- Choudhury, S., Bahadur, B. and Pullaiah, T. (2017). Ethnomedicinal plants of North-East India. *Ethnobotany of India*. 3(1): 1-70.
- Choudhury, S., Sharma, P., Choudhury, M.D. and Sharma, G.D. (2012). Ethmoedicinal plants used by Chorei tribes of Southern Assam, North Eastern India. *Asian Pacific Journal of Tropical Diseases*. 2(1): 141-147.
- Chujo, H., Yamasaki, M., Nou, S., Koyanagi, N., Tachibana, H. and Yamada, K. (2003). Effect of conjugated linoleic acid isomers on growth factor induced proliferation of human breast cancer cells. *Cancer Letters*. 202(1): 81-87.
- Chung, K., Wong, T., Wei, C., Huang, Y. and Lin, Y. (1998). Tannins and Human Health: A Review. *Critical Review in Food Science and Nutrition*. 38(6): 421-464.

- Ciorita, A., Zagrean-Tuza, C., Mot, A.C., Carpa, R., and Parvu, M. (2021). The Phytochemical Analysis of Vinca L. Species Leaf Extracts Is Correlated with the Antioxidant, Antibacterial and Antitumor Effects. *Molecules*. 26 (10): 1-21.
- Cl, K.C. and Indira, G. (2016). Quantitative estimation of total phenolic, flavonoids, tannin and chlorophyll content of leaves of *Strobilanthes kunthiana* (Neelakurinji). *Journal of Medicinal Plants Studies*. 4(60): 282-286.
- Coghlan, M. L., Haile, J., Houston, J., Murray, D. C., White, N. E., Moolhuijzen, P., Bellgard, M.I. and Bunce, M. (2012). Deep sequencing of plant and animal DNA contained within traditional Chinese medicines reveals legality issues and health safety concerns. *PLoS genetics*. 8(4): e1002657.
- Coleman, A.W. (2007). Pan-eukaryote ITS2 homologies revealed by RNA secondary structure. *Nucleic Acids Research*. 35(10): 3322-3329.
- Comerford, K.B. and Pasin, G, (2016). Emerging evidence for the importance of dietary protein source on glucoregulatory markers and type 2 diabetes: different effects of dairy, meat, fish, egg, and plant protein foods. *Nutrients*. 8(8): e446. <https://doi.org/10.3390/nu8080446>.
- Cordeiro, de. C.B.M.P., Santos, de.L.N.D., Ferreira, M.R.A., Araujo, de.L.C.C., Junior, A.R.C., Santos, da.C.A.D., Oliveira, de.A.P., Silva, da.A.G., Silva, da F.E.P., Correia, dos.S.M.T., Almeida, da.S.J.R.G., Silva, da.L.C.N., Soares, L.A.L., Napoleao1, T.H., Silva, da.M.V. and Paiva, P.M.G. (2018). Hexane extract from *Spondias tuberosa* (Anacardiaceae) leaves has antioxidant activity and is an anti-Candida agent by causing mitochondrial and lysosomal damages. *BMC Complementary and Alternative Medicine*. 18(1):284. doi: 10.1186/s12906-018-2350-2.
- Cox, P.A. (2000). Will tribal knowledge survive the millennium Science? *Science*. 287 (5450): 44-45.
- Cragg, G.M. and Newman, D.J. (2005). Plants as a source of anti-cancer agents. *Journal of Ethnopharmacology*. 100(1-2): 72-9.
- Cumbane, P. and Munyemana, F. (2017). Antioxidant and antibacterial activity of leaves and stem extracts of *Bridelia cathartica* Bertol. *Sky Journal of Microbiology Research*. 5(2): 18-26.
- Das, A.J., Mohammad, A., Kumarm M. and Kumar, R. (2013). FTIR analysis for screening variations in antimicrobial activity of fresh and dried leaf extract of

Rhynchostylis retusa: A threatened orchid species of Assam, North East India. *International Research Journal of Pharmacy*. 4(7): 187-189.

Das, A.K., Dutta, B.K. and Sharma, G.D. (2008). Medicinal plants used by different tribes of Cachar district, Assam. *Indian Journal of Traditional Knowledge*. 7(3): 446-454.

Das, M.K., Singh, N. and Rajamani, P. (2023). Comprehensive Assessment of the Antioxidant and Anticancer Potential of Selected Ethnobotanical Plants. *Oxygen*. 3(2): 203-221.

Das, R. (2016). Biodiversity of ethnomedicinal plants used by the ethnic tribal people of Barpeta District of Assam, North East India. *Asian Journal of Pharmaceutical Science and Technology*. 6 (1): 27-32.

Das, S., Kandali, R. and Baishya, S. (2023). Proximate composition of some indigenous plant species having ethno-medicinal importance. *Journal of Agrisearch*. 10(1): 41-45.

Dasgupta, N. and De, B. (2007). Antioxidant activity of some leafy vegetables of India: A comparative study. *Food Chemistry*. 101(2): 471-474.

De Boer, H.J., Ghorbani, A., Manzanilla, V., Raclariu, A. C., Kreziou, A., Ounjai, S., Osathanunkul, M. and Gravendeel, B. (2017). DNA metabarcoding of orchid-derived products reveals widespread illegal orchid trade. *Proceedings of the Royal Society B: Biological Sciences*. 284(1863): 20171182.

Deka, J. and Kalita, J.C. (2013). Ethnobotanically important medicinal plants of Kamrup District, Assam, India, used in fertility treatment. *International Research Journal of Pharmacy*. 4(3): 229-232.

Deka, K. and Nath, N. (2014). Indigenous medicinal plant knowledge of cough or bronchial problems in Goalpara district (N.E. India). *International Journal of Pure and Applied Bioscience*. 2(5): 80-87.

Demirezen, D. and Ahmet, A. (2006). Heavy metal levels in vegetables in turkey are within safe limits for Cu, Zn, Ni and exceeded for Cd and Pb. *Journal of Food Quality*. 29(3): 252-265.

Despande, V.H. and Shastri, R.K. (1977). Chemical investigation of 3 *Zanthoxylum* species: *Zanthoxylum alatum*, *Zanthoxylum oxyphyllum* and *Zanthoxylum acanthopodium*. *Indian Journal of Chemistry*. 15(1): 95-96.

- Dhanani, T., Shah, S., Gajbhiye, N.A. and Kumar, S. (2017). Effect of extraction methods on yield, phytochemical constituents and antioxidant activity of *Withania somnifera*. *Arabian Journal of Chemistry*. 10(1): S1193-9.
- Dhivya, S.M. and Kalaichelvi, K. (2017). UV-Vis Spectroscopic and FTIR analysis of *Sarcostemma brevistigma*, Wight. And Arn. *International Journal of Current Pharmaceutical Research*. 9(3): 46-49.
- Dietary Reference Intakes (DRIs) (2004). Recommended Intake for individuals, Elements. Food and Nutrition Board, Institute of Medicine, National Academies.
- Dinesh-Kumar, G. and Rajakumar, R. (2018). GC-MS analysis of bioactive compounds from ethanolic leaves extract of *Eichhornia crassipes* (Mart) Solms. and their pharmacological activities. *Journal of Pharmaceutical Innovation*. 7(8): 459-462.
- Duan, X.J., Zhang, W.W., Li, X.M. and Wang, B.G. (2006). Evaluation of antioxidant property of extract obtained from a red alga. *Polysiphonia ureceolta*. *Food Chemistry*. 95(1): 37-43.
- Dubey, S.K., Gairola, K., Gururani, S., Kumar, R., Prakash, O., Agrawal, S. and Kumar, S. (2022). Composition, Antioxidant and Anti-inflammatory activities of hexane and methanol extracts of *Acmella uliginosa* from Terai region of Uttarakhand. *Brazilian Journal of Pharmaceutical Sciences*. 58: 1-14.
- Dung, N.X., Loi, D.T., Huang, D.T. and Leclercq, P.A. (1991). Chemical Composition of the Oil of *Blumea lanceolaria* (Roxb.) Druce from Vietnam. *Journal of Essential Oil Research*. 3(4): 285-286.
- Dwivedi, S.K. and Dey, S. (2002). Medicinal herbs: A potential source of toxic metal exposure for man and animals in India. *Archives of Environmental Health*. 57(3): 229-31.
- Eberhardt, T.L., Li, X., Shupe, T.F. and Chung, Y.H. (2007). Chinese tallow tree (*Sapium sebiferum*) utilization: Characterization of extractives and cell-wall chemistry. *Wood Fiber Science*. 39(2): 319-324.
- Ebrahimzadeh, M.A., Nabavi, S.F., Nabavi, S.M. and Eslami, B. (2010). Antihypoxic and antioxidant activity of *Hibiscus esculentus* seeds. *Grasas y Aceites*. 61(1): 30- 36.
- Ebrahimzadeh, M.A., Nabavi, S.M., Nabavi, S.F. and Eslami, B. (2010). Antioxidant activity of *Ornithogalum sintenisii* bulbs and aerial part at flowering stage. *Tropical Journal of Pharmaceutical Research*. 9(2): 141-148.

- Edeoga, H.O., Okwu, D.E. and Mbaebie, B.O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*. 4 (7): 685-688.
- Ejoh, A.R., Nkonga, D.V., Innocent, G. and Mbofung, C.M. (2007). Nutritional component of some Non-conventional leafy vegetables consumed in Cameroon. *Pakistan Journal of Nutrition*. 6(6): 712-717.
- Ekor, M. (2014). The growing use of herbal medicines: Issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Neurology*. 4(177): 1-10.
- El, Cadi H, El Cadi A, Kounoun A, Oulad El Majdoub Y, Lovillo MP, Brigui J, Dugo P, Mondello L, Cacciola F. (2020). Wild strawberry (*Arbutus unedo*): phytochemical screening and antioxidant properties of fruits collected in northern Morocco. *Arabian Journal of Chemistry*. 13(8): 6299-6311.
- El-Atroush, H., Magdy, M. and Werner, O. (2015). DNA Barcoding of two endangered medicinal Plants from Abou Galoom protectorate. *Life Science Journal*. 12(9): 101-109.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*. 35(4): 495-516.
- Elufioye, T., Bamgbose, M. and Alabi, S. (2016). Evaluation of antioxidant and antiacne activity of *Psorospermum febrifugum* (Spach) and *Psorospermum corymbiferum* (Hochr.). *Journal of Pharmaceutical Research International*. 12(5): 1-10.
- ElZanaty, S.A., Seif Eldein, N.A., ElGebaly, E.A., ElGhaly, E.M. and ElGizawy, H.A. (2022). Chemical composition and antimicrobial activity of the essential oils of *Thevetia peruviana* and *Plumeria rubra* cultivated in Egypt. *Azhar International Journal of Pharmaceutical and Medical Sciences*. 2(2): 34-42.
- Emebu, P.K. and Anyika, J.U. (2011). Proximate and Mineral composition of Kale (*Brassica oleracea*) grown in Delta State, Nigeria. *Pakistan Journal of Nutrition*. 10(2): 190-194.
- Emmanuel, T.V., Njoka, J.T., Catherine, L.W. and Lyaruu, H.V.M. (2011). Nutritive and anti-nutritive qualities of mostly preferred edible woody plants in selected drylands of Iringa District, Tanzania. *Pakistan Journal of Nutrition*. 10(8): 786-791.

- Engtipi, M. and Raju, N.L. (2022). Nutritional Value and Anti-nutritional Factor's Analyses of Five Indigenous Spices used by the Karbi Group of Assam, India. *Agricultural Science Digest*. 42(2): 242-245.
- Epand, R., Savage, P. and Epand, R. 2007. Bacterial lipid composition and the antimicrobial efficacy of cationic steroid compounds (Ceragenins). *Biochimica et Biophysica Acta (BBA)-Biomembrane*. 1768(10): 2500-2509.
- Evans, W.C. 2009. Trease and Evans Pharmacognosy. London: Saunders Limited.
- FAO, (2003). Food energy methods of analysis and conversion factors. Food and nutrition paper 77, Food and Agriculture Organization of the United Nations, Rome, Italy. 1-93.
- FAO/WHO/UNO. (1985). Protein and Energy Requirements. In FAO/WHO/UNO1985.
- Farahmandfar, R., Kenari, R.E., Asnaashari, M., Shahrampour, D. and Bakhshandeh, T. (2019). Bioactive compounds, antioxidant and antimicrobial activities of *Arum maculatum* leaves extracts as affected by various solvents and extraction methods. *Food Science and Nutrition*. 7(2): 465-75.
- Fazekas, A.J., Kuzmina, M.L., Newmaster, S.G. and Hollingsworth, P.M. (2012). DNA Barcoding Methods for Land Plants. *Methods in Molecular Biology*. 858(1): 223-252.
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39(4): 783-791.
- Firoozi, S., Jamzad, M. and Yari, M. (2016). Biologically synthesized silver nanoparticles by aqueous extract of *Satureja intermedia* C.A. Mey and the evaluation of total phenolic and flavonoid contents and antioxidant activity. *Journal of Nanostructure in Chemistry*. 6(4): 357-364.
- Fisher, D.E. (1994). Apoptosis in cancer therapy: Crossing the threshold. *Cell*. 78(4): 539-542.
- Flaster, T. (1996). Ethnobotanical approaches to the discovery of bioactive compounds. "Progress in new crops". Alexandria, VA (USA): ASHS Press, ISBN 09-615-02738.
- Food and Agriculture Organization (FAO) (1980). Basic Texts of the Food and Agriculture Organization of the United Nations, Vols. I and II. FAO, Rome. 242p.

- Furlan, C.P.B., Marques, A.C., Marineli, R.da.S., Marostica, M. R. and Junior. (2013). Conjugated linoleic acid and phytosterols counteract obesity induced by high-fat diet. *Food Research International*. 51(2): 429-35.
- Gairola, S., Sharma, J. and Bede, Y.S. (2014). A cross-cultural analysis of Jammu, Kashmir and Ladakh (India) medicinal plant use. *Journal of Ethnopharmacology*. 155(2): 925-986.
- Gallo, M.B.C. and Sarachine, M.J. (2009). Biological Activities of Lupeol. *International Journal of Biomedical and Pharmaceutical Science*. 3(1): 46-66.
- Ganesh, M. and Mohankumar, M. (2017). Extraction and identification of bioactive components in *Sida cordata* (Burm. f.) using gas chromatography-mass spectrometry. *Journal of Food Science and Technology*. 54(10): 3082-3091.
- Garaniya, N. and Bapodra, A. (2014). Ethnobotanical and phytopharmacological potential of *Abrus precatorius* L.: A review. *Asian Pacific Journal of Tropical Biomedicine*. 4(1): 27-34.
- Gaur, R.D. and Sharma, J. (2011). Indigenous knowledge on the utilization of medicinal plant diversity in the siwalik region of Garhwal Himalaya, Uttarakhand. *Journal of Science*. 27 (1): 23-31.
- Gawali, P. and Jadhav, B.L. (2011). Antioxidant activity and antioxidant phytochemical analysis of mangrove species *Sonneratia alba* and *Bruguiera cylindrica*. *Asian Journal Microbiology, Biotechnology & Environmental Sciences*. 13(2): 257-261.
- Geetha, D.H., Jayashree, I. and Rajeswari, M. (2015). GC-MS analysis of ethanolic extract of *Elaeocarpus serratus* L. *European Journal of Pharmaceutical and Medical Research*. 2(2): 296-302.
- Gemedo, H.F. and Ratta, N. (2014). Antinutritional factors in plant foods: potential health benefits and adverse effects. *International Journal of Nutrition and Food Sciences*. 3(4): 284-9.
- George, R.O., Radha, H.R. and Somasekariah, B.V. (2018). In-vitro antidiabetic activity and GCMS analysis of bioactive compounds present in the methanol extract of *Kalanchoe pinnata*. *Indian Journal of Chemistry*. 57B: 1213-1221.
- Ghuman, S., Ncube, B., Finnie, J.F., McGaw, L.J., Mfotje Njoya, E., Coopoosamy, R.M. and Staden, J.V. (2019). Antioxidant, anti-inflammatory and wound healing properties of medicinal plant extracts used to treat wounds and dermatological disorders. *South African Journal of Botany*. 126: 232-240.

- Gnansounou, S.M., Noudogbessi, J.P., Yehouenou, B. and Sohounhloue, D. (2014). Proximate composition and micronutrient potentials of *Dialium guineense* wild growing in Benin. *International Food Research Journal*. 21(4): 1603-1607.
- Gogoi, P. and Sen, S. (2023). Ethnobotanical study of medicinal plants used traditionally for managing cuts and wounds by the rural people of Kailashpur, Assam, India. *Indian Journal of Pharmaceutical Education and Research*. 57(3): 678-689.
- Gohel, A., McCarthy, M.B. and Gronowicz, G. (1999). Estrogen prevents glucocorticoid-induced apoptosis in osteoblasts *in vivo* and *in vitro*. *Endocrinology*. 140(11): 5339-47.
- Gong, P., Li, Y., Yao, C., Guo, H., Hwang, H., Liu, X., Xu, Y. and Wang, X. (2017). Traditional Chinese medicine on the treatment of coronary heart disease in recent 20 years. *The Journal of Alternative and Complementary Medicine*. 23(9): 659-666.
- Gonzalez, M.A., Baraloto, C., Engel, J., Mori, S.A., Petronelli, P., Riera, B., Roger, A., Thébaud, C. and Chave, J. (2009). Identification of Amazonian Trees with DNA barcodes. *PLoS ONE*. 4(10): 1-7.
- Gorinstein, S., Oscar, J., Vargas, M., Jaramillo, N.O., Salas, I.A., Ayala, A.L.M., Arancibia-Avila, P., Toledo, F., Katrich, E. and Trakhtenberg, S. (2007). The total polyphenols and the antioxidant potentials of some selected cereals and pseudocereals. *European Food Research and Technology*. 225(3-4): 321-328.
- Goulas, V., Gomez- Caravaca, A.M., Exarchou, V., Gerothanassis, I.P., Segura-Carretero, A. and Gutierrez, A.F. (2012). Exploring the antioxidant potential of *Teucrium polium* extracts by HPLC-SPE-NMR and on-line radical-scavenging activity detection. *LWT- Food Science and Technology*. 46(1): 104-109.
- Gowri, J., Sahayaraja, A.P., Dharmalingama, V., Shobanaa, R. and Prema, A.P. (2015) Phytochemical screening by FTIR spectroscopic analysis of leaf and stem Extracts of *wedelia biflora*. *International Journal of Nano Corrosion Science and Engineering*. 2(5): 322-334.
- Greenwell, M. and Rahman, P.K.S.M. (2015). Medicinal Plants: Their Use in Anticancer Treatment. *International Journal of Pharmaceutical Sciences and Research*. 6(10): 4103-12.
- Griffiths, P.R. and de Haseth, J.A. (1986). Fourier transform infrared spectroscopy. New York: Wiley. 90(12): 1240-41.

- Guha, G., Rajkumar, V., Mathew, L. and Kumar, R.A. (2011). The antioxidant and DNA protection potential of Indian tribal medicinal plants. *Turkish Journal of Biology*. 35(2): 233-242.
- Gupta, A.K., Tandon, N. and Sharma, M. (2008). Review on Indian Medicinal Plants. Vol 7, Indian Council of Medical Research, New Delhi. 110-111.
- Hadi, M.Y., Mohammed, G. J. and Hameed, I. H. (2016). Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognoy and Phytotherapy*. 8(2): 8-24.
- Haghighi, S.R., Asadi, M.H., Akrami, H. and Baghizadeh, A. (2017). Anti-carcinogenic and anti-angiogenic properties of the extracts of *Acorus calamus* on gastric cancer cells. *Avicenna Journal of Phytomedicine*. 7 (2): 145-156.
- Hajra, P.K., Verma, D.M. and Giri, G.S. (1996). Materials for the flora of Arunachal Pradesh. Botanical Survey of India, Kolkata, India, Vol.1. 270-271.
- Hameed, I.H., Hussein, H.J., Kareem, M.A. and Hamad, N.S. (2015). Dentification of five newly described bioactive chemical compounds in methanolic extract of *Mentha viridis* by using gas chromatography - mass spectrometry (GC-MS). *Journal of Pharmacognosy and Phytotherapy*. 7(7): 107-125.
- Hamid, A. A., Zubair, M.F., Olajide, F.O., Ibrahim, S.O., Shehu, A., Bale, M.I., Muhammed, S.O., Aina, S. D. and Jimoh, A. (2018). Chemical compositions from the leaf extracts of *Funtumia Africana* (Benth.) stapf with its antioxidant and anti-inflammatory activity. *Nigerian Journal of Chemical Research*. 23(2): 83-96.
- Harborne, J.B. 1998. Phytochemical methods: A guide to modern techniques of plant analysis. London: Chapman & Hall.
- Harshberger, J.W. (1896). The purposes of ethnobotany. *Botanical Gazette*. 21(3): 146-154.
- Hasegawa, S., Ichiyama, T., Sonaka, I., Ohsaki, A., Okada, S., Wakiguchi, H., Kudo, K., Kittaka, S., Hara, M. and Furukuwa, S. (2012). Cysteine, histidine and glycine exhibit anti-inflammatory effects in human coronary arterial endothelial cells. *Clinical and Experimental Immunology*. 167(2): 269-74.
- Haub, M.D. and Lattimer, J.M. (2010). Effects of dietary fiber and its components on metabolic health. *Nutrients*. 2(12): 1266-1289.
- Hebert, P.D.N., Cywinska, A., Ball, S.L. and deWaard, J.R. (2003). Biological identifications through DNA barcodes. Proceeding of Royal Society B: *Biological Sciences*. 270(1512): 313-321.

- Hema, R., Kumaravel, S., Gomathi, S. and Sivasubramaniam, C. (2010). Gas chromatography-mass spectroscopic analysis of Lawsonia inermis leaves. *Life Science Journal*. 7(4): 48-50.
- Henry, G.E., Momin, R.A., Nair, M.G. and Dewitt, D.L. (2002). Antioxidant and cyclooxygenase activities of fatty acids found in food. *Journal of Agricultural and Food Chemistry*. 50(2002): 2231-2234.
- Herrero, M., Castro-Puyana, M., Mendiola, J.A. and Ibanez, E. (2013). Compressed fluids for the extraction of bioactive compounds. *Trends in Analytical Chemistry*. 43: 67-83.
- Hodges, J. (2005). Cheap food and feeding the world sustainably. *Livestock Production Science*. 92(1): 1-16.
- Holecek, M. (2017). Branched-chain amino acid supplementation in treatment of liver cirrhosis: Updated views on tips how attenuate their harmful effects on cataplerosis and ammonia formation. *Nutrition*. 41: 80-5.
- Hollingsworth, P.M., Forrest, L.L., Spouge, J.L., Hajibabaei, M., Ratnasingham, S., Vander Bank, M., Chase, M.W., Cowan, R.S., Erickson, D.L., Fazekas, A.J., Graham, S.W., James, K.E., Kim, K.J., Kress, W.J., Scheider, H., Van AlphenStahl, J., Barrett, S.C.H., Van den Berg, C. and Little, D.P. (2009). A DNA barcode for land plants. *Proceeding of National Academy of Sciences*. 106(31): 12794-12797.
- Hollingsworth, P.M., Graham, S.W. and Little, D.P. (2011) Choosing and using a plant DNA barcode. *PLoS ONE*. 6(5): e19254. <https://doi.org/10.1371/journal.pone.0019254>.
- Hooker, J.D. (1875). The Flora of British India. L. Reeve & Co, London. 492-496.
- Hossain, M. A., Weli, A.M. and Ahmed, S.H.I. (2019). Comparison of total phenols, flavonoids and antioxidant activity of various crude extracts of Hyoscyamus galleri traditionally used for the treatment of epilepsy. *Clinical Phytoscience*. 9(6): 1-8.
- Ilaiyaraja, N. and Khanum, F. (2011). Antioxidant potential of Tinospora cordifolia extracts and their protective effects on oxidation of biomolecules. *Pharmacognosy Journal*. 3(20): 56-61.
- Ilelaboye, N.O.A., Amoo, I.A. and Pikuda, O.O. (2013). Effect of cooking methods on mineral and anti nutrient composition of some green leafy vegetables. *Archives of Applied Science Research*. 5(3): 254-60.

- Indhumathi, M. and Arunprasath, A. (2019) Identification of functional groups in *Corbichonia decumbens* by Fourier-Transform Infrared Spectroscopy. *Journal of Drug Delivery and Therapeutics*. 9(3): 583-587.
- Insel, P.M., Ross, D., McMahon, K. and Bernstein, M. (2010). Nutrition. (4th edition) Jones and Bartlett Publishers, Sudbury, MA. pp. 1002.
- Institute of Medicine, Food and Nutrition Board. (2002). Dietary Reference Intakes: energy, carbohydrates, fiber, fat, fatty acids, cholesterol, protein and amino acids. Washington: National Academics Press.
- Inuwa, H.M., Aina, V.O., Aimola, B.G.I. and Amao, T. (2011). Comparative Determination of Antinutrient factors in Groundnut Oil, Palm Oil. *Advance Journal of Food Science and Technology*. 3(4): 275-279.
- Iqbal, E., Salim, K.A. and Lim, L.B.L. (2015). Phytochemical screening, total phenolics and antioxidant activities of bark and leaf extracts of *Goniothalamus velutinus* (Airy Shaw) from Brunei Darussalam. *Journal of King Saud University Science*. 27(3): 224-32.
- Isah, A., Abdullahi, M. and Tsado, M.J. (2015). Evaluation of Phytochemical, Anti-nutritional and Antioxidant Potentials of Flower and Seed Methanol Extracts of *Senna alata* L. Grown in Nigeria. *American Journal of Applied Chemistry*. 3(3): 93-100.
- Islam, M.Z., Hoque, M.M., Asif-Ul-Alam, S.M. and Monalisa, K. (2015). Chemical composition, Antioxidant capacities and storage stability of Citrus macroptera and *Garcinia pedunculata* fruits. *Emirates Journal of Food and Agriculture*. 27(3): 275-282.
- Islary, A., Sarmah, J. and Basumatary, S. (2016). Proximate composition, mineral content, phytochemical analysis and in vitro antioxidant activities of a wild edible fruit (*Grewia sapida* Roxb. ex DC.) found in Assam of North-East India. *Journal of Investigational Biochemistry*. 5(1): 21-31.
- Islary, A., Sarmah, J. and Basumatary, S. (2017). Nutritional value, phytochemicals and antioxidant properties of two wild edible fruits (*Eugenia operculata* Roxb. and *Antidesma bunius* (L.) from Assam, North-East India. *Mediterranean Journal of Nutrition and Metabolism*. 10(1): 29-40.
- Islary, A., Sarmah, J. and Basumatary, S. (2018). Antibacterial property and anti-nutritional contents of five wild fruits of Assam, India. *Journal of Applied Pharmaceutical Science*. 8(12): 170-175.

- Jaddoa, H.H., Hameed, I.H. and Mohammed, G.J. (2016). Analysis of volatile metabolites released by *Staphylococcus aureus* using gas chromatography-Mass spectrometry and determination of its antifungal activity. *Oriental Journal of Chemistry*. 32(4): 8-24.
- Jaimes-Morales, J., Marrugo-Ligardo, Y.A. and Acevedo-Correa, D. (2022). Analysis of Mesquite (*Prosopis juliflora*) Protein Concentrate for Possible Use as Supplementary Protein. *International Journal of Food Science*. 1-7. <https://doi.org/10.1155/2022/7621818>.
- Jain, J.B., Kumane, S.B. and Bhattacharya, S. (2016). Medicinal flora of Madhya Pradesh and Chattishgarh-A Review. *Indian Journal of Traditional Knowledge*. 5(2): 237-42.
- Jamuna, K.S., Ramesh, C.K., Srinivasa, T.R. and Raghu, K.I. (2011). In vitro antioxidant studies in some common fruits. *International Journal of Pharmacy and Pharmaceutical Science*. 3(1): 60-63.
- Jananie, R.K., Priya, V. and Vijayalakshmi K. (2011). Determination of Bioactive Components of *Cynodon dactylon* by GC-MS Analysis. *New York Science Journal*. 4(4): 16-20.
- Javed, M.R., Salman, M., Tariq, A., Tawab, A., Zahoor, M.K., Naheed, S., Shahid, M., Ijaz, A. and Ali, H. (2022). The Antibacterial and Larvicidal Potential of Bis-(2-Ethylhexyl) Phthalate from *Lactiplantibacillus plantarum*. *Molecules*. 27(21): 1-15.
- Jegadeeswari, P., Nishanthini, A., Muthukumarasamy, S. and Mohan, V.R. (2012). GC-MS analysis of bioactive components of *Aristolochia krysagathra* (aristolochiaceae) *Journal of Current Chemical and Pharmaceutical Sciences*. 2(4): 226-232.
- Jha, R.R. and Verma, S.K. (1996). Ethnobotany of Sauria Paharias of Santhal Pargana, Bihar, Indian Medicinal plants. *Ethnobotany*. 8(1): 31-35.
- Jiang, G., Xiao, X., Zeng, Y., Nagabhushanam, K., Majeed, M. and Xiao, D. (2013). Targeting beta-catenin signaling to induce apoptosis in human breast cancer cells by z-guggulsterone and Gugulipid extract of Ayurvedic medicine plant *Commiphora mukul*. *BMC Complement. The Journal of Alternative and Complementary Medicine*. 13(1): 1-12.
- Jima, T.T. and Megersa, M. (2018). Ethnobotanical study of medicinal plants used to treat human diseases in Berbere District, Bale Zone of Oromia Regional State, South East Ethiopia. *Evidence-Based Complementary and Alternative Medicine*. 1-16. doi: 10.1155/2018/8602945.

- Jimenez-Sepulveda, G., Aquino-Reyna, C., Martinez- Chaires, L., Torres-Bermudez, K. and Monroy-Rodriguez, M. (2009). Antioxidant activity and content of phenolic compounds and flavonoids from *Justicia spicigera*. *Journal of Biological Sciences*. 9(6): 629-632.
- Jisha, M., Zeinul Hukuman, N.H. and Leena, P. (2016). GC- MS analysis of leaves and flowers of *Pogostemon quadrifolius* (benth.) F. muell. (Lamiaceae). *World Journal of Pharmaceutical Research*. 5(12): 667-681.
- Junwei, L., Juntao, C., Changyu, N. and Peng, W. (2018). Molecules and functions of rosewood: *Pterocarpus cambodianus*. *Arabian Journal of Chemistry*. 11(6): 763-770.
- Just, M.J., Giner, R.M., Recio, C.M., Cuellar, M.J., Manez, S., Bilia, A. R. and Rios, J. L. (1998). Anti-inflammatory activity of unusual Lupane Saponins from *Bupleurum frutescens*. *Planta Medica*. 64(5): 404-407.
- Kala, C.P., Nehal, A. and Farooquee, N.A. (2005). Traditional uses and conservation of Timur (*Zanthoxylum armatum* DC) through social institutions in Uttarakhand Himalaya. *India Conservation and Society*. 3(1): 224-230.
- Kalakandan, S.K., Pakkirisamy, M. and Ravichandran, K. (2017). Phytochemical Screening, GC-MS, FT-IR Analysis of Methanolic Extract of *Curcuma caesia* Roxb (Black Turmeric). *Pharmacognosy Journal*. 9(6): 952-956.
- Kalia, A.N., Dhar, K.L., Singh, M.P. and Mukhija, M. (2015). Cytotoxic and antioxidant activity of *Zanthoxylum alatum* stem bark and its flavonoid constituents. *Journal of Pharmacognosy and Phytochemistry*. 4(4): 86-92.
- Kalidass, C. and Mohan, V.R. (2011). Nutritional and antinutritional composition of itching bean (*Mucuna pruriens* (L.) dc var. *pruriens*): an underutilized tribal pulse in Western Ghats, Tamil Nadu. *Tropical and Subtropical Agroecosystems*. 14(2): 279-283.
- Kalita, D. and Phukan, B. (2010). Some ethnomedicines used by the Tai Ahom of Dibrugarh district, Assam, India. *Indian Journal of Natural Products and Resources*. 1(4): 507-511.
- Kaliwal, B.B., Chinchali, J.F. and Sanakal, R.D. (2011). Evaluation of anticarcinogenic activity of *Clerodendrum serratum* leaf extract on liver and kidney of 7, 12 dimethylbenz [a] anthracene (DMBA) induced skin carcinogenesis in mice. *European Journal of Experimental Biology*. 1(4): 130-41.

- Kalustian, P. (1985). Pharmaceutical and cosmetic uses of palm and lauric products. *Journal of the American Oil Chemists Society*. 62(2): 431-433.
- Kamble, V. and Gaikwad, N. (2016). Fourier Transform Infrared Spectroscopy Spectroscopic Studies in Embelia Ribes Burm. F.: A Vulnerable Medicinal Plant. *Asian Journal of Pharmaceutical and Clinical Research*. 9(3): 41-47.
- Kanjilal, U.N (1992). Flora of Assam. Periodical Expert Book Agency, pp. 196-202.
- Kar, M.K., Swain, T.R. and Mishra S.K. (2014). Antidiabetic activity of Clerodendrum serratum (L) moon leaves in streptozotocin-induced diabetic rats. *Asian Journal of Pharmaceutical and Clinical Research*. 7(5): 260-263.
- Karthikeyan, A., Shanthi, V. and Nagasathaya, A. (2009). Preliminary phytochemical and antibacterial screening of crude extracts of the leaf of Adhatoda vasica L. *International Journal of Green Pharmacy*. 3(1): 78-80.
- Karunamoorthi, K. and Tsehaye, E. (2012). Ethnomedicinal knowledge, belief and self reported practice of local inhabitants on traditional antimicrobial plants and phytotherapy. *Journal of Ethnopharmacology*. 141(1): 143-150.
- Kaur, S. and Mondal, P. (2014). Study of total phenolic and flavonoid content, antioxidant activity and antimicrobial properties of medicinal plants. *Journal of Microbiology & Experimentation*. 1(1): 23-28.
- Kavipriya, K. and Chandran, M. (2018). FTIR and GCMS Analysis of bioactive Phytocompounds in methanolic leaf extract of Cassia alata. *Biomedical and Pharmacology Journal*. 11(1): 141-147.
- Kavitha, R. (2021). Phytochemical screening and GCMS analysis of bioactive compounds present in ethanol extracts of leaf and fruit of Trichosanthesis dicoica Roxb. *International Journal of Pharmaceutical Sciences and Research*. 12(5): 2755-2764.
- Kebede, T., Gadisa, E. and Tufa, A. (2021). Antimicrobial activities evaluation and phytochemical screening of some selected medicinal plants: A possible alternative in the treatment of multidrug-resistant microbes. *PLoS ONE* 16(3): e0249253. <https://doi.org/10.1371/journal.pone.0249253>.
- Khakhalary, S. and Narzari, S. (2022). Phytochemical profiles of leaf extracts of *Rothea serrata* (L.) Steane & Mabb: a medicinal herb of Assam. *The Journal of the Indian Botanical Society*. 102(1): 2455-7218.

- Khalid, A.S., Shakya, A.K., Naik, R.R., Jaradat, N., Farah, H.S., Shalan, N., Khalaf, N.A. and Oriquat, G.A. (2021). Phenolic content and antioxidant and antimicrobial activities of *Malva sylvestris* L., *Malva oxyloba* Boiss., *Malva parviflora* L., and *Malva aegyptia* L. Leaves Extract. *Journal of Chemistry*. 10(1): 1-10.
- Khalil, E., Esoh, R., Rababah, T., Ali, M. A. and Muhammad, H. A. (2010). Minerals, proximate composition and their correlations of medicinal plants from Jordan. *Journal of Medicinal Plants Research*. 6(47): 5757-5762.
- Khan, M.S., Yusufzai, S.K., Kaun, L.P., Shah, M.D. and Idris, R. (2016). Chemical composition and antioxidant activity of essential oil of leaves and flowers of *Alternanthera sessilis* red from Sabah. *Journal of Applied Pharmaceutical Science*. 6(12): 157-161.
- Khan, S., Richa, Kaur, H. and Jhamta, R. (2019). Evaluation of antioxidant potential and phytochemical characterization using GCMS analysis of bioactive compounds of *Achillea filipendulina* (L.) Leaves. *Journal of Pharmacognosy and Phytochemistry*. 8(3): 258-265.
- Khan, S.A., Baeshen, M.N., Ramadan, H.A. and Baeshen, H.A. (2018). TS2: An ideal DNA barcode for the arid medicinal plant *Rhazya stricta*. *Academia Journal of Medicinal Plants*. 6(7): 000-000.
- Khan, S.M., Khan, S.Y., Hanry, E.L., Rafatullah, M. and Elison, B.B. (2019). GC-MS Analysis of chemical constituents and in vitro antioxidant activity of the organic extracts from the stem of *Bridelia Stipularis*. *Sains Malaysiana*. 48(5): 999-1009.
- Khan, T., Ali, M., Khan, A., Nisar, P., Jan, S.A., Afridi, S. and Shinwari, Z.K. (2019). Anticancer plants: A review of the active phytochemicals, applications in animal models, and regulatory aspects. *Biomolecules*. 10(1): 1-30.
- Khare, C.P. (2004). *Encyclopedia of Indian Medicinal Plants*. Springer, Berlin.
- Kiliobas, K.S.A., Clarkson, G.P. and Artimas, S.P. (2019). Phytochemical analysis, proximate composition and antinutritional factors of *Corchorus oliterius* plant. *International Journal of Biological and Chemical Sciences*. 13(4): 2147-2157.
- Kim, S., Chen, J., Cheng, T., Gindulyte, A., He, J., He, S., Li, Q., Shoemaker, B.A., Thiessen, P.A., Yu, B., Zaslavsky, L., Zhang, J. and Bolton, E.E. (2019). PubChem in 2021: new data content and improved web interfaces. *Nucleic Acids Research* 49(D1):D1388-D1395.

- Kim, S.J., Chung, W.S., Kim, S.S., Ko, S.G. and Um, J.Y. (2011). Antiinflammatory effect of *Oldenlandia diffusa* and its constituent, hentriacontane through suppression of caspase-1 activation in mouse peritoneal macrophages. *Phytotherapy Research*. 25(10): 1537-1546.
- Kim, S.M., Bhosale, P.B., Ha, S.E., Vetrivel, P., Kim, H.H. and Kim, G.S. (2020). Functions of polyphenols and its anticancer properties in biomedical research: a narrative review. *Translational Cancer Research*. 9(12): 7619-7631.
- Kimura, M. (1980). A Simple Method for Estimating Evolutionary Rates of Base Substitutions through Comparative Studies of Nucleotide Sequences. *Journal of Molecular Evolution*. 16(2): 111-120.
- Kinghorn, A.D. and Gupta, A.K. (2003). Quality Standards of Indian Medicinal Plants, Coordinator. (Medicinal Plants Unit, Indian Council of Medical Research). Indian Council of Medicinal Research, Ansari Nagar, New Delhi, India. (1): 0972-13.
- Kirtikar, K.R. and Basu, B.D. (1993). Indian Medicinal Plants. 1. Periodical Expert Book Agency. 459-465.
- Kma, L., Marbaniang, C. and Nath, R.S. (2020). In-vitro comparative studies of *Apium graveolens* L. extracts for antioxidant and anti-inflammatory activity. *The NEHU Journal*. XVIII (1): 43-59.
- Konstantynowicz-Nowicka, K., Berk, K., Chabowski, A., Kasacka, I., Bielawiec, P., Łukaszuk, B. and Harasim-Symbor, E. (2019). High-fat feeding in time-dependent manner affects metabolic routes leading to nervonic acid synthesis in NAFLD. *International Journal of Molecular Science*. 20(15): 3829.
- Kosalge, S.B. and Fursule, R.A. (2009). Investigation of ethnomedicinal claims of some plants used by tribals of Satpuda Hills in India. *Journal of Ethnopharmacology*. 121(3): 456-461.
- Kotoky, J., Choudhury, B., Kandimalla, R., Bharali, R., Monisha, J., Kunnumakara, A.B. and Kalita, K. (2016). Anticancer activity of *Garcinia Morella* on T-Cell Murine Lymphoma via Apoptotic Induction. *Frontiers in Pharmacology*. 7(3): 1-13.
- Kozat, S. (2007). Serum T3 and T4 concentrations in lambs with nutritional myodegeneration. *Journal of Veterinary Internal Medicine*. 21(5): 1135-1137.

- Krause, M., Fayh, A.P. and Reischak-Oliveira, A. (2017). L-arginine usage in Type I diabetes: From the autoimmune event to human dietary supplementation. In: *L-Arginine in Clinical Nutrition*. Switzerland: Humana Press. 395-406.
- Kress, W.J., Garcı-Robledo, C., Uriarte, M. and Erickson, D.L. (2014). DNA barcodes for ecology, evolution and conservation. *Trends in Ecology and Evolution*. 30(1): 25-35.
- Kress, W.J., Wurdack, K.J., Zimmer, E.A., Weigt, L.A. and Janzen, D.H. (2005). Use of DNA barcodes to identify flowering plants. *Proceeding of the National Academic Sciences of the United States of America*. 102(23): 8369-74.
- Krippner, R. and Staples, J. (2003). Suspected allergy to artemether-lumefantrine treatment of malaria. *Journal of Travel Medicine*. 10(5): 303-305.
- Krishna, V., Vidya, S.M., Manjunatha, B.K., Ahmed, M. and Singh, J.S.D. (2007). Evaluation of hepatoprotective activity of *Clerodendrum serratum*. *Indian Journal Experimental Biology*. 45(6): 538-542.
- Krishnamoorthy, B. and Kanabiran, K. (2020). Actinomycetes mediated targeting of drug resistant MRSA pathogens. *Journal of King Saud University- Science*. 32(1): 260-264.
- Krishnamoorthy, K. and Subramaniam, P. (2014). Phytochemical profiling of leaf, stem and tuber parts of *Solena amplexicaulis* (Lam.) Gandhi using GCMS. *International Scholarly Research Notices*. 1-13. <http://dx.doi.org/10.1155/2014/567409>.
- Krishnan, K., Mani, A. and Jasmine, S. (2014). Cytotoxic activity of bioactive compound 1, 2- Benzene dicarboxylic acid, mono 2- ethylhexyl ester extracted from a Marine derived *Streptomyces* sp. *International Journal of Molecular and Cellular Medicine*. VITSJK8. 3(4): 246-254.
- Kroemer, G., Dallaporta, B. and Resche-Rigon, M. (1998). The mitochondrial death/life regulator in apoptosis and necrosis. *Annual Review Physiology*. 60(1): 619-642.
- Kumar, A., Iyer, K., Shanthi, V. and Ramanathan, K. (2014). Extraction of Bioactive Compounds from *Millingtonia hortensis* for the Treatment of Dapsone Resistance in Leprosy. *Journal of Microbial and Biochemical Technology*. 6(5): 1-6.
- Kumar, A., Mansoori, A., Singh, N., Dubey, S.K., Thakur, T.K., Alkan, N. and Das, S.N. (2020). Phytochemical characterization and assessment of crude extracts from

Lantana camara L. for antioxidant and antimicrobial activity. *Frontiers in Agronomy*. 2: 1-14.

Kumar, A.A., Abuthahi, S.S.S. and Aboul-Enein, H.Y. (2022). Phytochemical extraction and comparative analysis of antioxidant activities of Areca catechu L. nut extracts. *Pharmacia*. 69(2): 447-451.

Kumar, P. and Niteshwar, K. (2013). Phytochemical and pharmacological profiles of *Clereodendrum serratum*. *International Journal of Research in Ayurveda and Pharmacy*. 4(2): 276-8.

Kumar, S. and Muller, K. (1999). Inhibition of keratinocyte growth by different Nepalese *Zanthoxylum* species. *Phytotherapy Research*. 13: 214-217.

Kumar, G.M.P. and Shiddamallayya, N. (2021). Nutritional and anti-nutritional of wild edible plants in Hassan district of Karnataka, India. *Indian Journal of Natural Products and Resources*. 12(2): 281-290.

Kumar, S. and Oanday, A.K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*. 1-17.

Kuppuswamy, K.M., Jonnalagadda, B. and Arockiasamy, S. (2013) .GC-MS analysis of chloroform extract of *Croton bonplandianum*. *International Journal of Pharma and Bio Science*. 4(4): 613-617.

Kurutas, E.B. (2015). The Importance of antioxidants which play the role in cellular response against oxidative/ nitrosative stress: Current State. *Nutrition Journal*. 15(1): 1-22.

Kusmardiyani, S., Novita, G. and Fidrianny, I. (2016). Antioxidant activities from various extracts of different parts of kelakai (*Stenochlaena palustris*) grown in central kalimantan Indonesia. *Asia Journal of Pharmaceutical Clinical Research*. 9(2): 215-219.

Labaran, I., Lukman, O., Afolabi, A.A., Jamil, D. and Umar, M. (2016). Analysis of some phytochemical and minerals found in aqueous stem bark extract of *Albizia lebbeck*. *Dutse Journal of Pure Applied Science*. 2(1): 231-7.

Laglaoui, A., Barchan, A., Bakkali, M., Arakrak, A. and Pagan, R. (2014). The effects of solvents polarity on the phenolic contents and antioxidant activity of three *Mentha* species extracts. *International Journal of Current Microbiology and Applied Science*. 3(11): 399-412.

- Lakshmi, N. D. M., Mahitha, B. and Madhavi, T.(2015). Phytochemical screening and FTIR analysis of Clitoria ternatea leaves. *International Journal of Scientific & Engineering Research*. 6(2): 287-290.
- Lalrinzuali, K., Vabeiryureilai, M. and Jagetia, G.C. (2021). Sonapatha (*Oroxylum indicum*) mediates cytotoxicity in cultured HeLa cells by inducing apoptosis and suppressing NF- κ B, COX-2, RASSF7 and NRF2. *Bioorganic Chemistry*. 114: 105126.
- Lalthanpuui, P.B. and Lalchhandama, Z.K. (2019). Chemical profiling, antibacterial and antiparasitic studies of Imperata cylindrical. *Journal of Applied Pharmaceutical Science*. 9(12): 117-121.
- Langley, M.R., Triplet, E.M. and Scarisbrick, I.A. (2020). Dietary influence on central nervous system myelin production, injury, and regeneration. *BBA-Molecular Basis of Diseases*. 1866(7): 165779.
- Lapornik, B., Prosek, M. and Wondra, A.G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering*. 71(2): 214-222.
- Lawal, I. (2013). Phytotherapy approach for the treatment of Gynaecological disorder among women in Ido Local Government Area of Ibadan, Oyo State, Nigeria. *Journal of Advanced Science Research*. 4(3): 41-44.
- Lee, L.H., Bakrim, S., Omari, N.E., Hachlafi, N.E., Bakri, Y. and Bouyahya, A. (2022). Dietary phenolics compounds as anticancer natural drugs: Recent update on molecular mechanisms and clinical trials. *Foods*. 11(21): 3323.
- Lee, Y.L., Huang, G.W., Liang, Z.C. and Mau, G.L. (2007). Antioxidant properties of three extract from Pleurotus citrinopileatus. *Food Science and Technology*. 40(5): 823-833.
- Leea, J. and Finn, C.E. (2012). Lingonberry (*Vaccinium vitis-idaea* L.) grown in the Pacific Northwest of North America: Anthocyanin and free amino acid composition. *Journal of Functional Foods*. 4(1): 213-218.
- Li, K., Sinclair, A.J., Zhao, F. and Li. D. (2018). Uncommon fatty acids and cardiometabolic health. *Nutrients*. 10(10):1559. Doi: 10.3390/nu10101559.
- Li, M., Cao, H., But, P.P.H. and Shaw, P.C. (2011). Identification of herbal medicinal materials using DNA barcodes. *Journal of Systematics and Evolution*. 49(3): 271-283.

- Li, M., Zhou, L., Yang, D., Li, T. and Li, W. (2012). Biochemical composition and antioxidant capacity of extracts from *Podophyllum hexandrum* rhizome. *BMC Complementary Alternative Medicine*. 12: 263-263.
- Li, Z., Huang, Y., Chen, X. and Chen, J. (2019). DNA barcoding and phylogenetic relationships in *Omphalogramma* (Primulaceae) from the Hengduan Mountain region of China. *Phytotaxa*. 402 (2): 097-106.
- Linder, M.C. and Azam, M.H. (1996). Copper biochemistry and molecular biology. *American Journal of Clinical Nutrition*. 63(5): 797S-811S.
- Liu SC, Lin JT, Wang CK, Chen, H. and Yang, D. (2009). Antioxidant properties of various solvent extracts from lychee (*Litchi chinensis* sonn.) flowers. *Food Chemistry*. 114(2): 577-81.
- Liu, K., Liu, P.C., Liu, R. and Wu, X. (2015). Dual AO/EB staining to detect apoptosis in osteosarcoma cells compared with flow cytometry. *Medical Science Monitors Basic Research*. 21: 15-20.
- Liz, A. (2006). Nutrition Basics for better health and performance. USA: Kendall/Hunt Publishing Company.
- Lowe, S.W. and Lin, A.W. (2000). Apoptosis in cancer. *Carcinogenesis*. 21(3):485-495.
- Lozano-grande, M.A., Gorinstein, S., Espitia-rangel, E., Dávila-ortiz, G. and Martínez-Ayala, A.L. (2018). Plant sources, extraction methods, and uses of squalene. *International Journal of Agronomy*. 1829160. 1-13.
- Lux, C., Joshi-Barr, S., Nguyen, T., Mahmoud, E., Schopf, E., Fomina, N. and Almutairi, A. (2012). Biocompatible Polymeric Nanoparticles Degrade and Release Cargo in Response to Biologically Relevant Levels of Hydrogen Peroxide. *Journal of American Chemical Society*. 134(38): 15758-15764.
- Machlup, F. and Leeson, K. (1978). Information through the printed word: The dissemination of scholarly, scientific and intellectual knowledge. Praeger, New York.
- Madaan, R., Bansal, G., Kumar, S. and Sharma, A. (2011). Estimation of total phenols and flavonoids in extracts of *Actaea spicata* roots and antioxidant activity studies. *Indian Journal of Pharmaceutical Sciences*. 73(6): 666-669.
- Madiha, Y.I., Rukayadi, Y. and Norhayati, H. (2017). Effects of extraction conditions on yield, total phenolic contents and antibacterial activity of methanolic

Cinnamomum zeylanicum Blume leaves extract. *International Food Research Journal*. 24(2): 779-86.

Magnusson, J., Ekstrom, S., Kull, I., Hakansson, N., Nilsson, S., Wickman, M., Melen, E., Riserus, U. and Bergstrom, A. (2018). Polyunsaturated fatty acids in plasma at 8 years and subsequent allergic disease. *Journal of Allergy and Clinical Immunology*. 142(2): 510-516.

Mahadani, P., Sharma, G.D. and Ghosh, S.K. (2012). Identification of ethnomedicinal plants (Rauvolfioideae: Apocynaceae) through DNA barcoding from Northeast India. *Pharmacognosy Magazine*. 9(35): 255-263.

Mahato, K., Kakoti, B.B., Borah, S. and Kumar, M. (2015). In-vitro antioxidative potential of methanolic aerial extracts from three ethnomedicinal plants of Assam: A Comparative Study. *Journal of Applied Pharmaceutical Science*. 5(12): 111-116.

Maitera, O.N. and Chukkol, I.B. (2016). Phytochemical and Fourier Transform Infrared Spectroscopy Analysis of Faidherbia albida (Del) as a preservative agent. *World Journal of Research and Review*. 3(3): 25-29.

Majumder, R., Tiwari, K.C., Bhattacharjee, S. and Nair, A.R. (1978). Some folklore medicines from Assam and Meghalaya. *International Journal of Crude Drug Research*. 16(4): 185-189.

Mallick, S. and Ojah, M. (2022). Ethnomedicinal uses of different ailments among the Koch-Rajbongshis of Agia, Goalpara District, Assam. *Turkish Online Journal of Qualitative Inquiry*. 13(1): 104-113.

Mallikadevi, T., Paulsamy, S., Jamuna, S. and Karthika, K. (2012). Analysis for phytoceuticals and bioinformatics approach for the evaluation of therapeutic properties of whole plant methanolic extract of Mukia maderaspatana (L.) M. Roem. (Cucurbitaceae) - A traditional medicinal plant in western districts of Tamil Nadu, India. *Asian Journal of Pharmaceutical Clinical Research*. 5(4): 163-168.

Mammen, D., Daniel, M. and Sane, R.T. (2010). Seasonal and geographical variations in chemical constituents of Leptadenia reticulata. *International Journal of Pharmaceutical Sciences Review and Research*. 4(2): 111-6.

Man, S., Gao, W., Zhang, Y., Huang, L. and Liu, C. (2010). Chemical study and medical application of saponins as anti-cancer agents. *Fitoterapia*. 81(7): 703-714.

Manikandan, R., Anand, A.V., Rengaraj, R.L., Vinayakamoorthi, P. and Ramalingam, R. (2017). In vitro Antioxidant Activity of Extracts of Psidium

guajava Leaves. *A Multifaceted Peer Reviewed Journal in the field of Pharmacology, Toxicology and Biomedical Reports*. 3(3): 50-54.

Manikrao, V.G. and Singh, S.P. (2013). Ethnomedicinal plants used in the treatment of skin diseases in Hyderabad Karnataka region, Karnataka, India. *Asian Pacific Journal of Tropical Biomedicine*. 3(11): 882-886.

Mao, A.A. and Hynniewta, T.M. (2000). Floristic diversity of North East India. *Journal of Assam Science Society*. 41(4): 255-266.

Maobe, M.A.G. and Nyarango, R.M. (2013). Fourier Transformer InfraRed Spectrophotometer Analysis of *Warburgia ugandensis* Medicinal herb used for the Treatment of Diabetes, Malaria and Pneumonia in Kisii Region, Southwest Kenya. *Global Journal of Pharmacology*. 7(1): 61-68.

Marangoni F., Agostoni C., Borghi C., Catapano A.L., Cena H., Ghiselli A., La Vecchia C., Lercker G., Manzato E., Pirillo A. (2020). Dietary linoleic acid and human health: Focus on cardiovascular and cardiometabolic effects. *Atherosclerosis*. 292: 90-98.

Marineli, R.S., Furlan, C.P.B., Marques, A.C., Bicas, J., Pastore, G.M. and Marostica, M.R. (2012). Phytosterols: Biological effects and mechanisms of hypocholesterolemic action. *Biotechnology of Bioactive Compounds: Sources and Applications*, First Edition. Published 2015 by John Wiley & Sons, Ltd. 565-581.

Marinova D., Ribarova F. and Atanassova M. (2005). Total phenolics and total flavonoids in Bulgarian fruits and vegetables. *Journal of University and Chemical Technology and Metallurgy*. 40(3): 255-260.

Mark, F., McCarty, B.A., James, H.O.K.M.D., James, J. D. and Pharm, D. (2018). Dietary Glycine Is Rate-Limiting for Glutathione Synthesis and May Have Broad Potential for Health Protection. *Ochsner Journal*. 18(1): 81-87.

Martinez, Y., Li, X., Liu, G., Bin, P., Yan, W., Mas, D. and Yin, Y. (2017). The role of methionine on metabolism, oxidative stress and diseases. *Amino Acids*. 49(12): 2091-2098.

Medhi, K., Purohit, B.P. Gogoi, A.J. Saikia, S.P. Kanjilal, P.B. and Bhau, B.S. (2009). Molecular marker based genetic diversity analysis of two economically important plants (*Zanthoxylum hamiltonianum* and *Nepenthes khasiana*) of North-East India. *Biodiversity in herbal medicine*. 47-55.

- Mekonnen, A., Atlabachew, M. and Kassie, B. (2018). Investigation of antioxidant and antimicrobial activities of *Euclea schimperii* leaf extracts. *Chemical and Biological Technologies in Agriculture*. 5(16): 1-12.
- Mensah-Agyei, G.O., Ayeni, K.I. and Ezeamagu. C.O. (2020). GC-MS analysis of bioactive compounds and evaluation of antimicrobial activity of the extracts of *Daedalea elegans*: a Nigerian mushroom. *African Journal of Microbiology Research*. 14(6): 204-210.
- Mereen, J.E. and Daiz, J.H.J. (2020). GCMS analysis and Inflammatory activity of *Murex tribulus*. *International Journal of Pharmaceutical Sciences*. 11(12): 6184-6188.
- Miao, M., Warren, A., Song, W.B., Wang, S, Shang, H.M. and Chen, Z. (2008). Analysis of the internal transcribed spacer 2 (ITS2) region of scuticociliates and related taxa (Ciliophora, Oligohymenophorea) to infer their evolution and phylogeny. *Protist*. 159(4): 519-533.
- Miliauskas, G., Venskutonis, P.R. and Van, B.T.A. (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chemistry*. 85(2): 231-237.
- Min, G. and Chun-Zhao, L. (2005). Comparison of techniques for the extraction of flavonoids from cultured cells of *Saussurea medusa* Maxim. *World Journal of Microbiology and Biotechnology*. 21: 1461-1463.
- Mishra, V.K., Passari, A.K., Vanlalhmangaihi, K., Kumar, N.S. and Singh, B.P. (2015). Antimicrobial and antioxidant activities of *Blumea lanceolaria* (Roxb.). *Journal of Medicinal Plant Research*. 9(4): 84-90.
- Mohamed, A. (2016). DNA Barcoding of five medicinal plants from Siwa Oasis, Egypt. *KMITL Science and Technology Journal*. 16(2): 49-56.
- Mohammad, S.A., Saka, S., Mairiga, I.A., Sanda, K.A., Shuaibu, A. and Ibrahim, A. (2014). Preliminary phytochemical and elemental analysis of aqueous and fractionated pod extracts of *acacia nilotica* (Thorn mimosa). *Veterinary Research Forum*. 5(2): 95-100.
- Mohammed, L., Elkhamlichia, A., El Hajajia, H., Farajb, H., Alamib, A. and Balia, B.E. (2017). Phytochemical screening and evaluation of antioxidant and antibacterial activities of seeds and pods extracts of *Calycotome villosa* subsp. *Intermedia*. *Journal of Applied Pharmaceutical Science*. 7(04): 192-198.

- Mohammed, S.A., Saka, S., Ismail, A.M., Sanda, K.A., Shuaibu, A. and Ibrahim, A. (2014). Preliminary phytochemical and elemental analysis of aqueous and fractionated pod extracts of *Acacia nilotica* (Thorn mimosa). *Veterinary Research Forum*. 5(2): 95-100.
- Mohan, V.R., Muthulakshmi, A. and Jothibai, M.R. (2012). GC-MS Analysis of bioactive components of *Feronia elephantum* Correa (Rutaceae). *Journal of Applied Pharmaceutical Science*. 02 (02): 69-74.
- Mohanty, B., Mahanty, A., Ganguly, S., Sankar, T.V., Chakraborty, K., Rangasamy, A., Paul, B., Sarma, D., Mathew, S., Asha, K.K., Behera, B., Aftabuddin, M., Debnath, D., Vijayagopal, P., Sridhar, N., Akhtar, M.S., Sahi, N., Mitra, T., Banerjee, S., Paria, P., Das, D., Vijayan, K.K., Laxmanan, P.T. and Sharma, A.P. (2014). Amino acid compositions of 27 food fishes and their importance in clinical nutrition. *Journal of Amino Acids*: 1-7.
- Mohsen, M. S. and Ammar, S. M. A. (2008). Total phenolic contents and antioxidant activity of corn tassel extracts. *Food Chemistry*. 112(3): 595-598.
- Monga, N., Sethi, G.S., Kondepudi, K.K. and Naura, A.S. (2020). Lipid mediators and asthma: Scope of therapeutics. *Biochemical Pharmacology*. DOI: 10.1016/j.bcp.2020.113925. in press.
- Moreno-Rojas, J.M., Samtiya, M., Aluko, R.E. and Dhewa, T. (2021). Potential Health Benefits of Plant Food-Derived Bioactive Components: An Overview. *Foods*. 10(4): 839.
- Moshahid, A.R.M., Irshad, M., Zafaryab, M. and Singh, M. (2012). Comparative Analysis of the Antioxidant Activity of *Cassia fistula* Extracts. *International Journal of Medicinal Chemistry*. Article ID 157125: 1-6.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival - application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*. 65(1-2): 55-63.
- Motlhanka, D.M.T., Habtemariam, S. and Houghton, P. (2014). Free Radical Scavenging Activity of Crude Extracts and 4'-O-24. *Methylepigallocatechin Is Organic and Bioorganic Chemistry*. 4(1):1-5.
- Moussa, T.A.A. and Almaghrabi, O.A. (2016). Fatty acid constituents of *Peganum harmala* plant using Gas Chromatography-Mass Spectroscopy. *Saudi Journal of Biological Sciences*. 23(3): 397-403.

- Mugendhiran, S. and Sheeja, B. D. (2020). Evaluation of phytoconstituents of *Erigeron canadensis* L. by FTIR and GC-MS analysis. *International Journal of Pharmaceutical Sciences and Research*. 12(5): 2823-2834.
- Muhammed, I., Muh, S., Olorunju, S., Bale, J., Abdullahi, U. and Lawal, R. (2002). Response of nutrients and anti-nutritional constituents in the seeds of *Cassia tora* L. to treatments. *Journal of Agriculture and Environmental Sciences*. 3: 225-34.
- Muhammed, R., Khalid, R., Muhammad, A., Fozia, A., Fatima, H., Zia, S., Ghulam, R., Egbuna, C., Andrew, G.M. and Chukwuemeli, Z.U. (2023). Phytobioactive compounds as therapeutic agents for human diseases: A review. *Food Science & Nutrition*. 11: 2500- 2529.
- Mujtaba, A., Masud, T., Ahmad, A., Naqvi, S.M.S., Qazalbash, M.A. and Levin, R.E. (2016). Effect of solvents on extraction yield, total flavonoid, total phenolic contents, DPPH scavenging activity and antibacterial potential of three apricot cultivars. *Transylvanian Review*. 10(6): 1662-1676.
- Mukesh, K.R., Gaurav, K., Shiv, K.I., Gotmi, S. and Tripathi, D.K. (2012). *Clerodendrum serratum*: A clinical approach. *Journal of Applied Pharmaceutical Science*. 2(2): 11-15.
- Munda, S. and Kakoti, B.B. (2017). Pharmacological evaluation of methanolic extract of *Zanthoxylum oxyphyllum* Edgew with special reference to anti-inflammatory and analgesic activity. *International Journal of Applied Biology and Pharmaceutical Technology*. 8: DOI: 10.21276/Ijabp.
- Mussarat, S., Salam, N.M.A., Tariq, A., Wazir, S.M., Ullah, R. and Muhammad, A. (2014). Use of Ethnomedicinal plants by the People living around Indus River. *Evidence-Based Complementary and Alternative Medicine*. 1-14.
- Muthuswami, R. and Fathimath, S.V.P. (2020). GC-MS analysis of ethanolic leaf extract of *Achyranthes aspera* Linn. *Journal of Pharmacognosy and Phytochemistry*. 9(4): 1207-1210.
- Mwakalukwa, R., Maregesi, S.M. and Mwashuiya, J. (2016). Comparative Analysis on Nutritional and Antinutritional factors in Raw and Blanched *Crotalaria laburnoides* Klotzsch leaves from Iramba District, Tanzania. *Scholars Academic Journal of Pharmacy (SAJP)*. 5(8): 342-349.
- Nabavi, S., Ebrahimzadeh, M.A., Nabavi, S.F., Hamidinia, A. and Bekhradnia, A.R. (2008). Determination of antioxidant activity, phenol and flavonoids content of *Parrotia persica* Mey. *Pharmacologyonline*. 2: 560-567.

- Nabavi, S.F., Nabavi, S.M., Hellio, C., Alinezhad, H., Zare, M., Azimi, R. and Baharfar, R. (2012). Antioxidant and antihemolytic activities of methanol extract of *Hyssopus angustifolius*. *Journal of Applied Botany and Food Quality*. 85: 1-4.
- Nabi, M., Tabassum, N. and Ganai, B.A. (2022). Phytochemical screening and antibacterial activity of *Skimmia anquetilia* N.P. Taylor and Airy Shaw: A first study from Kashmir Himalaya. *Frontiers in Plant Science*. 13: 937946. doi: 10.3389/fpls.2022.937946.
- Nachiyar, V.C., Renugadevi, K. and Zaveri, M. (2020). Bioactivity of Dodecanoic Acid Extracted from *Geitlerinema* sp. TRV57. *Indian Journal of Pharmaceutical Education and Research*. 55(1): 224-231.
- Naeini Z., Toupchian O., Vatannejad A., Sotoudeh G., Teimouri M., Ghorbani M., Nasli-Esfahani E., Koohdani F. (2020). Effects of DHA-enriched fish oil on gene expression levels of p53 and NF- κ B and PPAR- γ activity in PBMCs of patients with T2DM: A randomized, double-blind, clinical trial. *Nutritional, Metabolism and Cardiovascular diseases*. 30: 441-447.
- Naim, D.M., Husin, C.C.N. and Ahmad, M.I. (2018). Authentication of a selected medicinal plants using DNA barcoding technique. *SHS Web of Conferences* 45, 05004.
- Nakamura, M., Ra, J.H., Jee, Y. and Kim, J.S. (2017). Impact of different partitioned solvents on chemical composition and bioavailability of *Sasa quelpaertensis* Nakai leaf extract. *Journal of Food and Drug Analysis*. 25(2):316-326.
- Namukobe, J., Sekandi, P., Byamukama, R., Murungi, M., Nambooze, J., Ekyibetenga, Y., Nagawa, C.B. and Asimwe, S. (2021). Antibacterial, antioxidant, and sun protection potential of selected ethno medicinal plants used for skin infections in Uganda. *Tropical Medicine and Health*. 49(1): 1-10.
- Naseem, U., Muhammad, Z., Farhat, A.K. and Shazeb, K. (2014). A review on general introduction to medicinal plants, its phytochemicals and role of heavy metal and inorganic constituents. *Life Science Journal*. 11(7s): 520-527.
- Nath, N. (2008). Morphological and palynological features of some common medicinal plants of Goalpara district, Assam. *Environement & Ecology*. 24(4C). 2285-2287.
- Navarro, A., De, I.H.B. and Villar, A. (2001). Anti-inflammatory and immunomodulating properties of a sterol fraction from *Sideritis foetens*. *Biological & Pharmaceutical Bulletin*. 24(5): 470-473.

- Nemati, F., Ali, A.D., Bahman, E., Vahid, M. and Sepideh, M. (2013). Cytotoxic Properties of Some Medicinal Plant Extracts from Mazandaran, Iran. *Iranian Red Crescent Medical Journal*. 15(11): e8871.
- Nengroo, Z.R. and Rauf, A. (2019). Fatty acid composition and antioxidant activities of five medicinal plants from Kashmir. *Industrial Crops and Products*. 140():<https://doi.org/10.1016/j.indcrop.2019.111596>.
- Newmark, H.L. (1999). Squalene, olive oil and cancer risk. Review and hypothesis. *Annals of the New York Academy Science*. 889(1): 193-203.
- Newsholme, P., Keane, K.N., Elahy, M. and Cruzat, V.F. (2017). L-arginine, pancreatic beta cell function, and diabetes: Mechanisms of stimulated insulin release and pathways of metabolism. In: Patel V, Preedy V, Rajendram R, editors. In: L-Arginine in Clinical Nutrition. Switzerland: Humana Press; p. 85-4.
- Nguyen, Q.V. and Eun, J.B. (2011). Antioxidant activity of solvent extracts from Vietnamese medicinal plants. *Journal of Medicinal Plants Research*. 5(13): 2798-2811.
- Niciforovic, N., Mihailovic, V., Maskovic, P., Solujic, S., Stojkovic, A. and Muratspahic, D.P. (2010). Antioxidant activity of selected plant species, potential new sources of natural antioxidants. *Food and Chemical Toxicology*. 48(11): 3125-3130.
- Ningombam, D.S. and Singh, P.K. (2014). Ethnobotanical study of *Phlogacanthus thyriformis* Nees: A conserved medicinal plant of Manipur, Northeast India. *International Journal of Herbal Medicine*. 1(5): 10-14.
- Nishat, F.S., Neog, N. and Saikia, M.B. (2012). Phytochemical analysis of *Lasia spinosa* and *Alpinia nigra*, potential medicinal plants of Assam. *Medicinal Plants. International Journal of Phytomedicines and Related Industries*. 4(3): 0975- 4261.
- Nithyadevi, J. and Sivakumar, R. (2015). Phytochemical screening and GC-MS, FT-IR analysis of methanolic extract leaves of *Solanum torvum* Sw. *International Journal of Research Studies in Bioscience*. 3(9): 61-66.
- Njoku N. E., Ubbaonu C. N., Alagbaoso S.O., Agunwa I.M. and Eluchie C.N. (2016). Proximate, Anti-Nutritional and Phytochemical Composition of the Yellow Variety of the *Synsepalum Dulcificum* (Miracle Fruit) Berry. *American Journal of Food Science and Technology*. 4(4): 102-108.
- Njoku, C.E., Alaneme, K.K., Omotoyinbo, J.A., Ekeleme, A.C., Ugwu, E.I. and Ikele, U.S. (2021). Phytochemical, Proximate and Mineral Analysis of *Urena lobata*

Stems from Imo State Nigeria. *International Conference on Engineering for Sustainable World. Material Science and Engineering*. doi:10.1088/1757-899X/1107/1/012183.

Nonglang, F.P., Bhan, S., Khale, A. and Wankhar, W. (2022). Pharmacognostic evaluation of *Eranthemum indicum* extracts for its in-vitro antioxidant activity, acute toxicology and investigation of potent bioactive phytochemicals using HPTLC and GCMS. *Beni Suez University Journal of Basic and Applied Sciences*. 11(129): 1-17.

Novak, W.K. and Haslberger, A.G. (2000). Substantial equivalence of antinutrients and inherent plant toxins in genetically modified novel foods. *Food and Chemical Toxicology*. 38(6): 473-483.

Noviany, N., Amrulloh, M.H., Mohamad, R., Irawan, B., Kusuma, W.A., Hadi, S., Supriyanto, R., Nofiani, R., Hussin, M.H. and Yuwono, S.W. (2023). FTIR-Based metabolomics for characterization of antioxidant activity of different parts of *Sesbania grandiflora* plant. *Sains Malaysiana*. 52(1): 165-174.

Obazelu, P.A., Aruomaren, A.L. and Ugboaja, E.E. (2021). Phytochemical Analysis, Nutrients and Mineral Composition of *Combretum platypterum* Aqueous Leaf Extract. *Journal of Applied Sciences and Environmental Management*. 25(9): 1625-1630.

Ogie, B.M., Ha-thi, A.D., Mulokozi, G. and Hambraeus, L. (2001). Micronutrient composition and nutritional importance of gathered vegetables in Vietnam. *International Journal of Food Science Nutrition*. 52(6): 485-499.

Oguchi, Y., Weerakkody, W.A.P., Tanaka, A., Nakazawa, S. and Ando, T. (1996). Varietal differences of quality-related compounds in leaves and petioles of spinach grown at two locations. *Bulletin of Hiroshima Prefectural Agricultural Research Centre*. 64(5): 1-9.

Ogunka-Nnoka, C.U., Igwe, F.U., Agwu, J., Peter, O.J. and Wolugbom, P.H. (2020). Nutrient and Phytochemical Composition of *Centella asiatica* Leaves. *Medicinal & Aromatic Plants*. 9(2): 1-7.

Oguoma, O.I., Ezeifeke, G.O., Adeleye, S.A., Oranusi, S. and Amadi, E.S. (2015). Antimicrobial activity, proximate and amino acid analysis of *Tetrapleura tetraptera*. *Nigerian Journal of Microbiology*. 27(1): 2709-2718.

Ogwok, P., Bamuwamye, M., Apili, G. and Musalima, J.H. (2014). Health risk posed by lead, copper and iron via consumption of organ meats in Kampala City (Uganda). *Journal of Environment Pollution and Human Health*. 2(3): 69-73.

- Oktay, M., Gulcin, I. and Kufrevioglu, O.I. (2003). Determination of in-vitro antioxidant activity of fennel (*Foeniculum vulgare*) seed extracts. *Food Science and Technology*. 36(2): 263-271.
- Ola, F.L. and Oboh, G. (2000). Anti-nutritional factors in nutritional quality of plant foods. *Journal of Technology*. 4(15): 1-3.
- Olanrewaju, C. A. and Ahmed, F. (2014). Proximate analysis and phytochemical screening of some medicinal plants commonly used by Guaris of Fct, Nigeria. *International Journal of Current Research*. 6(06): 6964-6967.
- Olayinka, B.U., Abdulbaki, A.S., Alsamadany, H., Alzahrani, Y., Omorinoye, O.A., Olagunju, G.R. and Sulyman, A. (2023). Proximate composition, amino acid and fatty acid profiles of eight cultivars of ground nut grown in Nigeria. *Legume Research- An International Journal*. 46(1): 95-99.
- Olivia, N.U., Goodness, U.C. and Obinna, O.M. (2021). Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of *Hibiscus asper* leaves. *Future Journal of Pharmaceutical Sciences*. 7(59): 1-5.
- Olumide, M.D., Ajayi, O.A. and Akinboye, O.E. (2019). Comparative study of proximate, mineral and phytochemical analysis of the leaves of *Ocimum gratissimum*, *Vernonia amygdalina* and *Moringa oleifera*. *Journal of Medicinal Plant Research*. 13(15): 351-356.
- Olusanya, J. O. (2008). Proteins, In: Essential of food and nutrition, Apex Books Limited. Pp. 13-21.
- Omar, W.A.W. and Zain, S.N.D.M. (2018). Antioxidant activity, total phenolic content and total flavonoid content of water and methanol extracts of *Phyllanthus* species from Malaysia. *Pharmacognosy Journal*. 10(4): 677-681.
- Omotoso, O.T. (2006). Nutritional quality, functional properties and antinutrient composition of the larva of *Cirna forda* (Westwood) (Lepidoptera: Saturniidae). *Journal of Zhejiang University, Science B*. 7(1): 51- 55.
- Oni, J.O., Akomaye, F.A., Uman, E.J. and Damian, A.O. (2021). Amino acid profiles and bioactive compounds of four inedible mushrooms from Oban Division Cross River National Park (CRNP), Nigeria. *European Journal of Biology and Biotechnology*. 2(2): 11-18.
- Onwuka, G.I. (2005). Food Analysis and Instrumentation; Theory and Practice. Naphthalic prints, Surulere, Lagos, Nigeria. 219- 230.s

- Orienti, I., Zuccari, G., Bergamante, V., Fini, A., Corosio, R. and Montaldo, P.G. (2007). Enhancement of oleyl alcohol anti-tumor activity through complexation in polyvinyl alcohol amphiphilic derivatives. *Drug Delivery*. 14(4): 209-17.
- Otunola, G.A., Abifarin, T.O. and Afolayan, A.J. (2020). Nutritional composition and antinutrient content of *Heteromorpha arborescens* (Spreng.) Cham. & Schltldl. leaves: An underutilized wild vegetable. *Food Science and Nutrition*. 9(1): 172-179.
- Ouyang, H., Kong, X.Z, He, W., Qin, N., He, Q.N., Wang, Y., Wang, R. and Xu, F. (2012). Effects of five heavy metals at sub-lethal concentrations on the growth and photosynthesis of *Chlorella vulgaris*. *Chinese Science Bulletin*. 57(25): 3363-3370.
- Oyeyinka, B.O. and Afolayan, A.J. (2019). Comparative Evaluation of the Nutritive, Mineral and antinutritive composition of *Musa sinensis* L. (Banana) and *Musa paradisiaca* L. (Plantain) fruit compartments. *Plants*. 8(12): 598.
- Ozen, T. and Kinalioglu, K. (2008). Determination of antioxidant activity of various extracts of *Parmelia saxatilis*. *Biologia*. 63(2): 211-216.
- Ozsoy, N., Can, A., Yanardag, R. and Akev, A. (2008). Antioxidant activity of *Smilax excels* L. leaf extracts. *Food Chemistry*. 110: 571-583.
- Padmashree, M.S., Roopa, B., Ashwathanarayana, R. and Naika, R. (2018). Antibacterial properties of *Ipomoea staphylina* Roem & Schult. plant extracts with comparing its preliminary qualitative phytochemical and quantitative GC-MS analysis. *Tropical Plant Research*. 5(3): 349-369.
- Palhares, R. M., Drummond, M. G., Cosenza, G. P., Das Graças Lins Brandao, M. and Oliveira, G. (2015). Medicinal plants recommended by the world health organization: DNA barcode identification associated with chemical analyses guarantees their quality. *PLoS One*. 10(5): 1-29.
- Pamulaparathi, A., Prathap, V.R., Banala, M. and Nanna, R. S. (2016). Total phenolic, flavonoid contents and antioxidant assays in leaf extracts of *Senna alata* (L.) Roxb. *Journal of Pharmaceutical Sciences and Research*. 8(9): 981-985.
- Pan, Y., Wang, K., Huang, S.Q., Wang, H.S., Mu, X.M., He, C.H., Ji, X.W., Zhang, J. and Huang, F.J. (2008). Antioxidant activity of microwave-assisted extract of longan (*Dimocarpus Longan* Lour.) peel. *Food Chemistry*. 106(3): 1264-1270.
- Pandey, A.K. and Kumar, S. (2013). Chemistry and biological activities of Flavonoids: An overview. *The Scientific World Journal*. Volume 2013 | Article ID 162750. 1-6.

- Pandey, G., Jain, G.C. and Mathur, N. (2012). Therapeutic potential of metals in managing diabetes mellitus: a review. *Journal of Molecular Pathophysiology*. 1(1): 63-76.
- Pandey, M.M., Rastogi, S. and Rawat, A.K. (2013). Indian traditional ayurvedic system of medicine and nutritional supplementation. *Evidence based Complementary and Alternative Medicine*. 13:1-12.
- Pandit, B.R., Kotiwar, O.S., Oza, R.A. and Kumar, R.M. (1996). Ethno-medicinal plant lore from Gir Forest Gujrat. *Advance in Plant Science*. 9(1): 81-84.
- Park, G.H., Kwon, H.Y., Park, Y., Kwon, S.H., Jang, Y.L., Lee, D.H., Yang, S.O. and Hyun, J.E. (2022). Nutritional composition and phytochemical screening in different parts of *Hibiscus syriacus* L. *Food Science and Nutrition*. 10(9): 3034-3042.
- Pasalar, P., Khaghani, S., Razi, F., Yajloo, M.M., Paknejad, M. and Ahmad, S. (2011). Selective Cytotoxicity and Apoptogenic Activity of Hibiscus Sabdariffa Aqueous Extract against MCF-7 Human Breast Cancer Cell Line. *Journal of Cancer Therapy*. 2(3): 394-400.
- Patel, J.J., Acharya, S.R. and Acharya, N.S. (2014). Clerodendrum serratum (L.) Moon. - A review on traditional uses, phytochemistry and pharmacological activities. *Journal of Ethnopharmacology*. 154(2): 268-285.
- Patel, M.B. and Mishra, S. (2011). Hypoglycemic activity of alkaloidal fraction of *Tinospora cordifolia*. *Phytomedicine*. 18(12): 1045-52.
- Patel, S., Gheewala, N., Suthar, A. and Shah, A. (2009). In-Vitro cytotoxicity activity of *Solanum nigrum* extract against Hela Cell Line and Vero Cell Line. *International Journal of Pharmacy and Pharmaceutical Sciences*. 1(1): 38-46.
- Paudel, M.R. and Pant, B. (2017). Cytotoxic activity of crude extracts of *Dendrobium amoenum* and detection of bioactive compounds by GC-MS," *Botanica Orientalis: Journal of Plant Science*. 6(11): 38-42.
- Paul, M. and Devi, N. (2021). GC-MS and FT-IR Analysis of methanol Fruit Extract of *Ficus Racemosa* and *Ficus Auriculata*. *International Journal of Pharmaceutical Science and Research*. 12(3): 1679-1684.
- Paul, R.K., Irudayaraj, V., Johnson, M. and Patric, R. D. (2011). Phytochemical and anti-bacterial activity of epidermal glands extract of *Christella parasitica* (L.) H. Lev. *Asian Pacific Journal of Tropical Biomedicine*. 1(1): 8-11.

- Paulsamy, S., Mallikadevi, T., Jamuna, S. and Karthika, K. (2012). Analysis for phytochemicals and bioinformatics approach for the evaluation of therapeutic properties of whole plant methanolic extract of *Mukia maderaspatana* (L.) M.Roem. (Cucurbitaceae) – A traditional medicinal plant in western districts of Tamil Nadu, India. *Asian Journal of Pharmaceutical and Clinical Research*. 5(4): 163-168.
- Payum, T. (2020). Phytoconstituents and Proximate Composition of *Clerodendrum colebrookianum* Walp: A Widely Used Anti High Blood Pressure Medicinal Food Plant in Eastern Himalayas. *Pharmacognosy Journal*. 12(6): 1534-1540.
- Perez-Jimenez, J., Arranz, S., Tabernero, M., Díaz-Rubio, M.E., Serrano, J., Goñi, I. and Saura-Calixto, F. (2008). Updated methodology to determine antioxidant capacity in plant foods, oils and beverages: extraction, measurement and expression of results. *Food Research International*. 41(3): 274- 285.
- Peterson, J., Dwyer, J., Bhagwat, S., Haylowitz, D., Hoiden, J., Eldridge, A.L., Beecher, G. and Ala-Desamni, J. (2005). Major flavonoids in dry tea. *Journal of Food Composition and Analysis*. 18(6): 487-501.
- Phillips, R., Bibby, M. and Double, J. (1990). A critical appraisal of the predictive value of *in vitro* chemosensitivity assays. *Journal of the National Cancer Institute*. 82: 1457-68.
- Phillipson, J.D. (2001). Phytochemistry and medicinal plants. *Phytochemistry*. 56 (3): 237-243.
- Phongtongpasuk, S. and Poadang, S. (2014). Extraction of antioxidants from *Peperomia pellucida* L. Kunth. *Thammasat International Journal of Science and Technology*. 19(3): 38-43.
- Pietta, P.G. (2000). Flavonoids as antioxidants. *Journal of Natural Products*. 63(7): 1035-1042.
- Pioani, J.R. (1993). A new species and a new combination in *Zanthoxylum* (Rutaceae) from Brazil. *Brittonia*. 45: 154-158.
- Pourramezana, Z., Oloomib, M. and Kermanshahi, R.K. (2022). Effect of solvent polarity on the extraction yield of antioxidants from *Lactobacillus* Supernatants. *Iranian Journal of Pharmaceutical Sciences*. 18 (1): 47-54.
- Prajapati, N. D. Purohit, S. S. Sharma, A. K. and Kumar. T. (2003). A Handbook of Medicinal Plants Agrobios Publisher, Jodhpur, India.

- Prakash, N.K.U., Sripriya, N.S., Raj, D.D., Deepa, S. and Bhuvaneswari, S. (2019). Antioxidant potency and GC-MS composition of *Origanum majorana* Linn. *Pakistan Journal of Pharmaceutical Science*. 32(5): 2117-2122.
- Prarthana, J. and Maruthi, K.R. (2017). Screening of phytochemicals & bioactive antibacterial activity in *Spirogyra* Sp. *International Journal of Advanced Research*. 5(7): 1145-1154.
- Prasad, K. (2017). HPLC Analysis of amino acid and antioxidant composition of three medicinal plants of (Pithoragarh) Uttarakhand Himalayas. *Journal of Analytical and Pharmaceutical Research*. 6(5): 1-6.
- Prasad, S.B., Rosangkima, G. and Rongpi, T. (2010). Ethno-medicinal value of some anticancer medicinal plants from north-east India: an in vivo screening in murine tumor model. *Scientific Visualization*. 10(4): 123-132.
- Prejeena, V., Suresh, S.N. and Varsha, V. (2017). Phytochemical screening, antioxidant analysis and antiproliferative effect of *Costus pictus* d. don leaf extracts. *International Journal of Recent Advances in Multidisciplinary Research*. 4(3): 2373-2373.
- Pukhrambam, P.D., Devi, K.K. and Das, S. (2023). Molecular Identification of Medicinal Plant, *Polygonum posumbu* from Manipur, India. *Bioscience Biotechnology Research Communications*. 6(1): 2321-4007.
- Quadri, O. N., and Musa, T. Y. (2015). Phytochemical Composition, Mineral Contents and Amino Acid Profile of *Phyllanthus amarus* leaves.
- Rabeta, M.S., Mousavi, L. and Murugaiyah, V. (2019). Nutritional and antinutritional values of leaves and stems of *Ocimum tenuiflorum* L. *Food Research*. 3 (6): 798-807.
- Raclariu, A. C., Mocan, A., Popa, M. O., Vlase, L., Ichim, M. C., Crisan, G., Brysting A. K. and de Boer, H. (2017). *Veronica officinalis* product authentication using DNA metabarcoding and HPLC-MS reveals widespread adulteration with *Veronica chamaedrys*. *Frontiers in Pharmacology*. 8: 378. <https://doi.org/10.3389/fphar.2017.00378>.
- Radhakrishnan, S., Alarfaj, A. A. and Annadurai, G. (2015). Estimation of phytochemical analysis and invitro antioxidant activity of *calotropis gigantea* extract: wound healing activity and its biomedical application. *International Journal of Pharmaceutical and Sciences and Research*. 6(7): 3053-3060.

- Rahimi, R. and Rabani, M. (2010). Mineral content of some plants used in Iran. *Pharmacognosy Research*. 2(4): 267-70.
- Rahman, M.M., Islam, M.B., Biswas, M. and Alam, A.H.M.K. (2015). In vitro antioxidant and free radical scavenging activity of different parts of *Tabebuia pallida* growing in Bangladesh. *BMC Research Notes*. 8(621): 1-9.
- Rahuman, A.A., Gopalakrishnan, G., Ghouse, G., Arumugam, S. and Himalayan, B. (2000). Effect of *Feronia limonia* on mosquito larvae. *Fitoterapia*. 71(5): 553-555.
- Rai, P.K. and Lalramnghinglova, H. (2010). Ethnomedicinal plant resources of Mizoram, India. Implication of traditional knowledge in health care system. *Ethnobotanical Leaflets*. 10(3): 274-305.
- Rajalakshmi, K. and Mohan, V.R. (2016). GC-MS analysis of bioactive components of *Myxopyrum serratum* A.W. Hill (Oleaceae). *International Journal of Pharmaceutical Sciences Review and Research*. 38(1): 30-35.
- Rajiv, P., Deepa, A., Vanathi, P. and Vidhya, D. (2017). Screening for phytochemicals and FTIR analysis of *Myristica dactyloids* fruit extracts. *International Journal of Pharmacy and Pharmaceutical Sciences*. 9(1): 0975-1491.
- Rakholiya, K., Kaneria, M. and Chanda, S. (2011). Vegetable and fruit peels as a novel source of antioxidants. *Journal of Medicinal Plants Research*. 5(1): 63-71.
- Ralte, L., Kiangte, L., Thangjam, N.M., Kumar, A. and Singh, Y.T. (2022). GCMS and molecular docking analyses of phytochemicals from the underutilized plant, *Parkiatimoriana* revealed candidate anti-cancerous and anti-inflammatory agents. *Scientific Research Journal*. 12(1): 3395.
- Ram, V.R., Aadesariya, M.K. and Vyas, S.J. (2018). Study of Bioactive Components of n-Butanol by Gas Chromatography-Mass Spectrometry (GC-MS) In Leaves of *Grewia tenax* growing in Kachchh District. *World Journal of Pharmaceutical Research*. 7(1): 607-625.
- Ramalakshmi, S. and Muthuchelian, K. (2011). Analysis of Bioactive constituents from the Ethanolic leaf extract of *Tabebuia rosea* (Bertol.) DC by Gas Chromatography-Mass Spectrometry. *International Journal of ChemTech Research*. 3(3): 1054-1059.
- Ramalingam, S., Selvaraj, D., Shanmughanandhan, D., Sarma, D.K., Joseph, J.C. and Srinivasan, R.V. (2012). DNA Barcode ITS effectively distinguishes the medicinal plant *Boerhavia diffusa* from Its Adulterants. *Genomics Proteomics Bioinformatics*. 10: 364-367.

- Raman, B.V., Samuel, L., Pardha, S.M., Narashimha, R.B., Krishna, A.N.V. and Radhakrishnan, T.M. (2012). Antibacterial, antioxidant activity and GC-MS analysis of *Eupatorium odoratum*. *Asian Journal of Pharmaceutical and Clinical Research*. 5(2): 99-106.
- Ramanibai, R., Parthiban, E., Arokiyaraj, C. and Janarthanan, S. (2019). Antioxidant and GC-MS analysis of *Annona reticulata* leaves extract against unsecure free radicals. *SN Applied Sciences*. 1: 349.
- Ramya, B., Malarvili, T. and Velavan, S. (2015). GCMS of bioactive compounds in *Bryonopsis laciniata* fruit extract. *International Journal of Pharmaceutical Science and Research*. 6(8): 3375-3379.
- Ramesh, H.L. and Yogananda, M.V.N. (2012). Assessment of heavy metals contamination in green leafy vegetables grown in Bangalore urban district of Karnataka. *Advances in Life Science and Technology*. 6: 40-51.
- Ramya, B., Seetharaman, S., Indra, V., Sheela, D., Perinbam, K. and Augustain, G. (2017). Phytochemical profiling of aqueous extracts of selected medicinal plants. *Journal of Pharmaceutical and Scientific Innovation*. 6 (1): 2277- 4572.
- Ranganathan, D. (2014). Phytochemical analysis of *Caralluma nilagiriana* using GC-MS. *Journal of Pharmacognosy and Phytochemistry*. 3 (1): 155-159.
- Rao, C.V., Newmark, H.L. and Reddy, B.S. (1998). Chemopreventive effect of squalene on colon cancer. *Carcino genesis*. 19(2): 287-290.
- Rao, K.V.K., Schwartz, S.A., Nair, H.K., Aalinkeel, R., Mahajan, S., Chawda, R. and Nair, M.P.N. (2004). Plant derived products as a source of cellular growth inhibitory phytochemicals on PC-3M, DU-145 and LNCaP prostate cancer cell lines. *Current Science*. 87(11): 1585-1588.
- Rao, P.K., Hasan, S.S., Bhellum, B.L. and Manhas, R.K. (2015). Ethnomedicinal plants of Kathua district, Jammu and Kashmir, India. *Journal of Ethnopharmacology*. 2(171): 12-27.
- Razika, L., Thanina, A.C., Nadjiba, C.M., Narimen, B., Mahdi, D.M. and Arab, K. (2017). Antioxidant and wound healing potential of saponins extracted from the leaves of Algerian *Urtica dioica* L. *Pakistan Journal of Pharmaceutical Science*. 30(3): 1023-1029.
- Reddy, D.N. (2019). Essential oils extracted from medicinal plants and their applications. DOI: 10.1007/978-981-13-7154-7-9.

- Renvoize, C., Biola, A., Pallardy, M. and Breard, J. (1998). Apoptosis: Identification of dying cells. *Cell Biology and Toxicology*. 14(2): 111-20.
- Reyes-Garcia, V., Kightley, E., Ruiz-Mallen, I., Fuentes-Pelaez, N., Demps, K., Huanca, T. and Martinez-Rodriguez, M. (2010). Schooling and local environmental knowledge: Do they complement or substitute each other? *International Journal of Educational Development*. 30(3): 305-313.
- Rice-Evans, C., Sampson, J., Bramley, P.M. and Holloway, D.E. (1997). Why do we expect carotenoids to be antioxidants in vivo. *Free Radical Research*. 26(4): 381-398.
- Rimm, E.B., Appel, L.J., Chiuve, S.E., Djousse, L., Engler, M.B., Kris-Etherton, P.M., Mozaffarian, D., Siscovick, D.S. and Lichtenstein A.H. (2018). Seafood long-chain n-3 polyunsaturated fatty acids and cardiovascular disease: A science advisory from the American Heart Association. *Circulation*. 138(1): 35-47.
- Rogero, M.M. and Calder, P.C. (2018). Obesity, inflammation, toll like receptor 4 and fatty acids. *Nutrients*. 10(4): 432.10.3390/nu10040432.
- Romero-Diez, R., Rodriguez-Rojo, S., Cocero, M.J., Duarte, C.M.M., Matias, A.A. and Bronze, M.R. (2018). Phenolic characterization of aging wine lees; Correlation with antioxidant activities. *Food Chemistry*. 1(259): 188-195.
- Rosas-Rodriguez, J.A., Moran-Palacio¹, E.F., Tortoledo-Ortiz, O., Yanez-Farias, G.A., Zamora-Alvarez, L.A., Stephens-Camacho, N.A., Sonanez-Organis, J.G. and Ochoa-Lopez, L.M. (2014). Determination of Amino Acids in Medicinal Plants from Southern Sonora, Mexico. *Tropical Journal of Pharmaceutical Research*. 13 (4): 601-606.
- Roy, M.K., Swargiary, A. and Verma, A.K. (2021). Antiproliferative and apoptosis-inducing properties of selected medicinal plants of Assam, India. *Archives of Medicine and Health Sciences*. 9(2): 236-43.
- Rubilar, O., Benavides-Mendoza, A., Diez, M.C., Tortella, G., Cornejo, P., Bustamante, L., Quiroz, A., Ruiz, A., Nahuelcura, J., Berrios, D., Vera, J., Espinoza, J. and Fincheira, P. (2023). The Impact of 2-Ketones Released from Solid Lipid Nanoparticles on Growth Modulation and Antioxidant System of *Lactuca sativa*. *Plants*. 12(7): 3094-1-19.
- Sadeghi, Z. and Mahmood, A. (2014). Ethno-gynecological knowledge of medicinal plants used by Baluch tribes, southeast of Baluchistan. *Revista Brasileira de farmacognosia*. 24(6): 706-715.

- Saeed, N., Khan, M.R. and Shabbir, M. (2012). Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complementary Alternative Medicine*. 12(221): 1-12.
- Saha, D., Talukdar, A., Das, T., Ghosh, S.K. and Rahman, H. (2012). Evaluation of analgesic activity of ethanolic extract of *Cleodendrum serratum* Linn leaves in Rats. *International Research Journal of Pharmaceutical and Applied Science*. 2(6): 33-37.
- Sahoo, U.K. and Rocky, P. (2019). Proximate constituents of some homegarden food crops and household food security in Mizoram, northeast India. *Indian Journal of Hill Farming*. 32(1): 71-78.
- Saikia, A.P., Ryakala, V.K., Sharma, P., Goswami, P. and Bora, U. (2006). Ethnobotany of medicinal plants used by Assamese people for various skin ailments and cosmetics. *Journal of Ethnopharmacology*. 106(2): 149-157.
- Saikia, B. (2006). Ethnomedicinal plants from Gohpur of Sonitpur district, Assam. *Indian Journal of Traditional Knowledge*. 5(4): 529-530.
- Saikia, K., Lalawmpuii, R. and Borgohain, P. (2017). Evaluation of In-Vitro antioxidant and cytotoxic activity of methanolic leaf extract of *Blumea lanceolaria* Roxb. *International Journal of Technical Research and Applications*. 5(6): 54-57.
- Saikia, P. and Deka, D.C. (2013). Mineral content of some wild green leafy vegetables of North-East India. *Journal of Chemical and Pharmaceutical Research*. 5(3): 117-21.
- Saikia, S.P., Wann, S.B. and Gogoi, B. (2020). DNA Barcodes for delineating *Clerodendrum* species of North East India. *Scientific Reports*. <https://doi.org/10.1038/s41598-020-70405-3>.
- Sajib, M.A.M., Jahan, S., Islam, M.Z., Khan, T.A. and Saha, B.K. (2014). Nutritional evaluation and heavy metals content of selected tropical fruits in Bangladesh. *International Food Research Journal*. 21(2): 609-615.
- Sak, K. (2014). Cytotoxicity of dietary flavonoids on different human cancers types. *Pharmacognosy Review*. 8(8): 122-146.
- Salama, A.K. and Radwan, M.A. (2005). Heavy metals (Cd, Pb) and trace elements (Cu, Zn) contents in some foodstuffs from the Egyptian market. *Emirates Journal of Agriculture Science*. 17(1): 34-42.

- Samarghandian, S., Hadjzadeh, M.A.R., Afshari, J.T. and Hosseini, M. (2014). Antiproliferative activity and induction of apoptotic by ethanolic extract of *Alpinia galanga* rhizome in human breast carcinoma cell line. *BMC Complementary and Alternative Medicine*. 14: 192.
- Samell, K., Sonita, L., Chanseiha, N., Sokhany, L. and Kosalvorlak, C. (2018). Phytochemical analysis and antioxidant property of selected medicinal plants native to Cambodia. *Drug Designing and International Properties International Journal*. 1(2): 21-29.
- Sangma, A.J.T. and Sahoo, U.K. (2017). Utilization Pattern of Medicinal Plants by Different Tribes of Garo Hills of Meghalaya, North-East India. *American Journal of Ethnomedicine*. 4(1): 1-8.
- Sanjeevkumar, C.B., Londonkar, R.L., Kattagouda, U.M. and N. K. Asha Tukappa. (2016). Screening of in vitro antioxidant activity of chloroform extracts of *Bryonopsis laciniosa* Fruits. *International Journal of Current Microbiology and Applied Sciences*. 5(3): 590-597.
- Sar, S.K., Nair, L.D., Arora, A. and Mahapatra, D. (2013). Fourier Transform Infrared Spectroscopy Analysis of Few Medicinal Plants of Chhattisgarh, India. *Journal of Advanced Pharmacy Education & Research*. 3(3): 196-200.
- Satpathy, G., Tyagi, Y.K. and Gupta, R.K. (2011). Preliminary evaluation of nutraceutical and therapeutic potential of raw *Spondias pinnata* K. an exotic fruit of India. *Food Research International*. 44(7): 2076- 2087.
- Satter, M.M.A., Mohammed, M.R.L.K., Syeda, J.A., Nusrat, A., Mohammed, F.I. and Shaha, B. (2016). Nutritional quality and safety aspects of wild vegetables consume in Bangladesh. *Asian Pacific Journal of Tropical Biomedicine*. 6(2): 125-131.
- Satyral, P., Dosoky, N.S., Poudol, A. and Setzer, W.N. (2012). Essential oil constituents and their biological activities from the leaves of *Cassia fistula* growing in Nepal. *Open Access Journal of Medical and Aromatic Plants*. 3(2): 1-4.
- Saupi, N., Zakaria, M.H. and Bujang, J.S. (2009). Analytic chemical composition and mineral content of yellow velvet leaf (*Limnocharis flava* L. Buchenau)'s edible parts. *Journal of Applied Science*. 9(16): 2969-2974.
- Savithramma, N., Ankanna, S., Rao, M.L. and Saradvathi, J. (2012). Studies on antimicrobial efficacy of medicinal tuberous shrub *Talinum cuneifolium*. *Journal of Environmental Biology*. 33(4): 775-780.

- Savithamma, N., Sulochana, C. and Rao, K.N. (2007). Ethnobotanical survey of plants used to treat asthma in Andhra Pradesh, India. *Journal of Ethnopharmacology*. 113(1): 54-61.
- Sawmliana, M. The Book of Mizoram Plants. 1st Ed. Lois Bet, Chandmari, Aizawl. 2003: pp. 114.
- Schultz, J., Maisel, S., Gerlach, D., Muller, T. and Wolf, M. (2005). A common core of secondary structure of the internal transcribed spacer 2 (ITS2) throughout the Eukaryota. *RNA*. 11(4): 361-364.
- Seal, T. (2020). Effect of boiling and microwave cooking on nutritional, anti-nutritional and toxicity of wild edible plants of north-eastern region in India. *International Journal of Pharmacy and Pharmaceutical Sciences*. 12(11): 61-70.
- Seal, T. and Chaudhuri, K. (2015). Effect of solvent extraction system on the antioxidant activity of some selected wild leafy vegetables of Meghalaya state in India. *International Journal of Pharmaceutical and Phytopharmacological Research*. 5(3): 41-47.
- Seal, T. and Chaudhuri, K. (2016). Nutritional analysis of some selected wild edible plants consumed by the tribal people of Meghalaya state in India. *International Journal of Food Science and Nutrition*. 1(6): 39-43.
- Seal, T., Pillai, B. and Chaudhuri, K. (2017). Evaluation of Nutritional Potential of Five Unexplored Wild Edible Plants Consumed by the Tribal People of Arunachal Pradesh State in India. *Journal of Food and Nutrition Research*. 5(1): 1-5.
- Sears, B. and Perry, M. (2015). The role of fatty acids in insulin resistance. *Lipids in Health and Diseases*. 14: 121. 10.1186/s12944-015-0123-1.
- Sermakkani, M. and Thangapandian, V. (2011). GC-MS analysis of Cassia italica leaf methanol extract. *Asian Journal of Pharmaceutical and Clinical Research*. 5(2): 90-94.
- Shaayau, S. and Kurya, A.U. (2019). Nutritional, antinutritional and therapeutic potentials of *Solanum incanum* Linnaeus fruit cultivated in Sokoto, Northern Nigeria. *Nigerian Journal Pure & Applied Science*. 32(2): 3461-3468.
- Shah, R.K. (2016). Preliminary Nutritional analysis and Phytochemistry of *Smilax ovalifolia* Leaves. *International Journal of Current Microbiology and Applied Sciences*. 5(1): 270-273.
- Shah, T.K., Kumar, A., Tandel, R., Bhat, R.A.H. and Sarma, D. (2019). Identification of functional groups in *Thymus linearis* ethanol leaf extract through

fourier transform infrared spectroscopy (FTIR). *Journal of Pharmacognosy and Phytochemistry*. 8(4): 3013-3015.

Shalaby, E.A., El-fayoumy, E.A., Shanab, S.M.M., Gaballa, H.S. and Mohamed, A.T. (2021). Evaluation of antioxidant and anticancer activity of crude extract and different fractions of *Chlorella vulgaris* axenic culture grown under various concentrations of copper ions. *BMC Complementary Medicine and Therapies*. 21(51): 1-16.

Sharanabasappa, M., Santosh, D., Shaila, Y., Seetharam, and Sanjeevarao, I. (2007). Phytochemical studies on *Bauhinia racemosa* Lam. *Bauhinia purpurea* tinn. and *Hardwickia binata* Roxb. *E- Journal of Chemistry*. 4(1): 21-31.

Shareef, M.I., Leelavathi, S., Gopinath, S. and Shareef, M. (2013). Evaluation of in-vivo Activity of *Clerodendrum serratum* L. against Rheumatism. *International Journal of Innovative Research in Science, Engineering and Technology*. 2(12): 7750-58.

Sharma, C. and Chetri, A. (2017). Documentation of traditional medicinal plants used By Garo Tribe in Dimoria Tribal belt of Assam. *International Conference on Innovative Research in Science, Technology and Management*. 44-48.

Sharma, H. and Shrivastava, A. (2022). Antimicrobial and antioxidative properties of *Cleome viscosa* leaf extract. *Research Square*. 1-10.

Shedzad, A., Qayyum, A., Rehman, R., Nadeem, F. and Muhammed, R.S. (2018). A review of bioactivity guided medicinal uses and therapeutic potentials of noxious weed (*Alternanthera sessilis*). *International Journal of Chemical and Biochemical Sciences*. 14: 95-103.

Shettar, A.K., Vedamurthy, A.B., Sateesh, M.K. and Kaliwal, B.B. (2017). In vitro antidiabetic activities and GC-MS phytochemical analysis of *Ximenia americana* extracts. *South African Journal of Botany*. 113(108): 202-211.

Shodehinde, S.A., Adefegha, S.A., Oboh, G., Oyeleye, S.I., Olasehinde, T.A., Nwanna, E.E., Adedayo, B.C. and Boligon, A.A. (2016). Phenolic composition and evaluation of methanol and aqueous extracts of bitter melon (*Momordica charantia* L) leaves on Angiotensin-I- Converting Enzyme and some pro- oxidant- induced lipid peroxidation in vitro. *Journal of Evidence-Based Complementary & Alternative Medicine*. 21(4): 67-76.

Shukla, R.K., Painuly, D., Porval, A. and Shukla, A. (2014). Proximate analysis, nutritional value, phytochemical evaluation, and biological activity of *Litchi chinensis* Sonn. leaves. *Journal of Herbs, Spices & Medicinal Plants*. 20: 196-208.

- Shyam, P. and Suresh, P.K. (2013). Comparative analysis of three leaf extracts of *Ixora coccinea* Linn. for their protective and antioxidant potentials and correlation with analytical data. *International Journal of Pharma and BioSciences*. 4(4): 937-949.
- Sibiya, N.P., Moteetee, A.N. and Kayitesi, E. (2021). Proximate Analyses and Amino Acid Composition of Selected Wild Indigenous Fruits of Southern Africa. *Plants*. 10(4): 721. <https://doi.org/10.3390/plants10040721>.
- Siddhuraju, P. and Becker, K. (2003). Antioxidant properties of various extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera* lam.) leaves. *Journal of Agricultural and Food Chemistry*. 51(8): 2144-2155.
- Simbo, D.J. (2010). An ethnobotanical survey of medicinal plants in Babungo, Northwest Region, Cameroon. *Journal of Ethnobiology and Ethnomedicine*. 6(8):1-7.
- Singh J., Singh, A. and Praves, K.R. (2003). Proceedings of the 1st National Interactive Meet on Medicinal and Aromatic Plants. CIMAP; Lucknow, India: Production and trade potential of some important medicinal plants: An overview.
- Singh, J. and Basu, P. S. (2012). Non-nutritive bioactive compounds in pulses and their impact on human health: an overview. *Food and Nutrition Science*. 3(12):1664-1672.
- Singh, M. K., Khare, G., Iyer, S.K., Sharwan, G. and Tripathi, D.K. (2012). *Clerodendrum serratum*: A Clinical approach. *Journal of Applied Pharmaceutical Science*. 2: 11-15.
- Singh, P.K., Singh, J., Medhi, T. and Kumar, A. (2022). Phytochemical Screening, Quantification, FT-IR Analysis and In Silico Characterization of Potential Bioactive Compounds Identified in HR-LC/MS Analysis of the polyherbal formulation from Northeast India. *ACS Omega*. 7: 33067- 33078.
- Singh, P.K., Sonter, S., Mishra, S. and Dwivedi, M.K. (2021). Chemical profiling, in vitro antioxidant, membrane stabilizing and antimicrobial properties of wild growing *Murraya paniculata* from Amarkantak (M.P.) *Scientific Reports*. 11: 9691.
- Singh, R.C., Nelson, R., Muthu, K.P. and Pargavi, B. (2011). Identification of Volatile Constituents from *Premna serratifolia* L. through GC-MS. *International Journal of PharmTech Research*. 3(2): 1050-1058.

- Singh, R.K. and Negi, A.K. (2019). Traditional Knowledge and Use of Medicinal Plants A Case Study from Joshimath Block of Chamoli Uttarakhand. *Journal of Pharmacognosy and Phytochemistry*. 8(3): 122-127.
- Singha, B.L. and Hassan, Y. (2017). Analysis of carbohydrate, moisture content and specific gravity of bambusa tulda with special reference to its harvesting season. *International Journal of Science and Research*. 6(1): 279-283.
- Sinha, M.K. and Sinha, D. (2013). Herbal medicinal plants of Kororia district (C.G.) India. *International Journal of Green and Herbal Chemistry*. 2(4): 1114-20.
- Sinha, R. (2018). Nutritional Analysis of Few Selected Wild Edible Leafy Vegetables of Tribal of Jharkhand, India. *International Journal of Current Microbiology Applied Science*. 7(2): 1323-1329.
- Sinha, S. and Raghuwansh, R. (2021). Evaluation of Phytochemical, Antioxidant and Reducing Activity in Whole Plant Extract of *Andrographis paniculata*. Pharmaceutical Communication. *Bioscience Biotechnology Research Communications*. 14(1): 282-290.
- Skoog, A., Holler, E.J. and Crouch, S.R. (2007). Principles of instrumental analysis. 6 Edition, Brooks Cole, Belmont. 1039.
- Snively, G. and Corsiglia, J. (2000). Discovering indigenous science: Implications for science education. *Science Education*. 85(1): 6-34.
- Soad, M., El-din, M. and Amani, M.D. (2016). Bioactivity and phytochemical constituents of marine red seaweeds (*Jania rubens*, *Corallina mediterranea* and *Pterocladia capillacea*). *Journal of Taibah University for Science*. 10(4): 471- 484.
- Sobuj, M.K.A., Islam, M.A., Islam, M.S., Islam, M.M., Mahmud, Y. and Rafiquzzaman, S.M. (2021). Effect of solvents on bioactive compounds and antioxidant activity of *Padina tetrastrum* and *Gracilaria tenuistipitata* seaweeds collected from Bangladesh. *Scientific Reports*. 11(1): 19092.
- Sodamide A., Ayankoso A. S., Odoje O. F., and Owonikoko A.D. (2021). Nutritive evaluation, mineral composition, amino acid profile and phytochemical analysis of leaf protein concentrate of Bitter Gourd (*Mormodica charantia*). *European Journal of Material Science*. 8(2): 29-40.
- Solowey, E., Lichtenstein, M., Sallon, S., Paavilainen, H., Solowey, E. and Lorberboum-Galski, H. (2014). Evaluating medicinal plants for anticancer activity. *Scientific World Journal*. 1-12.

- Somashekar, G., Sudhakar, U., Srividya, S. and Suresh, S. (2023). Phytochemical Analysis and in vitro Cell Viability Effects of Ethanolic Extract of *Ormocarpum cochinchinense* on Mouse Embryonic Fibroblasts. *Indian Journal of Pharmaceutical Education and Research*. 57(1): 120-124.
- Soosairaj, S. and Dons, T. (2016). Bio-active compounds analysis and characterization in Ethanolic plant extracts of *Justicia tranquebariensis* L. (Acanthaceae)-using GC-MS. *International Journal of ChemTech Research*. 9(7): 260-265.
- Soriana-Santos, J., Gonzalez-Palma, I., Escalona-Buendia, H.B., Ponce-Alquicira, E., Tellez-Tellez, M., Gupta, V.K. and Diaz-Godinez, G. (2016). Evaluation of the antioxidant activity of aqueous and methanol extracts of *Pleurotus ostreatus* in different growth stages. *Frontiers in Microbiology*. 7: 1-9.
- Sowmiya, R., Balasubramani, G., Deepak, P., Aiswarya, D., Ravikumar, S. and Perumal, P. (2017). Characterization and screening of in vitro antimalarial and larvicidal activities of selected seaweeds from southeast coast of India against *Plasmodium falciparum* and *Anopheles stephensi*. *Journal of Coastal Life Medicine*. 5(6):242-248.
- Spanova, M. and Drum, G. (2011). Squalene-biochemistry, molecular biology, press biotechnology and applications. *European Journal of Lipid Science and Technology*. 113(11): 1299-1320.
- Sreeja, P.S., Arunachalam, K., Saikumar, S., Kasipandi, M., Dhivya, S., Murugan, R. and Parimelazhagan, T. (2018). Gastroprotective effect and mode of action of methanol extract of *Sphenodesme involucrata* var. *paniculata* (C.B. Clarke) Munir (Lamiaceae) leaves on experimental gastric ulcer models. *Biomedicine & Pharmacotherapy*. 97: 1109-1118.
- Staden, V.J., Finnie, J.F., Luterova, A., Gruz, J., Chukwujekwu, J.C., Aremu, A.O. and Vambe, M.M. (2020). Antibacterial, Mutagenic Properties and Chemical Characterisation of Sugar Bush (*Protea caffra* Meisn.): A South African Native Shrub Species. *Plants*. 9(1331): 1-12.
- Stankovic, M.S., Niciforovic, N., Topuzovic, M. and Solujic, S. (2014). Total phenolic content, flavonoid concentrations and antioxidant activity of the whole plant and plant parts extracts from *Teucrium montanum* L. var. *Montanum*, F. *Supinum* (L.) Reichenb. *Biotechnology & Biotechnological Equipment*. 25(1): 1310-2818.

- Starlin, T., Prabha, P.S., Thayakumar, B.K.A. and Gopalakrishnan, V.K. (2019). Screening and GC-MS profiling of ethanolic extract of *Tylophora pauciflora*. *Journal of Biomedical Informatics*. 15(6): 425-429.
- Starr, J.R., Naczi, R.F. and Chouinard, B.N. (2009). Plant DNA barcodes and species resolution in sedges (*Carex*, Cyperaceae). *Molecular ecology resources*. 9(6): 151-163.
- Steane, D.A., Scotland, R.W., Mabberley, D.J., Wagstaff, S.J., Reeves, P.A. and Olmstead, R.G. (1997). Phylogenetic relationships of *Clerodendrum serratum* L. (Lamiaceae) Inferred from chloroplast DNA. *Systematic Botany*. 22(2): 229-243.
- Stevenson, M.G. (1996). Indigenous knowledge in environmental assessment. *Arctic*. 49(3): 278-291.
- Stobiecki, M. and Kachlicki, P. (2006) In: The Science of Flavonoids (E. Grotewold, Ed.), Springer Science+Business Media, New York. 47-69.
- Stoddart, L.A., Smith, N.J. and Milligan, G. (2008). Free fatty acid receptors FFA1, -2, and -3: pharmacology and pathophysiological functions. *Pharmacological Reviews*. 60(4): 405-417.
- Subramanian, L., Thomas, S. and Koshy, O. (2017). Green Synthesis of Silver Nanoparticles using Aqueous Plant Extracts and its Application as Optical Sensor. *International Journal of Biosensors & Bioelectronics*. 2(3): 1-4.
- Sudha, S.S., Devakumar, J. and Keerthana, V. (2017). Identification of Bioactive Compounds by Gas Chromatography-Mass Spectrometry Analysis of *Syzygium Jambos* (L.) Collected From Western Ghats Region Coimbatore, Tamil Nadu. *Asian Journal of Pharmaceutical and Clinical Research*. 10(1): 364-369.
- Sulaiman, C.T. and Indira, B. (2012). Total phenolics and total flavonoids in selected Indian medicinal plants. 74(3): 258-260.
- Sultana, B., Anwar, F. and Przybylski, R. (2007). Antioxidant activity of phenolic components present in barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. *Food Chemistry*. 104(3): 1106-1114.
- Sun, S., Bleck, R., Benjamin, S.G., Green, B.W. and Grell, G.A. (2018). Sub seasonal forecasting with an icosahedral, vertically quasi-Lagrangian coupled model. Part I: Model overview and evaluation of systematic errors. *Weather Review* 146(5): 1601-1617.

- Surewicz, W.K., Mantsch, H.H. and Chapman, D. (1993). Determination of Protein Secondary Structure by Fourier Transform Infrared Spectroscopy: A Critical Assessment. *Biochemistry*. 32(2): 389-393.
- Suriyavathana, M., Subha Palanisamy, S.S., Selvaraj, S., Mari, K.R. and Kuppulingam, R. (2016). Phytochemical Profile of *Erythrina variegata* by Using High-Performance Liquid Chromatography and Gas Chromatography-Mass Spectroscopy Analyses. *Journal of Acupuncture and Meridian Studies*. 9(4): 207-212.
- Swamy, M.K., Sinniah, U.R. and Akhtar, M.S. (2015). In Vitro Pharmacological Activities and GC-MS Analysis of Different Solvent Extracts of *Lantana camara* Leaves Collected from Tropical Region of Malaysia. *Evidence-Based Complementary and Alternative Medicine*. 1-9.
- Swargiary, A., Daimari, M., Roy, M., Haloi, D. and Ramchiary, B. (2019). Evaluation of phytochemical properties and larvicidal activities of *Cynodon dactylon*, *Clerodendrum viscosum*, *Spilanthes acmella* and *Terminalia chebula* against *Aedes aegypti*. *Asian Pacific Journal of Tropical Medicine*. 12(5): 224-231.
- Swargiary, N., Dey, T. and Das, A.N. (2019). A Survey on Traditionally Used Medicinal Plants of Rangjuli (Belpara), Goalpara, Assam. *Periodic Research*. 7(4): 87-96.
- Syad, A.N., Shunmugiah, K.P. and Kasi, P.D. (2013). Antioxidant and anti-cholinesterase activity of *Sargassum wightii*. *Pharmaceutical Biology*. 51(11): 1401-1410.
- Taberlet, P., Coissac, E., Pompanon, F., Gielly, L., Miquel, C., Valentini, A., Vermat, T., Corthier, G., Brochmann, C. and Willerslev, E. (2007). Power and limitations of the chloroplast trnL (UAA) intron for plant DNA barcoding. *Nucleic Acids Research*. 35 (3): 1-8.
- Tahir, A., Hussain, F., Ahmed, N., Ghorbani, A. and Jami, A. (2018). Assessing universality of DNA barcoding in geographically isolated selected desert medicinal species of Fabaceae and Poaceae. *Peer Journal*. 6:e4499. Doi 10.7717/peerj.4499.
- Tanti, B., Roy, S.J., Baruah, P.S., Lahkar, L., Gurung, L. and Saikia, D. (2019). Phytochemical analysis and antioxidant activities of *Homalomena aromatica* Schott. *Journal of Pharmacognosy and Phytochemistry*. 8(1): 1379-1385.
- Taroq, A., Kamari, F.E., Aouam, I., Atki, Y.E., Lyoussi, B. and Abdellaoui, A. (2018). Antioxidant activities and total phenolic and flavonoid content variations of

leaf extracts of *laurus nobilis* L. from morocco. *Asian Journal of Pharmaceutical and Clinical Research*. 11(12): 540-543.

Techen, N., Parveen, I., Pan, Z. and Khan, I.A. (2014). DNA Bar-coding of medicinal plant material for identification. *Current Opinion in Biotechnology*. 30(25): 103-110.

Terto, M.V.C., Gomes, J.M., Araujo, D., Silva, T.S., Ferreira, J.M., Souza, J.J.N., Silva, M.S. and Tavares, J. (2020). Photoprotective activity of *Plectranthus amboinicus* extracts and HPLC quantification of rosmarinic acid. *Revista Brasileira de Farmacognosia*. 30(4): 183-8.

Teruel, M.R., Garrido, M.D., Espinosa, M.C. and Linares, M.B. (2015). Effect of different format solvent rosemary extracts (*Rosmarinus officinalis*) on chicken nuggets quality. *Food Chemistry*. 172: 40- 46.

Thakur, K., Bassi, K. and Sood, K. (2015). Textbook of Ethnobotany; S. Dinesh & Co: Jalandhar, India, pp. 1-70.

Thakuria, P., Nath, R., Sarma, S., Kalita, D.J., Dutta, D.J., Borah, P., Sharma, R., Barman, C. and Hussain, J. (2018). Phytochemical screening of medicinal plants occurring in local area of Assam. *Journal of Pharmacognosy and Phytochemistry*. 7(3): 186-188.

Tharmabalan, R.T. (2021). Nutritional analysis of five wild edible vegetables traditionally consumed by the Orang Asli in Perak. *International Journal of Food Science*.1-7.

Theja, B. and Kuber, R. (2023). Phytochemical screening and Fourier Transform IR Analysis of *Ficus sagittifolia* (Warburg Ex Mildbread and Burret) stem bark. *International Journal of of Pharmacy Research & Technology*. 13(1): 73-78.

Thomas, C. (2009). Plant barcode soon to become reality. *Science*. 325:526.

Thompson, J.D., Gibson, T.J., Plewnaik, F., Jeanmougin, F. and Higgins, D.G. (1997). The Clustal X Windows Interfare: Flexible Strategies for multiple sequence alignment aided by Quality Analysis Tools. *Nucleic Acid Research*. 25(24): 4876-4882.

Thushar, S., Mohammad, S. and Singh, R.K. (2023). Proximate Composition and Nutritional Values of Selected Wild Plants of the United Arab Emirates. *Molecules*. 28(3): 1504. doi.org/10.3390/molecules28031504.

- Tiwari, P., Kumar, B., Kaur, M., Kaur, G. and Kaur, H. (2011). Phytochemical screening and Extraction: A Review. *Internationale Pharmaceutica Scientia*. 1(1): 98-106.
- Tiwari, R.K., Udayabanu, M. and Chanda, S. (2017). GC-MS Analysis of essential oil composition of *Clerodendrum serratum* L: A Traditional plant of India. *Asian Journal of Pharmaceutical and clinical Research*. 10(7): 0974-2441.
- Tobias, T.A., Wood, L.G. and Rastogi, D. (2019). Carotenoids, fatty acids and disease burden in obese minority adolescents with asthma. *Clinical and Experimental Allergy*. 49(6): 838-846.
- Tomata, Y., Larsson, S.C. and Hagg, S. (2020). Polyunsaturated fatty acids and risk of Alzheimer's disease: A Mendelian randomization study. *European Journal of Nutrition*. 59(4): 1763-1766.
- Topwal, M. and Uniyal, S. (2018). Review on Important Ethno-Medicinal Plants in Uttarakhand. *International Journal of Pure and Applied Bioscience*. 6(2): 455-464.
- Toree, G.L.T.D., Arollado, E.C., Atienza, A. A. and Manalo, R. A. M. (2017). Evaluation of antioxidant capacity and identification of bioactive compounds of crude methanol extracts of *Caesalpinia pulcherrima* (L.) Swartz. *Indian Journal of Pharmaceutical Sciences*. 79(1): 113-123.
- Trowbridge F. and Martorell R. (2002). Forging effective strategies to combat iron deficiency. Summary and Recommendations. *Journal of Nutrition*. 85(7): 75-80.
- Tulucku, E., Yalcin, H., Ozturk, I. and Sagdic, O. (2012). Changes in the fatty acid compositions and bioactivities of clary sage seeds depending on harvest year. *Industrial Crops and Products*. 39(39): 69-73.
- Tuncturk, M. and Ozgokce, F. (2015). Chemical composition of some Apiaceae plants commonly used in herby cheese in Eastern Anatolia. *Turkish Journal of Agriculture and Forestry*. 39(1): 55-62.
- Turkmen, N., Sari, F. and Velioglu, Y.S. (2006). Effects of extraction solvents on concentration and antioxidant activity of black and black mate tea polyphenols determined by ferrous tartrate and Folin-Ciocalteu methods. *Food Chemistry*. 99(4): 835-41.
- Twine, W., Moshe, D., Netshiluvhi, T. and Siphugu, V. (2003). Consumption and direct-use values of savanna bio-resources used by rural households in Mametja, a

semi-arid area of Limpopo Province, South Africa. *South African Journal of Science*. 99(5): 467-473.

Tyagia, T. and Agarwala, M. (2017). GC-MS analysis of invasive aquatic Weed, *Pistia Stratiotes* L. and *Eichhornia Crassipes* (Mart.) Solms. *International Journal of Current Pharmaceutical Research*. 9(3): 111-117.

Uddin, S.J., Darren, G. and Tiralongo, E. (2011). Cytotoxic Effects of Bangladeshi Medicinal Plant Extracts. *Evidence-Based Complementary and Alternative Medicine*. 1-7. doi:10.1093/ecam/nep111.

Ukiva M., Akihisa T., Tokuda H., Suzuki H., Mukainaka T., Ichiishi E., Yasukawa, K., Kasahara, Y. and Nishino, H. (2002). Constituents of Compositae plants: III. Antitumor promoting effects and cytotoxic activity against human cancer lines of triterpene diols and triols from edible *Chrysanthemum* flowers. *Cancer Letters*. 177(1): 7-12.

Ullah, S. and Hyun, C.G. (2020). Evaluation of total flavonoid, total phenolic contents, and antioxidant activity of strychnobiflavone. *Indonesian Journal of Chemistry*. 20(3): 716-21.

Ullah, Z., Baloch, M.K., Khader, J.A., Baloch, I.B., Ullah, R., Abdeislam, N.M. and Noor, S. (2013). Proximate and nutrient analysis of selected medicinal plants of Tank and South Waziristan area of Pakistan. *African Journal of Pharmacy and Pharmacology*. 7(5): 179-184.

Umarani, G. and Nethaji, S. (2021). Determination of bioactive compounds from *Erythrina variegata* leaf extract using gas chromatographic and mass spectroscopic techniques. *International Journal of Botany Studies*. 6(2): 111-114.

Unuofin, J.O., Otunola, G.A. and Afolayan, A.J. (2017). Essential Oil Composition, Nutrient and Anti-nutrient Analysis of *Vernonia mespilifolia* Less. *Research Journal of Botany*. 12(2): 38-45.

Upadhyay, B., Parveen, Dhaker, A.K., Kumar, A. (2010). Ethnomedicinal and ethnopharmaco-statistical studies of Eastern Rajasthan, India. *Journal of Ethnopharmacology*. 129: 64-86.

Uraku, A.J. (2017). Phytochemical and mineral analysis of *Culcasia scandens* leaves. *EC Pharmacology Toxicology*. 3(5):146-151.

Usher, P. J. (2000). Traditional ecological knowledge in environmental assessment and management. *Arctic*. 53(2): 183-193.

- Usman, A., Onore, R.O., Oforghor, O.A., Mohammed, J. and Usman, N.L. (2020). Total phenolic and Flavonoid contents, Antioxidant Activity and Phytochemical Screening of Calotropis Procera Stem Bark Extracts. *Communication in Physical Sciences*. 5(3): 233-239.
- Usman, H., Abdulrahman, F.I. and Usman, A. (2009). Qualitative phytochemical screening and in vitro antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). *African Journal of Traditional, Complementary and Alternative Medicines*. 6(3): 289-295.
- Van de, P.M.C.G., Dejong, C.H.C. and Soeters, P.B. (2006). Adequate range for sulfur-containing amino acids and biomarkers for their excess: Lessons from Enteral and Parenteral Nutrition. *The Journal of Nutrition*. 136(6): 1694S-1700S.
- Vanamane, R.S., Vhankade, A.M., Bhosale, P.A., Salgar, P.S. and Manjunath, G. (2021). FTIR (Fourier transform infrared spectroscopy) spectroscopic analysis of dried leaf and fruit peel extract of *Capparis divaricata* lam. *International Journal of Environment, Agriculture and Biotechnology*. 6(6): 141-145.
- Venkanna, B., Uma, A., Suvarnalaxmi, Ch., Chandrasekharnath, N., Prakasham, R. S. and Jayalaxami, L. (2013). Antimicrobial property of *Datura* leaf extract against methicillin- resistant staphylococcus aureus isolated from urethral and skin suppurative infection. *Current Trends in Biotechnology and Pharmacy*. 7(3): 782 - 792.
- Victoria, S.H., Das, S., Lahlhenmawia, H., Phucho, L. and Shantabi, L. (2012). Study of analgesic, antipyretic and anti-inflammatory activities of the methanolic extract of *Blumea lanceolaria* (Roxb.) Druce. *International Journal of Life Sciences Biotechnology and Pharma Research*. 1(3): 2250-3137.
- Vidya, S.M., Krishna, V. and Manjunatha, B.K. (2007). Evaluation of hepatoprotective activity of *Clerodendrum serratum* L. *Indian Journal of Experimental Biology*. 45(6): 538-542.
- Vijay, R., Shukla, J. and Saxena, R. (2016). Invitro effect of various plant growth promoters on *Clerodendrum serratum*. *International Journal of Pure and Applied Bioscience*. 4(6): 186-192.
- Visveshwari, M., Subbaiyan, B. and Thangapandian, V. (2017). Phytochemical analysis, antibacterial activity, FTIR and GCMS analysis of *Ceropegia juncea* Roxb. *International Journal of Pharmacognosy and Phytochemical Research*. 9(7): 914-920.

- Wang, W., Li, J., Zhang, H., Wang, X., Fan, J. and Zhang X. (2019). Phenolic compounds and bioactivity evaluation of aqueous and methanol extracts of *Allium mongolicum* Regel. *Food Science and Nutrition*. 7(2): 779-787.
- Wani, Z.A., Kumar, N. and Akash. (2016). Ethnobotanical Study of Some Threatened Plants in District Baramulla, Kashmir, Jammu and Kashmir, India. *International Journal of Current Research Bioscience Plant Biology*. 3(2): 58-64.
- Wei, X., Koo, I., Kim, S. and Zhang, X. (2014). Compound identification in GC-MS by simultaneously evaluating the mass spectrum and retention index. *The Analyst*. 139(10): 2507-2514.
- WHO traditional medicine strategy. 2002-2005.
- WHO/FAO. (1985). WHO FAO report: Energy and protein requirements. WHO Technical Report Series No. 724. Geneva.
- Wintola, O.A. and Afolayan, A.J. (2011). Phytochemical constituents and antioxidant activities of the whole leaf extract of *Aloe ferox* Mill. *Pharmacognosy Magazine*. 7(28): 325-333.
- Wu, G. (2009). Amino acids: Metabolism, functions, and nutrition. *Amino Acids*. 37(1): 1-17.
- Wu, H., Xu, L. and Ballantyne, C.M. (2020). Dietary and pharmacological fatty acids and cardiovascular health. *The Journal of Clinical Endocrinology and Metabolism*. 105(4): 1030-1045.
- Wu, K., Zhao, T., Hogstrand, C., Xu, Y., Ling, S., Chen, G. and Luo, Z. (2020). XR-mediated inhibition of autophagy contributes to FA-induced TG accumulation and accordingly reduces FA-induced lipotoxicity. *Cell Communication and Signalling*. 18:1-16. doi: 10.1186/s12964-020-0525-1.
- Xi-Wen, Li., Liu, M., Bao-Sheng, L., Lu, L. and Yue-Ying, R. (2019). Species identification of poisonous medicinal plant using DNA barcoding. *Chinese Journal of Natural Medicines*. 17(8): 0585-0590.
- Xue-Na, B., J. Cheng, W. Liang, M. LanQing, L. Yu-Bo, S. Guang-Lu and W. You-Nian. (2012). Antifungal activity of extracts by supercritical carbon dioxide extraction from roots of *Stellera chamaejasme* L. and analysis of their constituents using GC-MS. In *Advances in Intelligent and Soft Computing*; Zhu, E., Sambath, S., Eds.; Springer: Berlin/Heidelberg, Germany. 134, pp. 653-662.

- Yadav, P. and Malpathak, N. (2016). Estimation of Antioxidant Activity and Total Phenol, Flavonoid Content among Natural Populations of Caper (*Capparis moonii*, Wight) from Western Ghats Region. *Indian Journal of Pharmaceutical Education and Research*. 50(3): 495-501.
- Yadav, R.N.S. and Agarwala, M. (2011). Phytochemical analysis of some medicinal plants. *Journal of Phytology*. 3(12): 10-14.
- Yamuna P., Abirami P., Vijayashalini P. and Sharmila M. (2017). GC-MS analysis of bioactive compounds in the entire plant parts of ethanolic extract of *Gomphrena decumbens* Jacq. *Journal of Medicinal Plants Studies*. 5(3): 31-37.
- Yang, Y.M., Jia, S.S., Xi, G.P., Zhang, M., Chen, Y.B., Lei, B. and Dhong, X.S. (2013). Induction of apoptosis by D-limonene is mediated by inactivation of Akt in LS174T human colon cancer cells. *Oncology Reports*. 29(1): 349-354.
- Yao, H., Song, J., Liu, C., Luo, K., Han, J., Li, Y. and Pang, X. (2010). Use of ITS2 region as the universal DNA barcode for plants and animals. *PLoS One*. 5(10): e13102. doi:10.1371/journal.pone.0013102.
- Yasir, A. and Ahmad, A. (2018). Impact of processing on nutritional and antinutritional factors of legumes: a review. *Annals Food Science and Technology*. 19(2): 199-215.
- Yasuo, F., Agustine, S and Made, S. (1988). Sesquiterpene lactones from *Blumea balsamifera*. *Phytochemistry*. 27: 1109-1111.
- Ye, Z., Liang, Z., Mi, Q. and Guo, Y. (2020). Limonene terpenoid obstructs human bladder cancer cell (T24 cell line) growth by inducing cellular apoptosis, caspase activation, G2/M phase cell cycle arrest and stops cancer metastasis. *Journal of the Balkan Union Oncology*. 25(1): 280-285.
- Yeboah, E.M.O. and Majinda, R.R.T. (2009). Radical Scavenging Activity and Total Phenolic Content of Extracts of the Root Bark of *Osyris lanceolata*. *Natural Product Communications*. 4(1): 89 -94.
- Yildirm, A., Mavi, A., Oktay, M., Kara, A. A., Algur, F., & Bilaloglu, V. (2000). Comparison of antioxidant and antimicrobial activities of Tilia (*Tilia Argentea* Desf Ex DC), Sage (*Salvia triloba* L.), and Black Tea (*Camellia sinensis*) extracts. *Critical Reviews in Food Science and Nutrition*. 48(10): 5030-5034.
- Yimer, A., Forsido, S.F., Addis, G. and Ayelind, A. (2023). Nutritional composition of some wild edible plants consumed in Southwest Ethiopia. *Heliyon*. 9(6): 10.1016/j.heliyon.2023.e16541.

Zalke, A.S., Kulkarni, A.V., Devendra, S.S. and Duraiswamy, B. (2010). In vivo anticancer activity of *Clerodendrum serratum* (L) Moon. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 1(3): 0975-8585.

Zavala-Sanchez, M.A., Rodriguez-Chavez, J.L., Figueroa-Brito, R., Quintana-Lopez, C.M., Bah, M.M., Campos-Guillen, J., Bustos-Martinez, J.A., Zamora-Avella, D. and Ramos-Lopez, M.A. (2020). Bioactivity of 1-octacosanol from *Senna crotalarioides* (Fabaceae: Caesalpinioideae) to control *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Florida Entomologists*. 102(4): 731-737.

Zhang, J., Hu, X., Wang, P., Huang, B., Sun, W., Xiong, C., Hu, Z., Hu, Z. and Chen, S. (2018). Investigation on species authenticity for herbal products of *Celestrus obiculatus* and *Tripterygium wilfordii* from markets using ITS2 barcoding. *Molecules*. 23(4): 967-980.

Zhao, H., Fan, W., Dong, J., Lu, J. Cheng, J., Shan, L., Lin, Y. and Kong, W. (2008). Evaluation of antioxidant activities and total phenolic contents of typical malting barley varieties. *Food Chemistry*. 107(1): 296-304.

Zhao, L. L., Feng, S. J., Tian, J. Y., Wei, A. Z. and Yang, T. X. (2018). Internal transcribed spacer 2 (ITS 2) barcodes: A useful tool for identifying Chinese *Zanthoxylum*. *Applications in plant sciences*. 6(6): e01157.

Zhao, W., Admassu, H., Abera, T., Abraha, B. and Yang, R. (2018). Proximate, mineral and amino acid composition of dried laver (*Porphyra* spp.) Seaweed. *Journal of Academia and Industrial Research*. (JAIR). 6(9): 149-154.

Zhou, K. and Yu, L. (2004). Effects of extraction solvent on wheat bran antioxidant activity estimation. *Food Science and Technology*. 37(7): 717-721.

Zhou, Y., Tao, X., Wang, Z., Feng, L., Wang, L., Liu, X., Pan, R., Liao, Y. and Chang, Q. (2019). Hippocampus metabolic disturbance and autophagy deficiency in olfactory bulbectomized rats and the modulatory effect of fluoxetine. *International Journal of Molecular Sciences*. 20(17): 4282.

SURVEY QUESTIONNAIRE

1. Name of the Healer/Informant/Practitioner:
2. Sex (Male/Female):
3. Tribe:
4. Years of Experience:
5. Occupation:
6. Locality:

Scientific Name	
Local Name	
Source (Wild/Cultivated/Both)	
Plant part used	
Diseases	
Mode of preparation and administration	
Tribe used	

Signature/Thumb Impression of the healer:

Collected by:

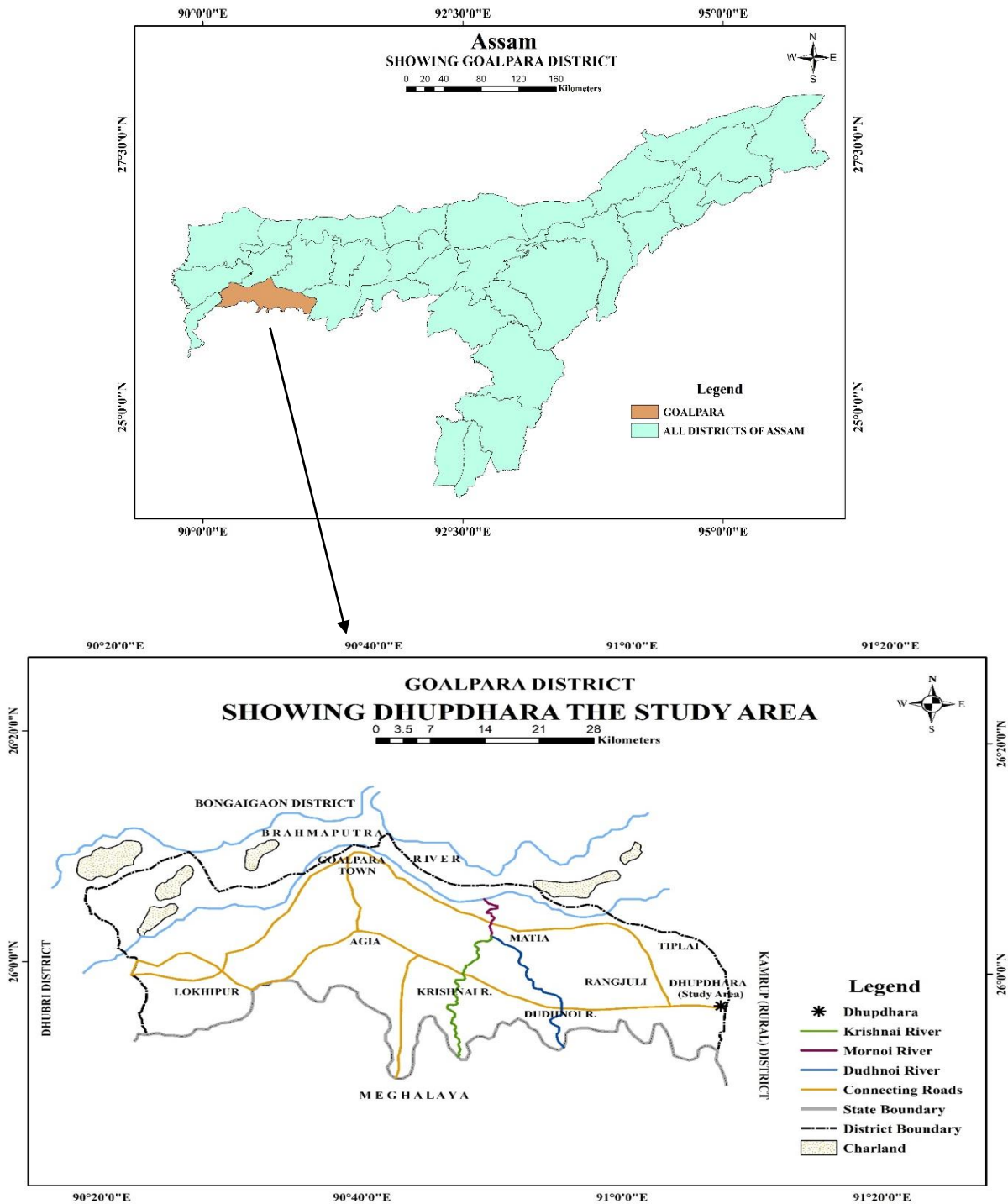


Plate 3.1: Map of Goalpara showing study area

RESEARCH ARTICLE

Phytochemical profiles of leaf extracts of *Rotheca serrata* (L.) Steane & Mabb: a medicinal herb of Assam

Seema Khakhalary and Silistina Narzari*

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Abstract The herb *Rotheca serrata* (L.) Steane & Mabb (*R. serrata*) locally known as Nangal Bhanga is a medicinal herb of Assam, India. It is broadly used in traditional medicine systems of Assam for curing various ailments including hepatitis, ulcer, diabetes and cancer. Through the present work it was intended to investigate the phytochemical constituents antioxidant potential and bioactive compounds of *R. serrata* leaves. Crude extracts were obtained through Soxhlet extraction, using solvents of increasing polarity, i.e., hexane, chloroform and methanol. Antioxidant activities including 2, 2-diphenyl-1-picryl hydrazyl (DPPH) free radical scavenging and H₂O₂ assays were performed using UV-Vis Spectrophotometer. Total phenolics, tannins and flavonoid content were estimated following standard protocols for quantitative phytochemical analysis. Gas Chromatography–Mass Spectrometry (GC–MS) was used to identify bioactive compounds that would account for the above recorded activities. Preliminary screening of phytochemicals indicated the existence of alkaloids, flavonoids, phenols, tannins. Higher concentrations of antioxidants, phenolics, tannins and flavonoids were extracted in methanol solvents compared to the other two solvents. The GC–MS analysis led to identification of 20 potential bioactive compounds of which 7 bioactive compounds were detected in methanol, 7 in hexane and 6 in chloroform extract. Bioactive compounds identified from leaves of *R. serrata* are reported for biological activities like antioxidant, anticancer, anti-tumor and chemo-preventive properties. Findings of this study indicate that methanol extract is a potent solvent for phytochemical extraction and analysis. Further, our study also suggests that isolation and elucidation of these bioactive compounds may play a vital role to find a new drug in near future.

Keywords: Antioxidant, Bioactive, Extract, Medicinal, Phytochemicals

Abbreviations

GC-MS: Gas Chromatography and Mass Spectroscopy
H₂O₂: Hydrogen Peroxide
R. serratum: *Rotheca serratum*
UV-Vis: Ultra Violet- Visible
GUBH: Gauhati University Botanical Herbarium
g: Gram

ml: Millilitre
TPC: Total Phenolic Content
TFC: Total Flavonoid Content
AlCl₃: Aluminium chloride
NaOH: Sodium Hydroxide
SD: Standard Deviation
SPSS: Statistical Package for the Social Sciences
OD: Optical Density
Ic50: Inhibitory Concentration

Introduction

Use of plants as a source of medicine is practised and is passed on through generations among many populations around the globe. So it forms an important component of the health care system. Assam is enriched with plant diversity and several plants have been used traditionally by Assamese people for therapeutic potentials. Plants used in traditional

medicines usually contain a wide range of bioactive compounds that can be used to treat various infectious and chronic diseases (Duraipandiyar *et al.* 2006). Bioactive compounds can be detected through preliminary phytochemical screening tests. The result acquired from preliminary phytochemical screening may aid in discovering novel drugs that come from natural sources.

R. serrata called Nangol bhanga in Assamese is an important medicinal plant belonging to family Lamiaceae. *Rotheca serrata* (L.) scientifically classified as

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Clerodendrum serratum earlier was placed under family Verbanaceae. Phylogenetic analysis of its mitochondrial DNA shifted it to the Lamiaceae family (Steane *et al.* 1997). Traditionally, this plant finds its wide applicability in ethnomedicines of Assam. Keshava (1994) reported the use of *R. serrata* roots in medicinal preparations for treating numerous disorders like asthma, bodyache, bronchitis, cholera, dropsy, eye disorder, fever, inflammations, malaria, ophthalmic, rheumatism, snakebite, tuberculosis, ulcers and wounds. Owing to its biological activities like anti-inflammatory and antipyretic activities, the use of *R. serratum* has been reported for treating diseases as typhoid, cancer, jaundice and hypertension (Mukesh *et al.* 2012). Saha *et al.* (2012) and Kar *et al.* (2014) informed about the analgesic and anti-diabetic potentials of its leaves.

Other scientific report published on extracts and formulations revealed anti-asthmatic, mast cell stabilization and anti-allergic effects in roots of *R. serrata*. Studies on pharmacological activities also include hepatoprotective, anti-oxidant, anti-inflammatory and anticancer potential (Acharya *et al.* 2014). Various phytochemicals including Apigenin-7-glucoside, (7-(β -D-glucopyranosyloxy)-5-hydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one have been previously isolated from the root of *R. serrata* (L) Moon (Bhujpal *et al.* 2010) and D-mannitol, stigmaterol, oleanolic acid, ferulic acid, lupeol, and ursolic acid (Kumar and Niteshwar 2013).

The study was undertaken to screen phytochemicals, evaluate in-vitro antioxidant activities and identify and characterize bioactive compounds in *R. serrata* extracts. This study will provide scientific validation for the therapeutic practice of using *R. serrata* leaves in traditional medicine systems of Assam. Identification of bioactive compounds in *R. serrata* essentially may lead to further dissemination of knowledge on its biological and pharmacological studies.

Materials and methods

Chemicals

All chemicals and solvents used were of

analytical grade and were purchased from Merck (Germany).

Collection of Plant material

Fresh leaves were collected in the month of January 2021 during the morning hours. The collected plant material was authenticated by the Gauhati University Botanical Herbarium (GUBH), Gauhati University, Assam. A voucher specimen of *Rotheca serrata* (L.) Steane & Mabb bearing accession number 18926 was then submitted to GUBH.

Preparation of Plant extract

Fresh leaves were collected, cleaned, washed and dried under shade for three weeks. The air dried leaves were crushed and pulverized using a clean and sterile electric grinder. 20g of powdered leaves were extracted successively in 200ml of hexane, chloroform and methanol solvent at room temperature for about 24 hours using Soxhlet apparatus. The solvents were evaporated in a rotary vacuum evaporator (Model no.#EV11) to obtain crude extracts. Finally, the yield percent of crude extract was calculated by the standard formula of Alebiosu and Yusuf (2015).

$$\text{Yield Percent (\%)} = a/b \times 100$$

Where, a = dry weight of extract obtained
b = initial weight of powdered material

Phytochemical Screening

The plant extracts were subjected to phytochemical screening tests by following standard protocols of Evans (2009), Harborne (1998) to confirm the presence of phytochemicals.

Determination of Total Phenolic Content

To evaluate total phenolic content (TPC) in *R. serrata* crude extracts the Folin-Ciocalteu reagent method of Shukla *et al.* (2014) was followed. For the analysis, 0.2 mL of extract (1mg/ml) was added to 2.5mL of 10% Folin-Ciocalteu reagent and then neutralized using 2ml of 7.5% sodium carbonate. The reaction mixture was then incubated in dark at normal room temperature for 30mins. Absorbance

value was measured at 765nm wavelength using a double beam UV-Vis spectrophotometer (UV Analyst-CT8200). The total phenolic content as mean SD (n=3) were calculated from the linear regression equation of gallic acid standard plot. The results are expressed as mg/g gallic acid equivalent (GAE) of dry extract.

Determination of Total Flavonoid Content

Total flavonoid content in *R. serrata* extracts was determined by the procedure described by Alhakmani *et al.* (2013). Calibration curve was constructed using quercetin as standard. 0.2mL of plant extract (1mg/ml) was diluted with 5ml of distilled water. To it 0.5ml of 5% sodium nitrite solution was added. After 5 mins, 0.6ml of 10% AlCl₃ solution was added. After another 6 mins, 2ml of 1M NaOH solution was added and final volume was adjusted to 3ml with distilled water. The solutions were thoroughly mixed and incubated for 15minutes. Absorbance value of the reaction mixture was measured at 510nm with double beam UV-Vis spectrophotometer (UV Analyst-CT8200) against blank. All the tests were performed in triplicates. Total flavonoid was calculated from the quercetin calibration curve. Results are expressed as mg quercetin equivalent per gram dry weight.

Determination of Total Tannin Content

The tannins were determined using the Folin-Ciocalteu method of CI and Indira (2016). To 0.1ml of plant extract (1mg/ml), 1ml of distilled water was added. To it 0.5 ml Folin-Ciocalteu reagent was added and mixed thoroughly. The mixture was alkalized by adding 1ml of 15% (w/v) Na₂CO₃ and kept in dark for 30 minutes at room temperature. The absorbance of the tannic acid standard solutions as well as sample was measured after colour development at 700nm using the UV-VIS spectrophotometer (UV Analyst-CT8200). Results calculated using the calibration curve were expressed as mg/g equivalent of tannic acid.

Determination of Antioxidant Activity

The antioxidant activities of the plant extract

vary with the solvent used for extraction. It is thus important to use different solvent extract for evaluating the effectiveness of the antioxidant. The antioxidant activity of all three solvent extract i.e. hexane, chloroform and methanol extract was determined using 2, 2-Diphenyl-1-Picryl-Hydrazyl Assay (DPPH) and Hydrogen Peroxide Assay (H₂O₂).

2, 2-Diphenyl-1-Picryl-Hydrazyl Assay (DPPH Method)

Free radical scavenging activity of the crude extracts were evaluated by using DPPH radical scavenging activity method of Alhakmani *et al.* (2013). Ascorbic acid was taken as the standard. Crude extracts and standard ascorbic acid solution of 1mg/ml of varying concentrations ranging from 50 to 250µg/mL were taken in separate test tubes. 2ml of 0.1mM DPPH prepared in methanol was added to each test tube. The solution was mixed and kept in dark at 37°C for 30mins. The decrease in absorbance of each solution was measured at 517nm using UV-Vis spectrophotometer (UV Analyst-CT8200). The solution used for the blank is methanol. Radical scavenging activity expressed as percentage inhibition of the extract and ascorbic acid were calculated using the standard formula:

$$\% \text{ Inhibition} = \frac{\text{OD control} - \text{OD test}}{\text{OD control}} \times 100$$

The concentration of sample required to scavenge 50% of DPPH free radical (IC₅₀) was calculated from the curve of percent inhibitions plotted against their respective concentrations.

Hydrogen Peroxide Scavenging Activity (H₂O₂)

The ability of leaf extracts to scavenge H₂O₂ was studied by the method of Nabavi *et al.* (2008). Different concentrations ranging from 50-250 µg/mL of crude extracts and standard ascorbic acid solution were taken in test tubes. To each test tubes, 0.6mL of H₂O₂ (40mmol/L) and 2ml of phosphate buffer (50mmol/L) (pH 7.4) was added. After 10 minutes, absorbance was measured at 230nm against a blank

solution containing phosphate buffer. The percentage of H₂O₂ scavenged was calculated using following formula:

$$\text{H}_2\text{O}_2 \text{ scavenge (\%)} = \frac{\text{OD control} - \text{OD test}}{\text{OD control}} \times 100$$

Gas Chromatography-Mass Spectroscopy (GC-MS)

GC separation of compounds was performed in Clarus 680 GC from Perkin Elmer, USA and MS study in Clarus 600C MS from Perkin Elmer, USA. For compound separation in GC, 2µl of extract was taken and injected into GC system through autosampler with a split ratio of 10:1 in splitmode. The GC system was fitted with 60m length capillary column of 0.25mm diameter and film thickness of 0.25µm. The column composition was 5% of diphenyl, 95% of dimethylpolysiloxane with a mass range around 50-600amu. Mass Spectra of the compounds were constructed at 70eV in Electron Impact positive (EI+) mode. The programming of column oven temperature was fixed between 60°C to 300°C and was held for 10mins. The temperature for the injector was kept at 280°C. The carrier gas used was Helium of 99.99% purity and the flow rate was fixed at 1ml/min⁻¹. The total run time for the whole GC-MS run was 51.83 minutes (Hema *et al.* 2010).

Identification of Compounds

Interpretation on Mass-Spectrum GC-MS was conducted using the database of National

Institute Standard and Technology (NIST 2014). The spectrum of components obtained from our study was compared with the spectrum of known components already stored in the NIST library. Through this comparison name, molecular weight and structure of the unknown components in *R. serrata* extracts were thus ascertained (Hema *et al.* 2010).

Statistical Analyses

All statistical analyses were performed in SPSS 26.0 version software. Experimental measurements were carried out in triplicates and are expressed as average of three analysis ± standard deviation (SD).

Results

Yield % of the crude extract

The yield % for hexane, chloroform and methanol extract of *R. serrata* were 2%, 2.8% and 3.5% respectively in 25g of powdered material used.

Preliminary Phytochemicals Screening

Results of the preliminary phytochemical screening disclosed the presence of various phytochemicals. It showed the presence of major classes of secondary metabolites such as tannins, flavonoids, phenolics, steroids, phytosterols etc. in all the extracts (Table-1). However, saponins, oils and fats were absent in chloroform and hexane extracts but present in

Table-1: Phytochemical components of solvent extracts of *R. serrata* based on preliminary screening.

Phytochemical constituents	Test	Methanol Extract	Chloroform Extract	Hexane extract
Alkaloids	Mayer's test	+	-	+
Tannins	Ferric chloride test	+	+	+
Saponins	Foam test	+	-	-
Phenolics	Ferric chloride test	+	+	+
Flavonoids	Alkaline reagent test	+	+	+
Phytosterols	Liebermann Burchard's test	+	+	+
Steroids	Salkowski's test	+	+	+
Terpenoids	Salkowski's test	-	+	+
Oils and fats	Spot test	+	-	-

(+)=Detected; (-)=Not detected.

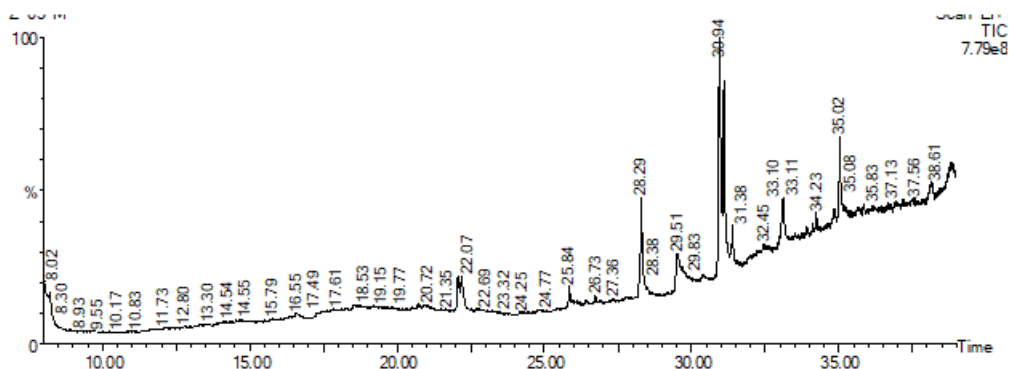


Figure 1: L GCMS chromatogram of methanolic extract of *R. serrata* leaves.

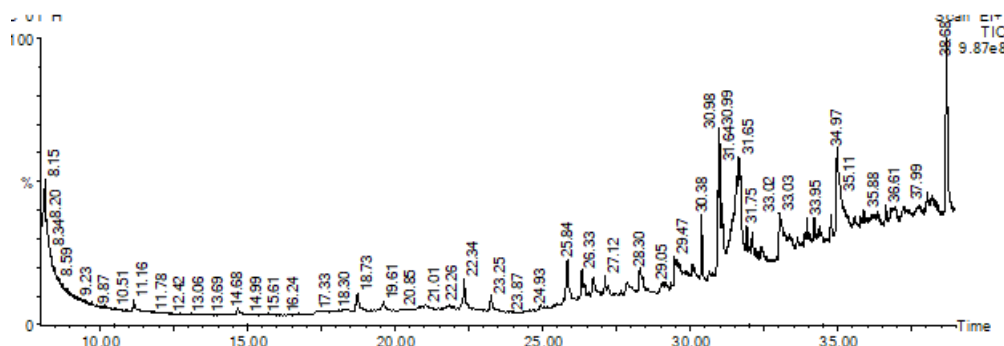


Figure 2: L GCMS chromatogram of hexane extract of *R. serrata* leaves.

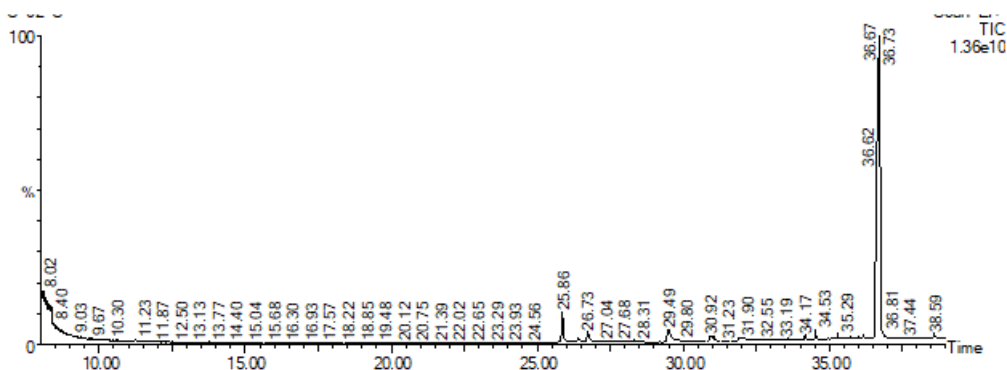


Figure 3: L GCMS chromatogram of chloroform extract of *R. serrata* leaves.

methanol extract.

Quantitative Phytochemical Screening

Total Phenolic Content (TPC)

The Total Phenolic Content (TPC) of leaf extracts is expressed in terms of GAE. The linear regression equation obtained from the

standard plot of gallic acid was $y = 0.004x + 0.059$, $R^2 = 0.984$ where y is absorbance and x is the amount of gallic acid in μg . The TPCs were calculated from the standard plot (Table-2).

Total Flavonoid Content (TFC)

The Total Flavonoid Content (TFC) of *R.*

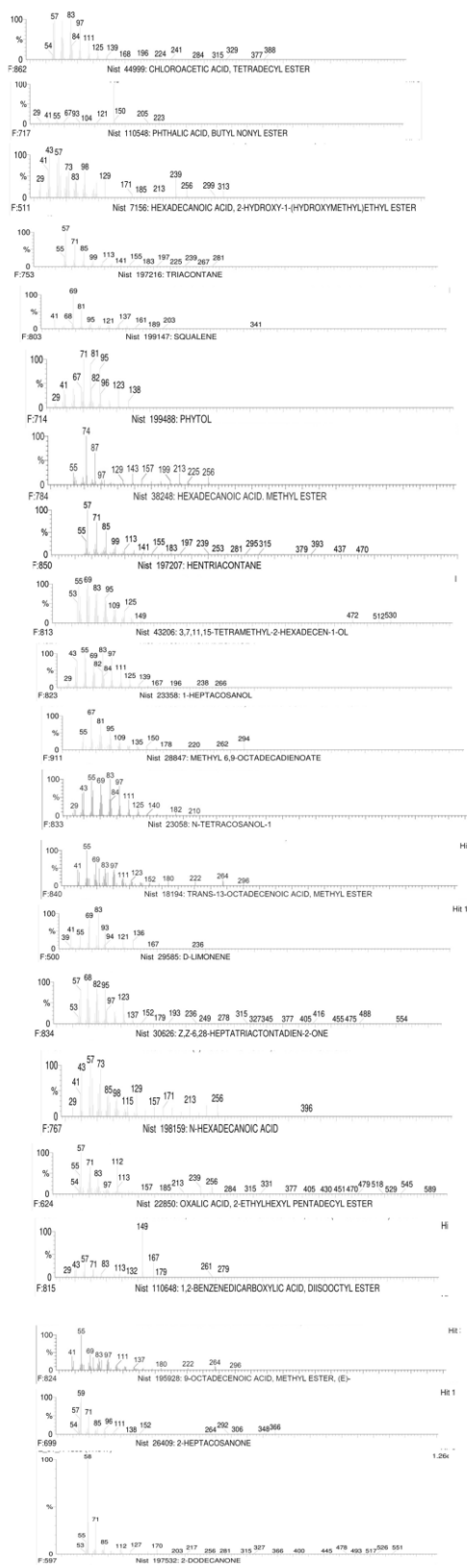


Figure 4: Mass spectrum of bioactive compounds present in *R. serrata* leaves extract

serrata leaf extracts is expressed in terms of Quercetin Equivalent (QE). The linear regression equation $y = 0.004x + 0.714$, $R^2 = 0.917$ where y is absorbance and x is the amount of quercetin in μg was obtained from standard plot of quercetin. The TFC was calculated from the standard plot and is presented in Table-2.

Total Tannin Content (TTC)

The Total Tannin Content (TTC) of leaf extracts is expressed in terms of Tannic Acid Equivalent (TAE). The linear regression equation $y = 0.074x - 0.517$, $R^2 = 0.901$ where y is absorbance and x is the amount of tannic acid in μg were obtained from the standard plot of tannic acid. The TTCs were calculated from the standard plot and is shown in Table-2

Among the three extracts, maximum amount of phytochemicals tested i.e phenolic, flavonoid and tannin were found in methanol extract followed by chloroform and hexane extracts.

Antioxidant Activity

The antioxidant activity of three crude extracts of *R. serrata* leaves was studied by commonly used radical scavenging methods such as DPPH and H_2O_2 . The scavenging effect of leaf extracts on the DPPH and H_2O_2 free radicals were calculated from their absorbance. Inhibitory concentrations (i.e. IC_{50} value) of each extracts were calculated from the calibration curve of their percentage inhibition and results were compared with the standard ascorbic acid. The highest antioxidant activity expresses the lowest IC_{50} (Table-3).

Methanol extract showed lowest IC_{50} value in DPPH radical scavenging activity compared to hexane and chloroform extracts. But, the H_2O_2 scavenging activity of extracts were found in the following order of chloroform>methanol> hexane>. Both the assays have lower antioxidant capacity compared to ascorbic acid (Standard).

GCMS Analysis of the Plant Extract

Gas chromatography mass spectroscopy analysis was carried out to identify bioactive

Table-2. Total phenolic, flavonoid and tannin content of the crude extracts of *R. serrata*.

Crude Extract	Total phenolic content (mg of GAE/g dry extract)	Total flavonoid content (mg of QE/g dry extract)	Total tannin content (mg of TAE/g dry extract)
Hexane	30.35±2.24	16.10±0.145	7.50±0.015
Chloroform	56.76±1.75	21.43±0.098	7.71±0.02
Methanol	73.41±1.66	27.41±0.635	12.68±0.032

Mean values ± standard deviations of triplicate determinations are reported.

Table-3: IC₅₀ value (in µg/ml) of *R. serrata* extracts from DPPH and H₂O₂ scavenging assay

Assays	Hexane	Chloroform	Methanol	Ascorbic Acid (Standard)
DPPH	226.95±0.997	220.96±2.096	156.37±0.910	115.30±1.35
H ₂ O ₂	288.64±4.976	202.36±2.835	224.58±1.123	161.13±1.84

compounds in *R. serrata* leaf extracts. A total of 20 bioactive compounds were identified from the GC-MS analysis. Out of which, 7 compounds were detected in hexane extract, 6 in chloroform and 7 in methanol extract. The GC-MS chromatogram and Mass Spectrum of bioactive compounds obtained from methanol, hexane and chloroform extract are presented in (Figures. 1, 2, 3 and 4) respectively while the chemical constituents along with their retention time (RT), molecular formula, molecular weight (MW), and peak area percentage and Pub Chem ID are presented in Table-4. The mass spectrum profile of GC-MS confirmed the presence of bioactive compounds with retention time ranging between 22.34 minutes - 35.02 minutes.

Discussion and conclusion


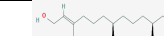





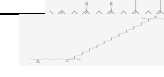



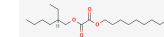
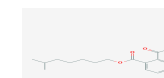


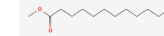

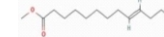


Crude extracts obtained from *R. serrata* leaves were observed for colour formation. The colour of hexane extract was yellow where as both chloroform and methanol extract appeared dark green. The dry weight and final weight of *R. serrata* extracts were significantly affected by solvent polarity used for extraction. The yield percentage of the extracts so calculated divulged that methanol extract had high extraction value as compared to chloroform and hexane. When the extracts were screened for phytochemicals, methanol extract contained higher amount of phytochemicals. So, methanol was found to be more potent for extracting phytochemicals compared to hexane and

chloroform. It indicates that the leaf extract contained more polar than non-polar compounds. The study revealed that differences arise in the composition of phytochemicals due to variations in solvent polarities used for sample extraction.

Based on the results of phytochemical screening, the total phenolic, flavonoid and the total tannin content were estimated. The quantitative tests results revealed that there are wide variations in the phytochemical contents of the extracts (Table-2). Quantitative data obtained from standard calibration curve and calculated by linear regression equation expressed that methanolic extract contained significant quantity of phenol, flavonoid and tannin (i.e. 73.4mg of GAE/g extract, 27.41mg of QE/g extract, and 12.68mg of TAE/g extract) in comparison to hexane and chloroform extracts. Our study, ascertained that methanol is a superior solvent for isolating polyphenolic compounds compared to hexane and chloroform. Plant phenolics constitute one of the major groups of compound acting as primary antioxidants and free radical terminators.

The total antioxidant activity of the plant extracts were also evaluated using DPPH and H₂O₂ scavenging assay. Ascorbic acid was used as positive control for both the assays. IC₅₀ value was calculated to evaluate the total antioxidant activity from the linear regression equation. A lower IC₅₀ value corresponds to higher effectiveness of the antioxidant. In the present study, methanol extract showed maximum ability in DPPH radical scavenging activity compared to other solvent extract which was measured by the lowest IC₅₀ value, but it has lower antioxidant capacity compared to ascorbic acid (Standard). The IC₅₀ value in (µg/ml) of the extracts were found in the order of Methanol>Chloroform>Hexane (Table-3). But in H₂O₂ assay, the chloroform extract exhibited the highest antioxidant activity followed by methanol and hexane extract. These variances result from the point that each method is established on the production and use of various radicals and species that are actively involved in oxidative process by different mechanisms. The variation might be

Table 4: Bioactive compounds identified in hexane, chloroform and methanol extract of *R. serrata*.

	Retention time (min)	Compound Name	Molecular formula	Molecular Weight	Peak Area (%)	2D Structure of compounds	Pub Chem ID
Hexane extract	22.34	Triacontane	C ₃₀ H ₆₂	422	1.179		12535
	25.84	Phytol	C ₂₀ H ₄₀ O	296	2.296		5280435
	30.98	Phthalicacid, butylnonyl ester	C ₂₁ H ₃₂ O ₄	348	6.099		6423814
	31.64	Hentriacontane	C ₃₁ H ₆₄	436	11.974		12410
	33.03	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester	C ₁₉ H ₃₈ O ₄	330	2.902		341733478
	34.98	Chloroacetic acid, tetradecylester	C ₁₆ H ₃₁ O ₂ Cl	290	8.220		519540
	38.68	Squalene	C ₃₀ H ₅₀	410	6.320		638072
Chloroform extract	26.72	Z,z-6,28-heptatriacontadien-2-one	C ₃₇ H ₇₀ O	530	1.443		5365964
	29.49	N-hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256	4.186		16213579
	30.92	1-heptacosanol	C ₂₇ H ₅₆ O	396	0.766		74822
	31.90	Methyl 6,9-octadecadienoate	C ₁₉ H ₃₄ O ₂	294	0.891		549027
	34.52	Oxalic acid, 2-ethylhexylpentadecylester	C ₂₃ H ₄₈ O ₄	412	0.637		6420799
	36.69	1,2-Benzenedicarboxylic acid, diisooctylester	C ₂₄ H ₃₈ O ₄	390	69.89		33934
Methanolic extract	22.07	2-Dodecanone	C ₁₂ H ₂₄ O	184	1.914		22556
	25.84	3,7,11,15-tetramethyl-2-hexadecen-1-ol	C ₂₀ H ₄₀ O	296	3.553		5366244
	28.29	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	270	1.983		8181
	29.51	N-tetracosanol-1	C ₂₄ H ₅₀ O	354	7.883		10472
	30.94	9-octadecenoic acid, methyl ester	C ₁₉ H ₃₆ O ₂	296	3.372		5280590
	31.38	Trans-13-octadecenoic acid, methyl ester	C ₁₉ H ₃₆ O ₂	296	15.453		5364506
	33.10	D-limonene	C ₁₀ H ₁₆	136	1.681		440917

the due to the complex nature of phytochemicals present in the extracts or the solvent used for extraction, etc (Rakholiya *et al.* 2011). It is thus important to perform several analytical methods for evaluating the effectiveness of antioxidants present in the plant. Phytoconstituents like flavonoids and phenolic compounds, commonly found in plants have been reported for its multiple biological effects, one of which is the antioxidant property (Tungmunnithum *et al.* 2018). Hence, in our study, the observed antioxidant activity might have ascended from the presence of phenolics and flavonoid contents in the extract of *R. serrata*. Tannins have also been reported to be associated in traditional treatment of ulcerated tissue and for its remarkable activity in cancer prevention (Batista *et al.* 2012). The optimum yield of tannin content of *R. serrata* leaves corroborates its traditional usage in treatment of cancer. Phytoconstituents having biological activities which include anti-inflammatory, antioxidant, antibacterial, antitumor, antidiabetic, hypercholesterolemia activities have been identified in the present study. The presence of these bioactive compounds stakes the reported utilization of *R. serrata* for various ailments. Based on abundance, the top three compounds present in the hexane extract were Hentriacontane (11.97%), Chloroacetic acid, tetradecyl ester (8.22%) and Squalene (6.32%). 1, 2-Benzene dicarboxylic acid, diisooctyl ester (69.89%) followed by N-hexadecanoic acid (4.18%) and Z, z-6,28-heptatriactontadien-2-one (1.443%) were the major compounds found in chloroform extract. Trans-13-octadecenoic acid methyl ester (15.453%), N-tetracosanol-1 (7.883%) and 3, 7, 11, 15-tetramethyl-2-hexadecen-1-ol (3.553%) were the top major bioactive compounds found in the methanol extract.

Most of the compounds identified in *R. serrata* leaves through GCMS analysis are biologically active compounds. Hentriacontane, the top compound obtained in the hexane extract is reported for its various pharmacological effects including antitumor activity (Kim *et al.* 2011). Chloroacetic acid, tetradecyl ester found in hexane extract (8.220%) is known for its antioxidant properties (Shyam and Suresh 2013). Triacontane is

reported for its antibacterial, antidiabetic and antitumor activities (Mallick and Dighe 2014). The compound Squalene has antioxidant, chemopreventive, anti-tumour and hypercholesterolemia activities (Gunes 2013). N-hexadecanoic acid have several biological activities like antioxidant, 5 alpha reductase inhibitor, anti-fibrinolytic, antimicrobial activity, hypercholesteromic nematocide, pesticide, antiandrogenic flavor, and haemolytic property (Starlin *et al.* 2019). 3,7,11,15-tetramethyl-2-hexadecen-1-ol is a diterpene associated with biological activities like antimicrobial, anti-inflammatory, anticancer and diuretic activities (Hamidi *et al.* 2016). Al-Abd *et al.* (2015) reported 1-heptacosanol to have antimicrobial & antioxidant property. Z, z-6, 28-heptatriactontadien-2-one has vasodilatory property (Mallikadevi *et al.* 2012). 1, 2-Benzene dicarboxylic acid, diisooctyl ester the top major bioactive compound obtained in the study has antioxidant property (Li *et al.* 2012). Phytol was reported to exhibit antioxidant and antinociceptive effects (Santos *et al.* 2013). Phytol has been reported with cytotoxic activities against breast cancer cell lines (MCF7) (Pejin *et al.* 2014). Hexadecanoic acid, methyl ester possesses anti-bacterial and antifungal properties (Chandrasekaran *et al.* 2011). Phthalic acid, butyl nonyl ester is not known for any biological activities yet. Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethyl ester has antioxidant activity (Arora and Kumar 2018). The minor bioactive compound detected in chloroform extract namely Oxalic acid, 2-ethylhexyl pentadecyl ester (0.637%) has not been informed for any biological activities. Methyl 6, 9 octadecadienoate has anti-oxidant activity (Berdeaux *et al.* 1998). 2-Dodecanone detected in methanol extract has insecticidal and repellent activity (Wang *et al.* 2019). N-tetracosanol-1, an alcoholic compound present in methanolic extract is known for its antioxidant properties (Lakshmi and Nair 2017). Trans-13-octadecenoic acid, methyl ester has anti-inflammatory, antiandrogenic, anticancerous, dermatitogenic, hypocholesterolemic, anemiagenic and insectifuge properties (Krishnamurthy and Subramaniam 2014). 9-octadecenoic acid, methyl ester has anticancer activity (Asghar *et*

al. 2011). Anandkumar *et al.* (2020) informed about the cardioprotective, hepatoprotective and anti-carcinogenic activities of D-limonene.

The above mentioned bioactive compounds from extracts of *R. serrata* leaves hold the reported biological activities. Triacotane in hexane extract justifies the reported anti-diabetic activity of the leaves. The reported anticancer activity of the Hexadecanoic acid, methyl ester, Phytol, 3,7,11,15-tetramethyl-2-hexadecen-1-ol and Squalene as informed by various authors supports the reported use of *R. serrata* in cancer treatment.

Based on the above results, it can be concluded that *R. serrata* is a good source of phytochemicals with potent pharmacological properties. It is also evident that methanol extract have superior antioxidant capacity compared to other solvent extract used in this study. 20 bioactive compounds identified in *R. serrata* leaves can contribute effective biological activities like antioxidant, antimicrobial, anticancer, anti-inflammatory, chemopreventive, anti-tumour, hypercholesterolemia activities, 5 alpha reductase inhibitor, anti-fibrinolytic activity if utilised properly. Biological activities of bioactive compounds in *R. serrata* leaf extract support the reported use of this plant in treating various ailments. Identification of bioactive compound in *R. serrata* can serve as the basis for determining possible health benefits of this plant. This study exposes new horizons for further biological and pharmacological research for better exploration of bioactive compounds from plants and their establishment for proper utilization in healthcare systems.

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References

- Acharya N S, Patel J J and Acharya S R 2014 *Clerodendrum serratum* (L.) Moon.- A review on traditional uses, phytochemistry and pharmacological activities. *J Ethnopharmacol* **154** 268-285.
- Al-Abd N M, Mohamed Z N, Mansor M, Azhar F, Hasan M S and Kassim M 2015 Antioxidant, antibacterial activity, and phytochemical characterization of *Melaleuca cajuputi* extract. *BMC Complement Altern Med* **15** 1-13.
- Alebiosu C O and Yusuf A J 2015 Phytochemical screening, thin-layer chromatographic studies and UV analysis of extracts of *Citrullus lanatus*. *J Pharm Chem Biol Sci* **3** 214-220.
- Alhakmani F, Kumar S and Khan SA 2013 Estimation of total phenolic content, in-vitro antioxidant and anti-inflammatory activity of flowers of *Moringa oleifera*. *Asian Pac J Trop Biomed* **3** 623-627.
- Anandakumar P, Kamaraj S and Vanitha MK 2020 D-limonene: A multifunctional compound with potent therapeutic effects. *J Food Biochem* **45** 1-10.
- Arora S and Kumar G 2018 Phytochemical screening of root, stem and leaves of *Cenchrus biflorus* Roxb. *J Pharmacogn Phytochem* **7** 1445-1450.
- Asghar S F, Rehman H U, Choudahry M I and Rahman A U 2011 Gas Chromatography-mass spectrometry (GC-MS) analysis of petroleum ether extract (oil) and bio-assays of crude extract of *Iris germanica*. *Int J Genet Mol Biol* **3** 95-100.
- Batista L M, Jesus N Z T de J, Falcao H de S, Gomes I F, Leite T J de A L, Lima G R de M, Filho J M B, Tavares J F, Silva M S de A S and Filho P F de A F 2012 Tannins, Peptic Ulcers and related mechanism. *Int J Mol Sci* **13** 3203-3228.
- Berdeaux O, Voinot L, Angioni E, Juanéda P and Sebedio J L 1998 A simple method of preparation of methyl trans-10, cis 12- and cis-9, trans-11-octadecadienoates from methyl linoleate. *Journal Am Oil Chem Soc* **75** 1749-1755.
- Bhujbal S S, Nanda R K, Deoda R S, Kumar D, Kewatkar S M, More L S and Patil M J 2010 Structure elucidation of a flavonoid glycoside from the roots of *Clerodendrum serratum* (L.) Moon, Lamiaceae. *Rev Bras Farmacogn* **20** 1001-1002.

- Chandrasekaran M, Senthilkumar A and Venkatesalu V 2011 Antibacterial and antifungal efficacy of fatty acid methyl esters from the leaves of *Sesuvium portulacastrum* L. *Eur Rev Med Pharmacol Sci* **15** 775-780.
- CI K C and Indira G 2016 Quantitative estimation of total phenolic, flavonoids, tannin and chlorophyll content of leaves of *Strobilanthes kunthiana* (Neelakurinji). *J Med Plants Stud* **4** 282–286.
- Duraipandiyan V, Ayyanar M and Ignacimuthu S 2006 Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India. *BMC Complement Altern Med* **6** 35-41.
- Evans WC 2009 Trease and Evans Pharmacognosy. London.
- Gunes F E 2013 Medical use of Squalene as a natural antioxidant. *Musbed* **3** 220-228.
- Hamidi N, Laid Z, Djellouli M and Ha L 2016 Chemical characterization by GC-MS from the aerial parts of *Fagonia longispina* (Zygophyllaceae). *Asian J Pharm Clin Res* **9** 152-153.
- Harborne JB 1998 *Phytochemical methods: A guide to modern techniques of plant analysis*. London: Chapman & Hall.
- Hema R S, Kumaravel S, Gomathi and Sivasubramaniam C 2010 Gas Chromatography-Mass Spectroscopic analysis of *Lawsonia inermis* leaves. *NY Sci J* **3** 141-143.
- Kar M K, Swain T R and Mishra S K 2014 Antidiabetic activity of *Clerodendrum serratum* (L) moon leaves in streptozotocin induced diabetic rats. *Asian J Pharm Clin Res* **7** 2455-3891.
- Keshava M K R 1994 *Medicinal Plants of Karnataka*. Karnataka Forest Dept, Bangalore, India.
- Kim S J, Chung W S and Ko S G 2011 Antiinflammatory effect of *Oldenlandia diffusa* and its constituent, Hentriacontane, through suppression of Caspase-1 activation in mouse peritoneal macrophages. *Phyto ther Res* **25** 1537-1546.
- Krishnamurthy K and Subramaniam P 2014 Phytochemical profiling of leaf, stem, and tuber parts of *Solenaam plexicaulis* (Lam.) Gandhi using GC-MS. *Int Sch Res Notices* 1-13.
- Kulkarni A, Govindappa M, Ramachandra Y L and Koka P 2015 GC-MS Analysis of methanol extract of *Cassia fistula* and it's invitro anticancer activity on human prostate cancer cell line. *Indo Am J Pharm Res* **5** 937-944.
- Kumar P and Niteshwar K 2013 Phytochemical and pharmacological profiles of *Cleodendrum serratum*. *Int J Res Ayurveda Pharm* **4** 276-8.
- Lakshmi M and Nair BR 2017 GCMS analysis of the chloroform extract of bark of *Terminalia travancorensi* Wight and Am. (Combretaceae). *Int J Pharm Sci Res* **8** 794-8.
- Li M, Zhou L, Yang D, Li T and Li W 2012 Biochemical composition and antioxidant capacity of extracts from *Podophyllum hexandrum* rhizome. *BMC Complement Altern Med* **12** 263-263.
- Mallick S S and Dighe V 2014 Detection and estimation of alpha-amyrin, beta-sitosterol, lupeol and n-tricontane in two medicinal plants by high performance thin layer chromatography. *Adv Chem* 1-7.
- Mallikadevi T, Paulsamy S, Jamuna S and Karthika K 2012 Analysis for phytoceuticals and bioinformatics approach for the evaluation of therapeutic properties of whole plant methanolic extract of *Mukia maderaspatana* (L.) M. Roem. (Cucurbitaceae)- A traditional medicinal plant in western districts of Tamil Nadu, India. *Asian J Pharm Clin Res* **5** 163-168.
- Mukesh K R, Gaurav K, Shiv K I, Gotmi S and Tripathi D K 2012 *Clerodendrum serratum*: A clinical approach. *J App Pharm Sci* **2** 11-15.
- Nabavi S, Ebrahimzadeh MA, Nabavi SF, Hamidinia A and Bekhradnia AR 2008 Determination of antioxidant activity, phenol and flavonoids content of *Parrotia persica* Mey. *Pharmacologyonline* **2** 560-567.
- Pejin B, Kojic V and Bogdanovic G 2014 An insight into the cytotoxic activity of phytol at in vitro conditions. *Nat Prod Res* **28** 2053-6.
- Rakholiya K, Kaneria M and Chanda S 2011 Vegetable and fruit peels as a novel source of antioxidants. *J Med Plant Res* **5** 63–71.
- Saha D, Talukdar A, Das T, Ghosh S K and Rahman H 2012 Evaluation of analgesic activity of ethanolic extract of *Cleodendrum serratum* Linn Leaves in Rats. *Int Res J Pharm Appl Sci* **2** 33-37.
- Santos CC de. MP, Salvadori MS, Mota V G, Costa L M, Almeida AAC de, Oliveira GAL de, Costa J P,

Sousa DPde, Freitas RMde and Almeida RNde 2013 Antinociceptive and antioxidant activities of phytol in-vivo and in-vitro models. *J Neuroscience Journal* 1-9.

Shukla R K, Painuly D, Porval A and Shukla A 2014 Proximate analysis, nutritional value, phytochemical evaluation, and biological activity of *Litch chinensis* Sonn. Leaves. *J Herbs Spices Med Plants* 20 196-208.

Shyam P and Suresh P K 2013 Comparative analysis of three leaf extracts of *Ixora coccinea* Linn for their protective and antioxidant potentials and correlation with analytical data. *Int J Pharma Bio Sci* 49 37-49.

Starlin T, Prabha P S, Thayakumar B K A and Gopalkrishnan VK 2019 Screening and GCMS profiling of ethanolic extract of *Tylophora pauciflora*. *J Biomed Inform* 15 425-429.

Steane DA, Scotland R W, Mabberley DJ, Wagstaff SJ, Reeves P A and Olmstead R G 1997 Phylogenetic relationships of *Clerodendrum serratum* L. (Lamiaceae) Inferred from chloroplast DNA. *Sys Bot* 22 229-243.

Tungmunnithum D, Thongboonyou A, Pholboon A and Yangsabai A 2018 Flavonoids and other phenolics compounds from medicinal plants for pharmaceutical and medical aspects: An overview. *Medicines* 5 2-16.

Wang Y, Zhang T Li, Feng Yi Xi, Guo SS, Pang X, Zhang D, Geng ZF and Du SS 2019 Insecticidal and repellent efficacy against stored-product insects of oxygenated monoterpenes and 2-dodecanone of the essential oil from *Zanthoxylum planispinum* var. *dintanensis*. *Environ Sci & Pollut Res* 26 24988-24997.



Nutrient Profiling, ICP-OES Mineral Analysis and UPLC Estimation of Amino Acids in Three Selected Ethnomedicinal Plants of North East India

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ABSTRACT

Background: Ethnomedicinal plants are rich traditional resources of Northeast India. Most of these plants are edible and are used as food by the tribal population of Northeast India. Quantitative nutritional analyses of three ethnomedicinal wild edible plant from Northeast India namely: *Zanthoxylum oxyphyllum* Edgew, *Rothea serrata* (L.) Steane and Mabb. and *Blumea lanceolaria* (Roxb.) Druce were carried out to identify the occurrence and abundance of nutrients, minerals and amino acids.

Methods: Proximate nutrients were estimated by standard AOAC methods. ICP-OES mineral analyzer was used to screen and quantify individual mineral elements. By performing UPLC analysis the presence and abundance of amino acids were investigated.

Result: Proximate analysis revealed that the plant species contains adequate amounts of protein, fibre and ash. From nutritional perspectives, low fat and carbohydrate in these species specifies that they are desirable diet for human nutrition. In regard to mineral content, the plant species were rich in macro minerals K, Ca, Na and Mg. Heavy metals were found within the recommended dietary permissible limits for humans. All the essential and non essential amino acids excluding cysteine, methionine and histidine were present in reasonable quantities. The plant species may be exploited to provide quality diets to mitigate nutrient deficiency. Adding together its nutritional and medicinal properties it may potentially be utilized in therapeutics. The further scope of the research involves ascertaining their potentiality in food-based strategies to perk up nutritional health.

Key words: Dietary, Medicinal, Nutritional, Therapeutics, Traditional, Wild edible.

INTRODUCTION

According to FAO at least two billion people, on earth, have been estimated to suffer from micronutrient deficiencies making them more susceptible to disease. Currently attention is focused on wild and natural food resources which if exploited can likely be a sustainable solution to mitigate nutrient deficiency. A large variety of natural resources are already present in North East India. Wild edible plants form an important part of the food culture in this region. Most importantly they contribute to the food and medicine baskets as well as livelihoods of tribal communities of Northeast India.

Among these, *Zanthoxylum oxyphyllum* Edgew a wild shrub indigenous to Northeast India is widely utilized as food and ethnomedicine in Northeast India. Tribal people in this region use the young leaves of this plant as vegetable. Tonics are prepared from the aqueous extracts of young *Z. oxyphyllum* leaves to treat a variety of diseases. According to Buragohain *et al.* (2011), *Z. oxyphyllum* leaves have blood purifying properties, can reduce the risk of leucoderma and help in solving stomach issues. The bark is used to treat leg pain, varicose veins, ulcers, rheumatism, hypotension, fevers and inflammatory conditions (Arun and Paridhavi, 2012).

Another plant, *Rothea serrata* (L.) Steane and Mabb. of Assam belongs to the family Lamiaceae. Locally, it is called as 'Nangal Bhanga' in Assamese. The use of *R. serrata* leaves has also been reported for the treatment of various diseases such as typhoid, cancer, jaundice and hypertension (Singh *et al.*, 2012). The leaves are also

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informed to have analgesic and anti-diabetic activity (Saha *et al.*, 2012; Kar *et al.*, 2014).

The third plant species discussed here is *Blumea lanceolaria* (Roxb.) Druce. Locally known as "Jwglaoori" in Assam is a tropical perennial herb belonging to the family Asteraceae. It is found in different parts of North-East India. In North East India, tribal people use this plant's young leaves as a vegetable. Traditionally, the leaves of the plant have also been used in the treatment of cough (Saikia *et al.*, 2017). In Mizoram of North East India, a decoction of the leaves is taken orally for the treatment of stomach ulcers, dysentery and wounds (Rai and Lalramnghinglova, 2010). The utility of *B. lanceolaria* in the treatment of ulcer and

cancer has also been mentioned by Sawmliana (2003) and Chawngkunga (2005).

Wild edible plants including *Alternanthera sessilis*, *Amaranthus species*, *Enhydra fluctuans*, *Houttuynia cordata*, *Centella asiatica*, *Ipomoea aquatic* and *B. lanceolaria* are used as food and in ethnomedicine in Northeast India (Bhattacharya *et al.*, 2001; Khakhalary *et al.*, 2022). These plant species are harvested from their native environments and sold in the markets of North East India.

It is postulated that plants contain nearly all of the nutritionally desirable micro and macronutrients including amino acids (Indrayan *et al.*, 2000). Onwordi *et al.* (2009) summarized that herbaceous vegetables contain good amounts of proteins, fats, carbohydrates, vitamins and minerals. The search for new natural therapeutic compounds which are less or non-toxic to humans, animals and the environment is a current research priority for scientists and it essentially begins with an emphasis on traditional, medical and veterinary procedures (Yeap *et al.*, 2010).

Vegetables are preferably consumed due to hunger, medicinal requirements and perceived nutrient benefits (Shumsky *et al.*, 2014). The tribal people in Northeast India claim that leaves of selected plant species holds medicinal properties as well as essential nutrients for human consumption. Therefore, this report provides a more comprehensive summary of the proximate, minerals and amino acid composition of an edible shrub of ethnomedical importance found in Northeast India. Studying their nutritional profiles will help in generating information about their potential nutritional benefits along with their medicinal properties.

MATERIALS AND METHODS

Collection of plant materials

Representative samples of *Z. oxyphyllum* Edgew, *R. serrata* (L.) Steane and Mabb and *B. lanceolaria* (Roxb.) Druce leaves were collected from Goalpara district of Assam, Northeast India. Herbarium of the collected samples were prepared and submitted to Gauhati University Botanical Herbarium (GUBH).

Sample preparation

Once in the laboratory the samples were cleaned and moisture content determined. The rest of the samples were shade dried, pulverized and stored in an airtight container for further analysis.

Estimation of proximate composition

Proximate composition analysis was performed in the Department of Biotechnology, Bodoland University, Kokrajhar in the year 2022. Moisture content was determined by oven-drying method, crude fat by extraction in Soxhlet apparatus, crude protein by micro Kjeldahl method, ash content by incineration in a muffle furnace, crude fibre content was estimated by treating the fat and moisture free sample with dilute acid and alkali and igniting the residue and carbohydrate content was calculated by subtracting the sum

of the percentages of moisture, fat, protein and ash from 100 (AOAC, 2005). Total solid was calculated using the method of James (1995). The energy content of plant sample was calculated by the method of FAO (2003).

Estimation of minerals

For mineral analysis 0.2 g of pulverized sample was digested with a mixture of 5 ml of 65% HNO₃ and 2 ml of 37% HCL. The sample was completely dissolved after the mixture was vortexed and boiled in a water bath (Ang and Lee, 2005). After filtering, the mixture was examined for the presence of Na, K, Mg, Ca, Mn, Zn, Fe, Cu, Ni and Cr using an ICP-OES mineral analyzer (Model No. Thermo Scientific TM iCAPTM 7600). Mineral estimation was carried out in the Sophisticated Analytical Instrumentation Facility (SAIF), NEHU, Shillong in the year 2022.

Determination of amino acid

Amino acid composition was determined by WATERS Acquity (make) Ultra Performance Liquid Chromatography. 1 mg of sample was taken in a clean glass tube to which 3 ml of 6N HCL was added and covered with paraffin. The tube was placed in the dry bath at 60°C under N₂ gas for 15 mins to maintain inertness. Then the temperature was increased at 110°C and incubated overnight. Derivatization was accomplished by combining 70 µl of Borate buffer and 20 µl of Accq Tag ultra-reagent to the sample and incubated for 10 mins at 55°C. After incubation 5 µl was loaded on to the instrument. Identification of amino acids was done by comparison to amino acid standard. Peaks were observed using 260 nm photodiode array detectors. Amino acid composition analysis was determined in Sandor Speciality Diagnostics Pvt. Ltd, Hyderabad in the year 2022.

Statistical analysis

Mean values and standard deviations were calculated using SPSS statistical software version 26.0 and the data were expressed as mean±standard deviation.

RESULTS AND DISCUSSION

Proximate composition

Results of proximate analysis of the three plant species are shown in Table 1. An attempt was made to compare the nutritional content of three most commonly consumed wild species. Findings showed that *B. lanceolaria* had the highest moisture (59.4±0.50 g/100 g) and fat (4±0.10 g/100 g) content among the samples analyzed. While analyzing the protein content of selected plants, it showed that *Z. oxyphyllum* had the highest protein content (24.30±0.34 g/100 g) followed by *R. serrata* (15.73±0.05 g/100 g) and *B. lanceolaria* (10.75±0.11 g/100 g). Among the proximate composition analyzed, *R. serrata* had the maximum amount of ash (12.02±0.06 g/100 g), fibre (17.43±0.05 g/100 g), carbohydrate (23.22±0.45 g/100 g), energy (189.1±1.04 kcal/100 g) as compared to *Z. oxyphyllum* and *B. lanceolaria*.

It has been reported by Ogie *et al.* (2001) that high moisture content promotes microorganism growth and

enzyme activity. Data shows that the moisture content was moderate and nearly identical in all the studied plant species (the values ranged from 44.97 g/100 g to 59.4 g/100 g). Thus, the presence of moderate moisture indicates its reasonable shelf life. Proteins are necessary for the formation of body tissues and regulating of compounds like hormones and enzymes (Akindahunsi and Salawu, 2005). Among the wild plants, the highest value of crude protein was found in *Z. oxyphyllum* (24.30 g/100 g). The protein content found in the samples exceeded the protein value reported by Tharmabalan (2021) in wild edible plants. The recommended dietary allowances (RDA) for protein for adult male and female set by ICMR-NIN (Indian Council of Medical Research-National Institute of Nutrition) (2020) is 54 g/d and 46 g/d respectively. The protein content in the studied sample can fulfill around 44% and 52% of daily requirement of protein in adult male and female at the maximum. Fats in foods are regarded as the primary source of energy but total fats also contains saturated and trans fatty acids which are undesirable for nutrition. Low fat foods are therefore preferred for human consumption. Fat content in the investigated samples is lower than that of other wild edible plants studied by Ullah *et al.* (2017). *Z. oxyphyllum* leaves has fat content of 2 g/100 g on dry weight because of which it can be marked as a low fat food and recommended to over weight or obese people as health food (Brahma *et al.*, 2014). According to Tunçturk and Özgökçe (2015), a plant's ash level is a good predictor of its overall mineral richness. Total ash content observed in this study corresponds to the findings of Aletan and Kwazo (2019) for wild edible plants. The data represented in Table 1 shows high amounts of ash indicating the presence of rich minerals desirable to our diet. In its 2002 report, the Institute of Medicine (IOM) set the recommended dietary allowances (RDA) for dietary fibre for adult to 25 gm per day. The recommended dietary allowances (RDA) set by ICMR-NIN (2020) for dietary fibre for adult male and female are 30 g/d and 25 g/d respectively. Fibre contents of all studied samples were high as compared to the report of Aletan and Kwazo (2019) and Tharmabalan (2021). The presence of high fibre lowers cholesterol level in the blood, reduces the risk of various cancers, bowel disease and improves general health and well being (Haub and Lattimer, 2010). Crude fibre content (14.43 g/100 g) in *Z. oxyphyllum*

can fulfill around half of the daily requirements for dietary fibre if consumed with other foods. The carbohydrate detected in all the samples was below the RDA values established by IOM (Institute of Medicines, 2002) in its reports. Food with low carbohydrate content is considered ideal for diabetic and hypertensive patients requiring low-sugar diets (Singha and Hassan, 2017). Results of the current study revealed that the selected plant species *Z. oxyphyllum*, *R. serrata* and *B. lanceolaria* could be used for nutritional purpose of human being due to their good nutritional qualities and adequate protection may be obtained against diseases arising from malnutrition. It also highlights the significance of wild vegetable species as cheap sources of nutrient for rural tribals.

Mineral composition

The plant species were analyzed for nine minerals of which one is heavy metals. Table 2, 3 and 4 represent the results of mineral composition of *Z. oxyphyllum*, *R. serrata* and *B. lanceolaria* (mg/100 g) dry matter. A comparison of mineral content in plants and their (%) fulfillment of recommended intake for particular nutrient as per the recommended dietary allowance (RDA) set by ICMR-NIN (2020) and Food Safety and Standards Authority of India, New Delhi is also shown in Table 2, 3 and 4. Results showed that selected plant species contain minerals like Na, K, Mg, Ca, Mn, Zn, Cu and Ni in varying amount. The plant species under investigation contained high amount of K and Ca with highest value recorded in *Z. oxyphyllum* and *B. lanceolaria* respectively. Na content was found equally in all the samples. Mg, Mn and Fe were of higher range in *R. serrata*, higher amount of Zn was detected in *Z. oxyphyllum* and Cu and Ni were highest in *B. lanceolaria*.

Macro minerals in *Z. oxyphyllum* including K, Ca, Na and Mg can fulfill about 0.12% to 4.13% of the RDA requirement in both human male and female (Table 2). On the contrary, micro nutrients such as Mn, Zn, Fe and Cu can mitigate nearly about 0.15% to 53.05% in male and 0.17% to 68.33% in female of the daily RDA. Heavy metals such as Ni can fulfil the RDA of 5% in both human male and female (Table 2). In *R. serrata*, macro minerals including K, Ca, Na and Mg can fulfill about 0.14% to 6.27% of the RDA in both human male and female. On the other hand, micro nutrients such as Mn, Zn, Fe and Cu can meet RDA of around

Table 1: Proximate composition of three ethnomedicinal plants (g/100 g) dry weight.

Parameters	<i>Z. oxyphyllum</i>	<i>R. serrata</i>	<i>B. lanceolaria</i>
Moisture	50.94±1.59	44.97±0.06	59.4±0.50
Total solid content	47.62±0.57	47.60±0.01	43.33±2.08
Protein	24.30±0.34	15.73±0.05	10.75±0.11
Fat	2.00±0.10	3.3±0.26	4±0.10
Ash	10.37±0.41	12.02±0.06	10.43±0.40
Fibre	14.43±0.52	17.43±0.05	15.46±0.35
Carbohydrate	10.98±0.12	23.22±0.45	15.61±0.32
Energy (kcal/100 g)	160.01±0.09	189.1±1.04	140.33±1.15

Note: Values are in triplicate mean±SD.

0.37% to 18.05% in male and 0.44% to 16.66% in female (Table 3). In *B. lanceolaria*, macro minerals including K, Ca, Na and Mg can fulfill about 0.14% to 6.63% of the RDA in both male and female. Among micro nutrients Mn, Zn, Fe and Cu can meet RDA of around 0.32% to 78.50% in male and 0.38 % to 78.50 % in female. Heavy metals such as Ni

can fulfil the RDA requirement of 47% in both male and female (Table 4).

Looking to the permissible levels set by the ICMR-NIN, 2020, the data obtained in the present study were within the specified limits demonstrating that a positive contribution of the mineral elements to the diet of consumers is provided

Table 2: Mineral profile of *Z. oxyphyllum* and recommended intake of essential minerals per day compared with *Z. oxyphyllum*.

Minerals	<i>Z. oxyphyllum</i> (mg/100 g)	Intake recommended for adults (mg per day)		% fulfilment of recommended intake	
		Male	Female	Male	Female
Na	2.57±0.02	2000	2000	0.12	0.12
K	104.7±0.20	3500	3500	2.99	2.99
Mg	4.39±0.09	400	310	1.09	1.41
Ca	41.38±0.01	1000	1000	4.13	4.13
Mn	0.66±0.01	440	370	0.15	0.17
Zn	9.02±0.02	17	13.2	53.05	68.33
Fe	3.08±0.02	19	29	16.21	10.62
Cu	0.06±0.02	2	2	3	3
Ni	0.05±0.03	1.0	1.0	5	5

Note: Values are in triplicate mean±SD. The data for recommended dietary allowances and estimated average requirements for Indians are as provided by Indian Council of Medical Research (ICMR).

Table 3: Mineral profile of *R. serrate* and recommended intake of essential minerals per day compared with *R. serrate*.

Minerals	<i>R. serrate</i> (mg/100 g)	Intake recommended for adults (mg per day)		% fulfilment of recommended intake	
		Male	Female	Male	Female
Na	2.90±0.1	2000	2000	0.14	0.14
K	47.76±0.04	3500	3500	1.36	1.36
Mg	6.53±0.32	400	310	1.63	2.10
Ca	62.73±0.20	1000	1000	6.27	6.27
Mn	1.65±0.05	440	370	0.37	0.44
Zn	2.20±0.02	17	13.2	12.94	16.66
Fe	3.43±0.40	19	29	18.05	11.82
Cu	0.09±0.01	2	2	4.5	4.5
Ni	0.01±0.02	1.0	1.0	1	1

Note: Values are in triplicate mean±SD. The data for recommended dietary allowances and estimated average requirements for Indians are as provided by Indian Council of Medical Research (ICMR).

Table 4: Mineral profile of *B. lanceolaria* and recommended intake of essential minerals per day compared with *B. lanceolaria*.

Minerals	<i>B. lanceolaria</i> (mg/100 g)	Intake recommended for adults (mg per day)		% fulfilment of recommended intake	
		Male	Female	Male	Female
Na	2.92±0.05	2000	2000	0.14	0.14
K	44.80±0.18	3500	3500	1.28	1.28
Mg	5.30±0.04	400	310	1.32	1.70
Ca	66.37±0.06	1000	1000	6.63	6.63
Mn	1.43±0.04	440	370	0.32	0.38
Zn	5.17±0.03	17	13.2	30.41	39.16
Fe	1.39±0.02	19	29	7.31	4.79
Cu	1.57±0.05	2	2	78.50	78.50
Ni	0.47±0.03	1.0	1.0	47	47

Note: Values are in triplicate mean±SD. The data for recommended dietary allowances and estimated average requirements for Indians are as provided by Indian Council of Medical Research (ICMR).

by these ethno medicinally important plants. Na and K are two essential macro minerals required by the body to maintain cellular homeostasis, metabolism and many other functions. The Na/K ratio in our body is very important to control high blood pressure and the ratio should be less than one (Akubugwo *et al.*, 2007). In the present study, all the samples have the Na/K ratio less than one that indicate the consumption of these vegetables are helpful for human and might be able to control the high blood pressure of our body. Low Na and high K intake also aid in the prevention of hypertension and atherosclerosis (Saupi *et al.*, 2009). Na and K of *Clerodendrum colebrookianum* recorded by Payum, (2020) is lower as compared to this study. The role of Mg and Ca in maintenance of heart function has been well pointed out by Insel *et al.* (2010). Mg content detected in the samples is high than Payum, (2020) in *Clerodendrum colebrookianum*. An analysis showed the percentage fulfillment of Ca is found high among macro minerals in all samples with the highest value recorded in *B. lanceolaria*. Ca content recorded by Njoku *et al.* (2021) from *Urena lobata* stems is comparable to the Ca content of *R. serrata* and *B. lanceolaria*. The data indicates that these wild vegetables could be a good source of Ca to our diet and provide health benefits by lowering the risk of such diseases. Through analysis, Mn and Zn content found in all the samples is lower than the recommended limit. Mn and Zn content studied by Payum (2020) in *Clerodendrum colebrookianum* is lower than the present study. Similarly, Zn content detected by Islary *et al.* (2016) in *Grewia sapida* fruit is also low. Demirezen and Ahmet (2006) analyzed various vegetables and reported that the Zn concentrations

in vegetables are within the recommended international standards. All analyzed samples possess an adequate amount of Fe and is higher than the study conducted by Payum, (2020) in *Clerodendrum colebrookianum*. Fe content of *Z. oxyphyllum* and *R. serrata* is comparable to Njoku *et al.* (2021) detected in *Urena lobata* stems. The percentage of RDA for Fe recorded shows that *R. serrata* has the highest percent fulfillment in comparison to *Z. oxyphyllum* and *B. lanceolaria* in both male and female. Cu an essential trace element in human body has an important role in oxidation reduction reactions and in scavenging of free radicals (Linder and Azam, 1996). When its concentration exceeds the safe limit, it can be toxic in some cases (Ogwok *et al.*, 2014). Among the analyzed samples, the per cent fulfillment of recommended intake for Cu was found high in *B. lanceolaria*. A trace quantity of Ni may promote healthy skin, iron metabolism and optimal growth in humans, but it can be toxic when its concentration exceeds the safe limit (Satter *et al.*, 2016). The percentage of RDA recorded for Ni was found high in *B. lanceolaria*.

Amino acid composition

Amino acid composition of the plant species and its percent fulfillment of the RDA requirement in adults as given by ICMR-NIN (2020) are presented in Table 5. UPLC chromatograms of standard and samples are shown in Fig 1, 2, 3 and 4. Both essential and non essential amino acids were found in the samples in varying quantities. Cysteine, among the nonessential amino acid was not detected in any of the plants. Among essential amino acids, histidine was not found in both *Z. oxyphyllum* and *R. serrata*. In *R. serrata*, methionine was not detected. Of the essential

Table 5: Amino acid composition of studied plants (mg/100 g) dry weight.

Amino acid	<i>Z. oxyphyllum</i>	<i>R. serrata</i>	<i>B. lanceolaria</i>	RDA (mg/100 g)
Essential amino acid				
Thr	2.90±0.02	ND	0.64±0.01	1.5
Val	2.57±0.02	0.19±0.01	0.66±0.02	2.6
Met	0.59±0.01	ND	0.70±0.01	1
Ile	1.28±0.005	0.17±0.005	0.47±0.02	2
Leu	4.51±0.03	0.84±0.001	1.12±0.02	3.9
Phe	7.08±0.02	1.76±0.03	5.77±0.03	NL
His	ND	ND	0.28±0.005	1
Lys	3.92±0.06	0.39±0.02	0.90±0.01	3
Arg	7.47±0.03	0.30±0.01	1.21±0.02	NL
Non-essential amino acid				
Asp	2.37±0.02	0.28±0.01	2.21±0.03	NL
Ser	3.02±0.01	0.83±0.02	0.28±0.005	NL
Glu	1.07±0.01	1.50±0.01	1.83±0.02	NL
Pro	6.63±0.02	0.91±0.01	0.70±0.01	NL
Gly	5.37±0.03	1.09±0.01	0.68±0.02	NL
Cys	ND	ND	ND	4
Ala	3.28±0.02	0.53±0.05	0.89±0.01	NL
Tyr	4.49±0.01	0.64±0.02	1.85±0.03	NL

Note: Values are in triplicate mean±SD. Recommended dietary allowances and estimated average requirements for Indians-2020 for age group of more than 24 months (2 years). "NL" indicates "not listed".

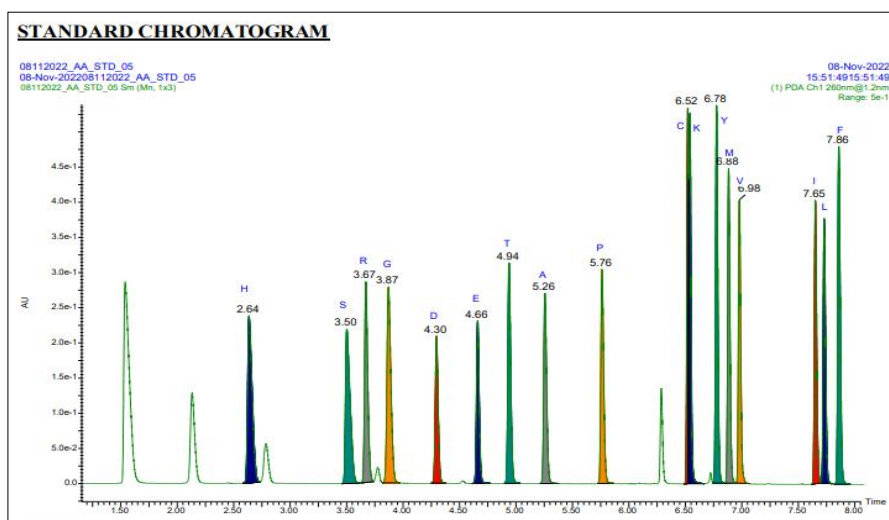


Fig 1: UPLC chromatogram of amino acid of standard.

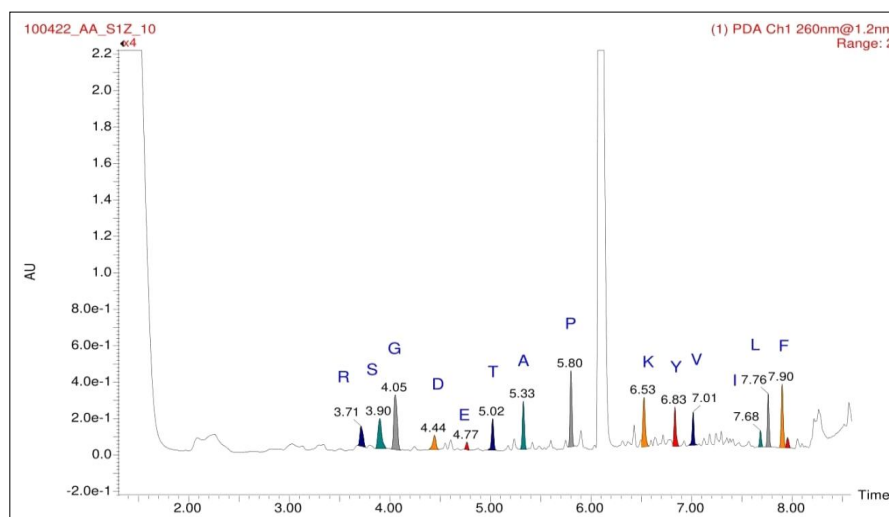


Fig 2: UPLC chromatogram of amino acid present in *Z. oxyphyllum*.

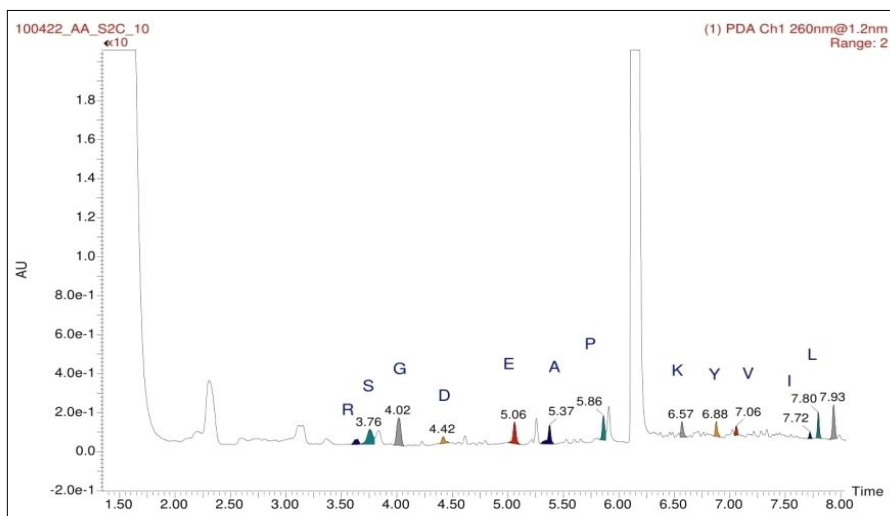


Fig 3: UPLC chromatogram of amino acid present in *R. serrata*.

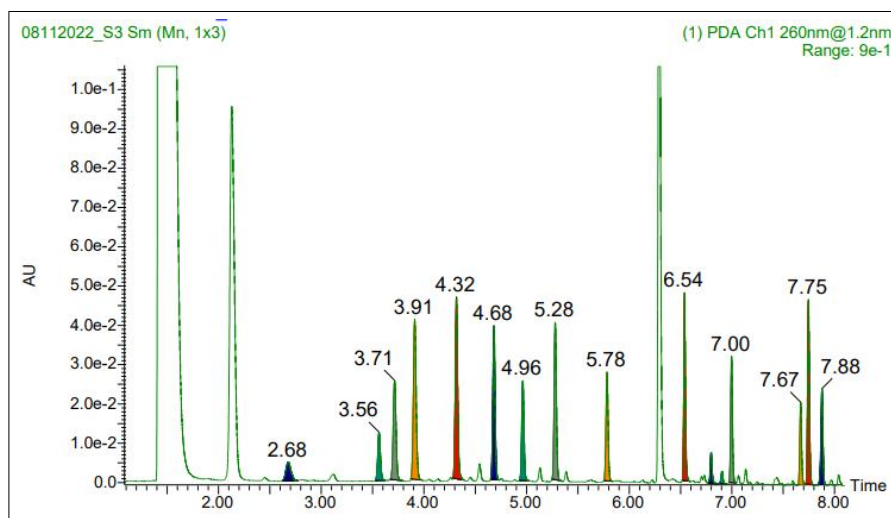


Fig 4: UPLC chromatogram of amino acid present in *B. lanceolaria*.

amino acid, leucine, threonine and lysine were detected slightly higher than RDA value. The UL (Upper limit) sets the experimentally determined safe upper limits for the highest average daily nutrient intake level that is likely to pose no risk of adverse health effects to almost all individuals in the general population as mentioned by National Academy of Medicine (2005). Elango (2023) reported that in some situations, amino acids above normal consumption amounts are considered beneficial such as exercise performance, recovery from injury, *etc.* The UL for the current adult leucine intake recommendations kept RDA of 2.9 g/d (Elango, 2023). RDA recommendations are “minimum” dietary intakes to prevent deficiency and considered adequate to meet 50% and 97.5% of the needs, respectively in a population. With nutrients being consumed in excess of the body's needs, there is a possibility of reaching intake levels where adverse effects can occur. Amino acids are precursors for the synthesis of secondary metabolites that have physiological beneficial effects in our bodies. They are essential components for healing processes and a lack of these components impedes recovery. Aside the structural functions, amino acids could also serve as valuable sources of energy especially in the absence of carbohydrate and fats in the body (Olusanya, 2008). Amino acids composition reveals information about the quality of food proteins. Collectively the present findings have clearly shown the presence of most essential and non essential amino acids indicating the studied plants can potentially contribute in mitigating protein deficiency. It also implicates the possibility of incorporating the studied plant material into the modern health care system through providing useful information for further application of these plants.

CONCLUSION

Findings of the current study provide an overview of nutrient composition of three wild edible ethnomedicinal plants of

Northeast India. It contains significant proportions of proximate nutrients, minerals and amino acids required for human nutrition and health in addition to its medicinal properties. Both essential and non-essential amino acids were found. Result availed from this study may be incorporated into the National Food Composition Database of India to enhance its food value.

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Conflict of interest

All authors declared that they have no conflict of interest.

REFERENCES

- Akindahunsi, A.A. and Salawu, S.O. (2005). Phytochemical screening and nutrient-antinutrient composition of selected tropical green vegetables. *African Journal of Biotechnology*. 4(6): 497-501.
- Akubugwo, I.E., Obasi, N.A., Chinyere, G.C., Ugbogu, A.E. (2007). Nutritional and chemical value of *Amaranthus hybridus* L. leaves from Afikpo, Nigeria. *African Journal of Biotechnology*. 6(24): 2833-2839.
- Aletan, U.I. and Kwazo, H.A. (2019). Analysis of the proximate composition, anti-nutrients and mineral content of *Maerua crassifolia* leaves. *Nigerian Journal of Basic and Applied Science*. 27(1): 89-96.
- Ang, H. and Lee, K. (2005). Analysis of mercury in Malaysian herbal preparations: A peer review. *Biomedical Science*. 4(1): 31-36.

- Arun, K.K.V. and Paridhavi, M. (2012). An ethnobotanical phytochemical and pharmacological utilization of widely distributed species *Zanthoxylum*: A comprehensive overview. *International Journal of Pharmaceutical Innovation*. 2(7): 24-35.
- AOAC (2005). *Official Methods of Analysis*. (18th Edn). Association of Official Analytical Chemists, Washington DC. pp. 935.14 and 992.24.
- Bhattacharya, R.K. (2001). *Horticultural Crops-fruits and Vegetables*, In: *Agriculture in Assam*, by [Thakur, A.C., Bhattacharya, A. and Sarma, D.K.], (Assam Agricultural University, Jorhat, Assam), pp. 104-113.
- Brahma, J., Singh, B., Rethy, P., Gajurel, P. (2014). Nutritional analysis of some selected wild edible species consumed by the Bodos Tribes of Kokrajhar District, BTC. Assam. *Asian Journal of Pharmaceutical and Clinical Research*. 7(3): 34-37.
- Buragohain, J., Konwar, B.K., Bordoloi, M.J. (2011). Isolation of antimicrobial compounds from the tender shoots of *Zanthoxylum*. *Der Pharmacia Sinicia*. 2(6): 149-152.
- Chawngkung, C. (2005). Medicinal plants used as healing agents by the ethnic people of Mizoram, Aizawl. *Journal of Biology and Life Science*. 4(2): 95-102.
- Demirezen, D. and Ahmet, A. (2006). Heavy metal levels in vegetables in Turkey are within safe limits for Cu, Zn, Ni and exceeded for Cd and Pb. *Journal of Food Quality*. 29(3): 252-265.
- Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein and Amino Acids. (2005). National Academy of Medicine.
- Elango, R. (2023). Tolerable upper intake level for individual amino acids in humans: A narrative review of recent clinical studies. *Advances in Nutrition*. 14(4): 885-894.
- FAO, (2003). *Food Energy Methods of Analysis and Conversion Factors*. Food and Nutrition Paper 77, Food and Agriculture Organization of the United Nations, Rome, Italy. 1-93.
- Haub, M.D. and Lattimer, J.M. (2010). Effects of dietary fiber and its components on metabolic health. *Nutrient*. 2(12): 1266-1289.
- ICMR (Indian Council of Medical Research) - NIN, (2020). Values for nutrients except amino acids. Food Safety and Standards Authority of India. F. No. Stds/SP-05/Orders/FSSAI.
- Indrayan, A.K., Shama, S.D., Durgapal, N., Kumar, N., Kumar, M. (2000). Determination of nutritive value and analysis of mineral elements for some medicinally valued plants from Uttarakhand. *Current Science*. 89(7): 1252-1253.
- Insel, P.M., Ross, D., McMahon, K., Bernstein, M. (2010). *Nutrition*. (4th edition) Jones and Bartlett Publishers, Sudbury, MA. pp. 1002.
- Institute of Medicine, Food and Nutrition Board, (2002). *Dietary Reference Intakes: Energy, carbohydrates, fiber, fat, fatty acids, cholesterol, protein and amino acids*. Washington: National Academics Press.
- Islary, A., Sarmah, J., Basumatary, J. (2016). Proximate composition, mineral content, phytochemical analysis and *in vitro* antioxidant activities of a wild edible fruit (*Grewia sapida* Roxb. ex DC.) found in Assam of North-East India. *Journal of Investigational Biochemistry*. 5(1): 21-31.
- James, C.S. (1995). *Analytical Chemistry of Food*. (1st Edn.) Chapman and Hall, New York. pp. 256-257.
- Kar, M.K., Swain, T.R., Mishra, S.K. (2014). Antidiabetic activity of *Clerodendrum serratum* (L.) moon leaves in streptozotocin-induced diabetic rats. *Asian Journal of Pharmaceutical and Clinical Research*. 7(5): 260-263.
- Khakhalary, S., Narzari, S., Sarmah, J., Paul, M., Dutta, T. (2022). Chromatographic analysis of *Blumea lanceolaria* (Roxb.) Druce leaf extract: A traditional herbal drug of North East India. *Applied Biological Research*. 24(1): 96-104.
- Linder, M.C. and Azam, M.H. (1996). Copper biochemistry and molecular biology. *American Journal of Clinical Nutrition*. 63(5): 797S-811S.
- Njoku, C.E., Alaneme, K.K., Omotoyinbo, J.A., Ekeleme, A.C., Ugwu, E.I., Ikele, U.S. (2021). Phytochemical, proximate and mineral analyses of *Urena lobata* stems from Imo State Nigeria. *International Conference on Engineering for Sustainable World (ICESW 2020)*. 1-13.
- Ogie, B.M., Ha-thi, A.D., Mulokozi, G., Hambraeus, L. (2001). Micronutrient composition and nutritional importance of gathered vegetables in Vietnam. *International Journal of Food Science Nutrition*. 52(6): 485-499.
- Ogwok, P., Bamuwamye, M., Apili, G., Musalima, J.H. (2014). Health risk posed by lead, copper and iron *via* consumption of organ meats in Kampala City (Uganda). *Journal of Environment Pollution and Human Health*. 2(3): 69-73.
- Olusanya, J. O. (2008). *Proteins*, In: *Essential of Food and Nutrition*. Apex Books Limited. pp. 13-21.
- Onwordi, C.T., Ogungbade, A.M., Wusu, A.D. (2009). The proximate and mineral composition of three leafy vegetables commonly consumed in Lagos, Nigeria. *African Journal of Pure and Applied Chemistry*. 3(6): 102-107.
- Payum, T. (2020). Phytoconstituents and proximate composition of *clerodendrum colebrookianum* walp: A widely used anti high blood pressure medicinal food plant in Eastern Himalayas. *Pharmacognosy Journal*. 12(6): 1534-1540.
- Rai, P.K. and Lalramnghinglova, H. (2010). Ethnomedicinal plant resources of Mizoram, India. Implication of traditional knowledge in health care system. *Ethnobotanical Leaflets*. 14(3): 274-305.
- Saha, D., Talukdar, A., Das, T., Ghosh, S.K., Rahman, H. (2012). Evaluation of analgesic activity of ethanolic extract of *Cleodendrum serratum* linn leaves in rats. *International Research Journal of Pharmaceutical and Applied Science*. 2(6): 33-37.
- Saikia, K., Lalawmpui, R., Borgohain, P. (2017). Evaluation of *in vitro* antioxidant and cytotoxic activity of methanolic leaf extract of *Blumea lanceolaria* Roxb. *International Journal of Technical Research and Applications*. 5(6): 54-57.
- Satter, M.M.A., Khan, M.M.R.L., Jabin, S.A., Abedin, N., Islam, M.F., Shaha, B. (2016). Nutritional quality and safety aspects of wild vegetables consume in Bangladesh. *Asian Pacific Journal of Tropical Biomedicine*. 6(2): 125-131.
- Saupi, N., Zakaria, M.H., Bujang, J.S. (2009). Analytic chemical composition and mineral content of yellow velvet leaf [*Limnocharis flava* (L.) Buchenau]'s edible parts. *Journal of Applied Science*. 9(16): 2969-2974.
- Sawmliana, M. (2003). *The Book of Mizoram Plants*. 1st Ed. Lois Bet, Chandmari, Aizawl. pp. 114.

- Shumsky, S.A., Hickey, G.M., Pelletier, B., Johns. T. (2014). Understanding the contribution of wild edible plants to rural social ecological resilience in semi-arid Kenya. *Ecology and Society*. 19(4): 34-55.
- Singh, M. K., Khare, G., Iyer, S.K., Sharwan, G., Tripathi, D.K. (2012). *Clerodendrum serratum*: A clinical approach. *Journal of Applied Pharmaceutical Science*. 2(2): 11-15.
- Singha, B.L. and Hassan, Y. (2017). Analysis of carbohydrate, moisture content and specific gravity of bambusa tulda with special reference to its harvesting season. *International Journal of Science and Research*. 6(1): 279-283.
- Tharmabalan, R.T. (2021). Nutritional analysis of five wild edible vegetables traditionally consumed by the Orang Asli in Perak. *International Journal of Food Science*. 71(3): 1-7.
- Tuncturk, M. and Ozgokce, F. (2015). Chemical composition of some Apiaceae plants commonly used in herby cheese in Eastern Anatolia. *Turkish Journal of Agriculture and Forestry*. 39(1): 55-62.
- Ullah, I., Gul, S., Rehman, H.U., Ahmad, N., Ullah, I., Aziz-ud-Din., Jawad, S.M., Ali, J., Ahmad, A., Akbar, M.U. (2017). Analysis of nutrients and minerals of some edible plants. *International Journal of Fauna and Biological Studies*. 4(6): 35-39.
- Yeap, S.K., Ho, W.Y., Beh, B.K., Liang, W.S., Ky, H., Yousef, A.H.N., Alitheen, N.B. (2010). *Vernonia amygdalina*, an ethnoveterinary and ethnomedicinal used green vegetable with multiple bioactivities. *Journal of Medicinal Plants Research*. 4(1): 2787-2812.