

REFERENCES

REFERENCES

- Abid, S., & Khan, A. H. (2002). Severe hemolysis and renal failure in glucose-6-phosphate dehydrogenase deficient patients with hepatitis E. *The American Journal of Gastroenterology*, *97*(6), 1544–1547.
- Agarwal, A., Guindo, A., Cissoko, Y., Taylor, J. G., Coulibaly, D., Koné, A., Kayentao, K., Djimde, A., Plowe, C. V., Doumbo, O., Wellems, T. E., & Diallo, D. (2000). Hemoglobin C associated with protection from severe malaria in the Dogon of Mali, a West African population with a low prevalence of hemoglobin S. *Blood*, *96*(7), 2358–2363.
- Agarwal, M. B., Varandani, D. G., Joshi, R. H., Bhargava, A. B., & Mehta, B. C. (1981). Haemoglobin D disorders in 13 unrelated families. *Indian Journal of Medical Research*, *73*, 554–557.
- Ahluwalia, A., Corcoran, C. M., Vulliamy, T. J., Ishwad, C. S., Naidu, J. M., Argusti, A., Stevens, D. J., Mason, P. J., & Luzzatto, L. (1992). G6PD Kalyan and G6PD Kerala; two deficient variants in India caused by the same 317 Glu-->Lys mutation. *Human Molecular Genetics*, *1*(3), 209–210.
- Ainoon, O., Boo, N. Y., Yu, Y. H., Cheong, S. K., Hamidah, H. N., & Lim, J. H. (2004). Complete molecular characterisation of Glucose-6-phosphate dehydrogenase (G6PD) deficiency in a group of Malaysian Chinese neonates. *Malaysian Journal of Pathology*, *26*(2), 89–98.
- Ainoon, O., Yu, Y. H., Amir Muhriz, A. L., Boo, N. Y., Cheong, S. K., & Hamidah, N. H. (2003). Glucose-6-phosphate dehydrogenase (G6PD) variants in Malaysian Malays. *Human Mutation*, *21*(1), 101.

- Ajlaan, S. K., al-Naama, L. M., & al-Naama, M. M. (2000). Correlation between normal glucose-6-phosphate dehydrogenase level and haematological parameters. *Eastern Mediterranean Health Journal*, 6(2-3), 391–395.
- Allen, S. J., O'Donnell, A., Alexander, N. D., Alpers, M. P., Peto, T. E., Clegg, J. B., & Weatherall, D. J. (1997). Alpha+-Thalassemia protects children against disease caused by other infections as well as malaria. *Proceedings of the National Academy of Sciences of the United States of America*, 94(26), 14736–14741.
- Altay, C., & Gumruk, F. (2008). Red cell Glucose-6-phosphate dehydrogenase deficiency in Turkey. *Turkish Journal of Haematology*, 25(1), 1–7.
- Amjad, F., Fatima, T., Fayyaz, T., Khan, M. A., & Qadeer, M. I. (2020). Novel genetic therapeutic approaches for modulating the severity of β -thalassemia (Review). *Biomedical Reports*, 13(5), 48.
- Arese, P., Gallo, V., Pantaleo, A., & Turrini, F. (2012). Life and Death of Glucose-6-Phosphate Dehydrogenase (G6PD) Deficient Erythrocytes - Role of Redox Stress and Band 3 Modifications. *Transfusion Medicine and Hemotherapy*, 39(5), 328–334.
- Au, S. W., Gover, S., Lam, V. M., & Adams, M. J. (2000). Human glucose-6-phosphate dehydrogenase: the crystal structure reveals a structural NADP(+) molecule and provides insights into enzyme deficiency. *Structure*, 8(3), 293–303.
- Awab, G. R., Aaram, F., Jamornthanyawat, N., Suwannasin, K., Pagornrat, W., Watson, J. A., Woodrow, C. J., Dondorp, A. M., Day, N. P., Imwong, M., & White, N. J. (2021). Protective effect of Mediterranean-type glucose-6-phosphate dehydrogenase deficiency against *Plasmodium vivax* malaria. *eLife*, 10, e62448.
- Awamy B. H. (1992). Effect of G6PD deficiency on sickle cell disease in Saudi Arabia. *Indian Journal of Pediatrics*, 59(3), 331–334.

- Azevedo, E., Kirkman, H. N., Morrow, A. C., & Motulsky, A. G. (1968). Variants of red cell glucose-6-phosphate dehydrogenase among Asiatic Indians. *Annals of Human Genetics*, 31(4), 373–379.
- Babu, B. V., Sridevi, P., Surti, S., Ranjit, M. R., Bhat, D., Sarmah, J., Sudhakar, G., & Sharma, Y. (2021). Prevalence of sickle cell disease among children of tribal population in India: Feasibility of screening at community level in low-resource settings. *Pediatric Blood & Cancer*, 68(6), e28911.
- Badar, M. S., Shamsi, S., Ahmed, J., Alam, M. A. (2022). Molecular Dynamics Simulations: Concept, Methods, and Applications. In: Rezaei, N. (Ed.) *Transdisciplinarity. Integrated Science*, vol 5. Springer, Cham.
- Badoum, E. S., Serme, S. S., Yaro, J. B., Coulibaly, S. A., Kargougou, D., Diarra, A., Ouédraogo, A. Z., Malik, L., Nebie, I., Soulama, I., Ouédraogo, A., Tiono, A. B., Traoré, Y., Sirima, S. B., & Bougouma, E. C. (2019). Abnormalities of hemoglobin and Glucose-6-phosphate-dehydrogenase deficiency in children with uncomplicated malaria and living in Banfora and Sapone, two different malaria setting of Burkina Faso. *International Journal of Tropical Disease and Health*, 37, 1–10.
- Balgir R. S. (2006). Do tribal communities show an inverse relationship between sickle cell disorders and glucose-6-phosphate dehydrogenase deficiency in malaria endemic areas of Central-Eastern India?. *Homo*, 57(2), 163–176.
- Balgir R. S. (2008). Hematological profile of twenty-nine tribal compound cases of hemoglobinopathies and G-6-PD deficiency in rural Orissa. *Indian Journal of Medical Sciences*, 62(9), 362–371.
- Balgir, R. S. (1996). Genetic epidemiology of the three predominant abnormal haemoglobins in India. *Journal of the Association of Physicians of India*, 44(1), 25–28.

- Bancone, G., & Chu, C. S. (2021). G6PD Variants and Haemolytic Sensitivity to Primaquine and Other Drugs. *Frontiers in Pharmacology*, *12*, 638885.
- Bancone, G., Chu, C. S., Somsakchaicharoen, R., Chowwiwat, N., Parker, D. M., Charunwatthana, P., White, N. J., & Nosten, F. H. (2014). Characterization of G6PD genotypes and phenotypes on the northwestern Thailand-Myanmar border. *PloS One*, *9*(12), e116063.
- Bancone, G., Menard, D., Khim, N., Kim, S., Canier, L., Nguong, C., Phommasone, K., Mayxay, M., Dittrich, S., Vongsouvath, M., Fievet, N., Le Hesran, J. Y., Briand, V., Keomany, S., Newton, P. N., Gorsawun, G., Tardy, K., Chu, C. S., Rattanapalroj, O., Dong, L. T., Quang, H. H., Tam-Uyen, N., Thuy-Nhien, N., Hien, T. T., Kalnoky, M., & Nosten, F. (2019). Molecular characterization and mapping of glucose-6-phosphate dehydrogenase (G6PD) mutations in the Greater Mekong Subregion. *Malaria Journal*, *18*(1), 20.
- Barman, L., Sharma, A., Kakati, S., Sarma, D. K., Hussain, E., & Saikia, L. (2023). Molecular detection of drug-resistant *Plasmodium falciparum* mutants in Assam. *Indian Journal of Medical Research*, *158*(1), 55–65.
- Barrera-Reyes, P. K., & Tejero, M. E. (2019). Genetic variation influencing hemoglobin levels and risk for anemia across populations. *Annals of the New York Academy of Sciences*, *1450*(1), 32–46.
- Basumatary, N., Baruah, D., Sarma, P. K., Wary, K. K., & Sarmah, J. (2023). The first report of three Glucose-6-phosphate dehydrogenase (G6PD) variants: Mediterranean, Orissa and Kalyan-Kerala from Northeast India. *Indian Journal of Hematology and Blood Transfusion*, <https://doi.org/10.1007/s12288-023-01686-7>
- Baxi, A. J., Balakrishnan, V., Undevia, J. V., & Sanghvi, L. D. (1963). Glucose-6-phosphate dehydrogenase deficiency in the Parsee community, Bombay. *Indian Journal of Medical Sciences*, *17*, 493–500.

- Bekker, H., Berendsen, H.J.C., Dijkstra, E.J., Achterop, S., van Drunen, R., van der Spoel, D., Sijbers, A., & Keegstra H. (1993). Gromacs: A parallel computer for molecular dynamics simulations; pp. 252–256 in *Physics computing 92*. Edited by R.A. de Groot and J. Nadrchal. World Scientific, Singapore, 1993.
- Beutler, E. (2007). Glucose-6-phosphate dehydrogenase deficiency: a historical perspective. *Blood*, *111*(1), 16–24.
- Beutler, E. (1996). G6PD: Population genetics and clinical manifestations. *Blood Reviews*, *10*(1), 45–52.
- Beutler, E., Westwood, B., and Kuhl, W. (1991). Definition of the mutations of G6PD Wayne, G6PD Viangchan, G6PD Jammu and G6PD “LeJeune”. *Acta Haematologica*, *86*(4), 179–182.
- Beutler, E., Westwood, B., Kuhl, W., & Hsia, Y. E. (1992). Glucose-6-phosphate dehydrogenase variants in Hawaii. *Human Heredity*, *42*(5), 327–329.
- Bharti, R. S., Vashisht, K., Ahmed, N., Nayak, A., Pande, V., & Mishra, N. (2020). First report of glucose-6-phosphate dehydrogenase (G6PD) variants (Mahidol and Acores) from malaria-endemic regions of northeast India and their functional evaluations in-silico. *Acta Tropica*, *202*, 105252.
- Bhasin, M. K. (2006). Genetics of castes and tribes of India: Glucose-6-phosphate dehydrogenase deficiency and abnormal hemoglobins (HbS and HbE). *International Journal of Human Genetics*, *6*(1), 49–72.
- Bhatia, H. M., & Rao, V. R. (1987). Genetic atlas of the Indian tribes. *Institute of Immunohaematology, Indian Council of Medical Research, Bombay*.
- Bickerton, G. R., Paolini, G. V., Besnard, J., Muresan, S., & Hopkins, A. L. (2012). Quantifying the chemical beauty of drugs. *Nature chemistry*, *4*(2), 90–98.
- Bordoloi, B. N., Thakur, G. C. S., & Saikia, M. C. (1987). Tribes of Assam Part I. *Tribal Research Institute, Assam*.

- Bouanga, J. C., Mouélé, R., Préhu, C., Wajcman, H., Feingold, J., & Galactéros, F. (1998). Glucose-6-phosphate dehydrogenase deficiency and homozygous sickle cell disease in Congo. *Human Heredity*, 48(4), 192–197.
- Bubp, J., Jen, M., & Matuszewski, K. (2015). Caring for Glucose-6-Phosphate Dehydrogenase (G6PD)-Deficient Patients: Implications for Pharmacy. *P & T*, 40(9), 572–574.
- Cai, W., Filosa, S., Martini, G., Zhou, Y., Zhou, D., Cai, L., and Kuang, Y. (2001). Molecular characterization of Glucose-6-phosphate dehydrogenase deficiency in the Han and Li nationalities in Hainan, China and identification of a new mutation in human G6PD gene. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi*, 18(2), 105–109.
- Camacho, L. H., Wilairatana, P., Weiss, G., Mercader, M. A., Brittenham, G. M., Looareesuwan, S., & Gordeuk, V. R. (1999). The eosinophilic response and haematological recovery after treatment for *Plasmodium falciparum* malaria. *Tropical Medicine & International Health*, 4(7), 471–475.
- Cao, A., and Kan, Y. W. (2013). The prevention of thalassemia. *Cold Spring Harbor Perspectives in Medicine*, 3(2), a011775.
- Cappellini, M. D., and Fiorelli, G. (2008). Glucose-6-phosphate dehydrogenase deficiency. *Lancet*, 371(9606), 64–74.
- Cayanis, E., Lane, A. B., Jenkins, T., Nurse, G. T., & Balinsky, D. (1977). Glucose-6-phosphate dehydrogenase Porbandar: a new slow variant with slightly reduced activity in a South African family of Indian descent. *Biochemical Genetics*, 15(7-8), 765–773.
- Census of India. (2011). Office of the Registrar General and Census Commissioner. Ministry of Home Affairs, Govt. of India. Available at <http://www.censusindia.gov.in>, Retrieved on November 15, 2023.

- Chalvam, R., Kedar, P. S., Colah, R. B., Ghosh, K., & Mukherjee, M. B. (2008). A novel R198H mutation in the Glucose-6-phosphate dehydrogenase gene in the tribal groups of the Nilgiris in Southern India. *Journal of Human Genetics*, 53(2), 181–184.
- Chalvam, R., Mukherjee, M. B., Colah, R. B., Mohanty, D., & Ghosh, K. (2007). G6PD Namoru (208T-C) is the major polymorphic variant in the tribal populations in Southern India. *British Journal of Haematology*, 136(3), 512–513.
- Chan, T. K., Todd, D., & Tso, S. C. (1976). Drug-induced haemolysis in Glucose-6-phosphate dehydrogenase deficiency. *British Medical Journal*, 2(6046), 1227–1229.
- Chao, L. T., Du, C. S., Louie, E., Zuo, L., Chen, E., Lubin, B., & Chiu, D. T. (1991). A to G substitution identified in exon 2 of the G6PD gene among G6PD deficient Chinese. *Nucleic Acids Research*, 19(21), 6056.
- Charoenkwan, P., Tantiprabha, W., Sirichotiyakul, S., Phusua, A., & Sanguansermisri, T. (2014). Prevalence and molecular characterization of Glucose-6-phosphate dehydrogenase deficiency in northern Thailand. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 45(1), 187–193.
- Chau, T. N., Lai, S. T., Lai, J. Y., & Yuen, H. (1997). Haemolysis complicating acute viral hepatitis in patients with normal or deficient glucose-6-phosphate dehydrogenase activity. *Scandinavian Journal of Infectious Diseases*, 29(6), 551–553.
- Chen, H. L., Huang, M. J., Huang, C. S., & Tang, T. K. (1996). G6PD NanKang (517 T->C; 173 Phe->Leu): A new Chinese G6PD variant associated with neonatal jaundice. *Human Heredity*, 46(4), 201–204.
- Chen, H. L., Huang, M. J., Huang, C. S., & Tang, T. K. (1997). Two novel glucose 6-phosphate dehydrogenase deficiency mutations and association of such mutations with F8C/G6PD haplotype in Chinese. *Journal of the Formosan Medical Association*, 96(12), 948-954.

- Chen, V. B., Arendall, W. B., 3rd, Headd, J. J., Keedy, D. A., Immormino, R. M., Kapral, G. J., Murray, L. W., Richardson, J. S., & Richardson, D. C. (2010). MolProbity: all-atom structure validation for macromolecular crystallography. *Acta Crystallographica. Section D, Biological Crystallography*, 66(Pt 1), 12–21.
- Chen, X., Yue, L., Li, C., & Li, C. (2010). A novel G473A mutation in the Glucose-6-phosphate dehydrogenase gene. *Pediatric Blood and Cancer*, 55(2), 383–385.
- Chinevere, T. D., Murray, C. K., Grant, E., Jr., Johnson, G. A., Duelm, F., & Hospenhal, D. R. (2006). Prevalence of Glucose-6-phosphate dehydrogenase deficiency in U.S. Army personnel. *Military Medicine*, 171(9), 905–907.
- Chiste, R. C., Freitas, M., Mercadante, A. Z., & Fernandes, E. (2014). Carotenoids inhibit lipid peroxidation and hemoglobin oxidation, but not the depletion of glutathione induced by ROS in human erythrocytes. *Life Sciences*, 99(1–2), 52–60.
- Chiu, D. T., Zuo, L., Chao, L., Chen, E., Louie, E., Lubin, B., Liu, T. Z., & Du, C. S. (1993). Molecular characterization of glucose-6-phosphate dehydrogenase (G6PD) deficiency in patients of Chinese descent and identification of new base substitutions in the human G6PD gene. *Blood*, 81(8), 2150–2154.
- Chiu, D. T., Zuo, L., Chen, E., Chao, L., Louie, E., Lubin, B., Liu, T. Z., & Du, C. S. (1991). Two commonly occurring nucleotide base substitutions in Chinese G6PD variants. *Biochemical and Biophysical Research Communications*, 180(2), 988–993.
- Chotivanich, K., Udomsangpetch, R., Pattanapanyasat, K., Chierakul, W., Simpson, J., Looareesuwan, S., & White, N. (2002). Hemoglobin E: A balanced polymorphism protective against high parasitemias and thus severe *P. falciparum* malaria. *Blood*, 100(4), 1172–1176.

- Choubisa, S. L. (2009). Sick cell haemoglobin, thalassaemia and G-6-PD enzyme deficiency genes in Garasiya tribe inhabited malaria endemic areas of Sirohi District, Rajasthan (India). *The Journal of Communicable Diseases*, 41(1), 13–18.
- Colovos, C., & Yeates, T. O. (1993). Verification of protein structures: patterns of nonbonded atomic interactions. *Protein science*, 2(9), 1511–1519.
- Corash, L., Spielberg, S., Bartsocas, C., Boxer, L., Steinherz, R., Sheetz, M., Egan, M., Schlesselman, J., & Schulman, J. D. (1980). Reduced chronic hemolysis during high-dose vitamin E administration in Mediterranean-type Glucose-6-phosphate dehydrogenase deficiency. *New England Journal of Medicine*, 303(8), 416–420.
- Cunningham, A. D., Colavin, A., Huang, K. C., & Mochly-Rosen, D. (2017). Coupling between protein stability and catalytic activity determines pathogenicity of G6PD variants. *Cell Reports*, 18(11), 2592–2599.
- D'Alessandro, A., Dzieciatkowska, M., Nemkov, T., & Hansen, K. C. (2017). Red blood cell proteomics update: Is there more to discover? *Blood Transfusion*, 15(2), 182–187.
- Daina, A., Michielin, O., & Zoete, V. (2017). SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Scientific reports*, 7, 42717.
- Deka, R., Reddy, A. P., Mukherjee, B. N., Das, B. M., Banerjee, S., Roy, M., Dey, B., Malhotra, K. C., & Walter, H. (1988). Haemoglobin E distribution in ten endogamous groups of Assam, India. *Human Heredity*, 38(5), 261–266.
- Deng, Z., Yang, F., Bai, Y., He, L., Li, Q., Wu, Y., Luo, L., Li, H., Ma, L., Yang, Z., He, Y., & Cui, L. (2017). Co-inheritance of Glucose-6-phosphate dehydrogenase deficiency mutations and hemoglobin E in a Kachin population in a malaria-endemic region of Southeast Asia. *Plos One*, 12(5), e0177917.

- Devendra, R., Gupta, V., Shanmugam, R., Singh, M. P. S. S., Patel, P., Valecha, N., Mishra, N., Ahmed, N., Hoti, S. L., Hegde, H. V., Warang, P., Chiddarwar, A., Kedar, P., Mayekar, P., & Mukherjee, M. B. (2020). Prevalence and spectrum of mutations causing G6PD deficiency in Indian populations. *Infection, Genetics and Evolution*, *86*, 104597.
- Devendra, R., Warang, P., Gupta, V., Chiddarwar, A., Kedar, P., Agarwal, M. B., & Mukherjee, M. B. (2019). A novel G6PD p.Gly 321 Val mutation causing severe hemolysis in an Indian infant. *Indian Journal of Hematology and Blood Transfusion*, *35*(2), 399–401.
- Doss, C. G., Alasmar, D. R., Bux, R. I., Sneha, P., Bakhsh, F. D., Al-Azwani, I., Bekay, R. E., & Zayed, H. (2016). Genetic epidemiology of Glucose-6-phosphate dehydrogenase deficiency in the Arab world. *Scientific Reports*, *6*, 37284.
- Durrant, J. D., & McCammon, J. A. (2011). Molecular dynamics simulations and drug discovery. *BMC Biology*, *9*, 71.
- Eisenberg, D., Lüthy, R., & Bowie, J. U. (1997). VERIFY3D: assessment of protein models with three-dimensional profiles. *Methods in Enzymology*, *277*, 396–404.
- El-Deen, Z.M., Hussin, N.N., Hamid, T.A., Migeed, O.R., & Samy, R.M. (2013). G6PD deficiency and G6PD (Mediterranean and Silent) polymorphisms in Egyptian infants with Neonatal Hyperbilirubinemia. *Labmedicine*, *44*, 228-234.
- Ellella, S. A., Tawfik, M., Barseem, N., & Moustafa, W. (2017). Prevalence of glucose-6-phosphate dehydrogenase deficiency in neonates in Egypt. *Annals of Saudi Medicine*, *37*(5), 362–365.
- El-Gezeiry, D., El-Gendy, W., El-Kaffash, D., Hassab, H., & Moez, P. (2005). Comparative Study For The Detection of 563 C---- T G6PD Mutation Using Restriction Enzyme Assay and Amplification and Refractory Mutation System (ARMS). *Alexandria Journal of Pediatrics*, *19*(1), 51-55.

- Elsea, S. H., Razjouyan, J., Lee, K. M., Lynch, J. A., Martini, S., & Pandit, L. M. (2023). Association of Glucose-6-Phosphate Dehydrogenase Deficiency With Outcomes in US Veterans With COVID-19. *JAMA Network Open*, 6(3), e235626.
- Fazary, A. E., Awwad, N. S., Ibrahim, H. A., Shati, A. A., Alfaifi, M. Y., & Ju, Y. H. (2020). Protonation equilibria of N-acetylcysteine. *ACS Omega*, 5(31), 19598–19605.
- Ferreira, L. G., Dos Santos, R. N., Oliva, G., & Andricopulo, A. D. (2015). Molecular docking and structure-based drug design strategies. *Molecules*, 20(7), 13384–13421.
- Fikrika, H., Ambarsari, L., & Sumaryada, T. (2016). Molecular Docking Studies of Catechin and Its Derivatives as Anti-bacterial Inhibitor for Glucosamine-6-Phosphate Synthase. *IOP Conference Series: Earth & Environmental Science*, 31(1), 012009.
- Filho, W. L., Scheday, S., Boenecke, J., Gogoi, A., Maharaj, A., & Korovou, S. (2019). Climate change, health and mosquito-borne diseases: Trends and implications to the pacific region. *International Journal of Environmental Research and Public Health*, 16(24), 5114.
- Fiorelli, G., Martinez di Montemuros, F., & Cappellini, M. D. (2000). Chronic non-spherocytic haemolytic disorders associated with glucose-6-phosphate dehydrogenase variants. *Bailliere's Best Practice & Research. Clinical Haematology*, 13(1), 39–55.
- Fischer, M., Coleman, R. G., Fraser, J. S., & Shoichet, B. K. (2014). Incorporation of protein flexibility and conformational energy penalties in docking screens to improve ligand discovery. *Nature chemistry*, 6(7), 575–583.
- Forget, B. G., & Bunn, H. F. (2013). Classification of the disorders of hemoglobin. *Cold Spring Harbor Perspectives in Medicine*, 3(2), a011684.

- Francis, R. O., D'Alessandro, A., Eisenberger, A., Soffing, M., Yeh, R., Coronel, E., Sheikh, A., Rapido, F., La Carpia, F., Reisz, J. A., Gehrke, S., Nemkov, T., Thomas, T., Schwartz, J., Divgi, C., Kessler, D., Shaz, B. H., Ginzburg, Y., Zimring, J. C., Spitalnik, S. L., & Hod, E. A. (2020). Donor Glucose-6-phosphate dehydrogenase deficiency decreases blood quality for transfusion. *Journal of Clinical Investigation*, *130*(5), 2270–2285.
- Frank, J. E. (2005). Diagnosis and management of G6PD deficiency. *American Family Physician*, *72*(7), 1277–1282.
- Fujii, T., Belletti, A., Carr, A., Furukawa, T. A., Luethi, N., Putzu, A., Sartini, C., Salanti, G., Tsujimoto, Y., Udy, A. A., Young, P. J., & Bellomo, R. (2019). Vitamin C therapy for patients with sepsis or septic shock: A protocol for a systematic review and a network meta-analysis. *BMJ Open*, *9*(11), e033458.
- Gandomani, M. G., Khatami, S. R., Nezhad, S. R., Daneshmand, S., & Mashayekhi, A. (2011). Molecular identification of G6PD Chatham (G1003A) in Khuzestan province of Iran. *Journal of genetics*, *90*(1), 143–145.
- Garcia, A. A., Koperniku, A., Ferreira, J. C. B., & Mochly-Rosen, D. (2021). Treatment strategies for Glucose - 6 - phosphate dehydrogenase deficiency: past and future perspectives. *Trends in Pharmacological Sciences*, *42*(10), 829–844.
- Gautam, N., Gaire, B., Manand har, T., Marasini, B. P., Parajuli, N., Lekhak, S. P., & Nepal, M. (2019). Glucose 6 phosphate dehydrogenase deficiency and hemoglobinopathy in South Western Region Nepal: A boon or burden. *BMC Research Notes*, *12*(1), 734.
- Georgakouli, K., Deli, C. K., Zalavras, A., Fatouros, I. G., Kouretas, D., Koutedakis, Y., & Jamurtas, A. Z. (2013). α -lipoic acid supplementation up-regulates antioxidant capacity in adults with G6PD deficiency. *Food and Chemical Toxicology*, *61*, 69–73.

- Georgakouli, K., Fatouros, I. G., Fragkos, A., Tzatzakis, T., Deli, C. K., Papanikolaou, K., Koutedakis, Y., & Jamurtas, A. Z. (2018). Exercise and redox status responses following alpha-lipoic acid supplementation in G6PD deficient individuals. *Antioxidants*, 7(11), 162.
- Ghosh, K., Colah, R. B., & Mukherjee, M. B. (2015). Haemoglobinopathies in tribal populations of India. *Indian Journal of Medical Research*, 141(5), 505-508.
- Ghosh, S., & Vishveshwara, S. (2014). Ranking the quality of protein structure models using sidechain based network properties. *FI000 Research*, 3, 17.
- Goheen, M. M., Campino, S., & Cerami, C. (2017). The role of the red blood cell in host defence against falciparum malaria: An expanding repertoire of evolutionary alterations. *British Journal of Haematology*, 179(4), 543–556.
- Gomez-Manzo, S., Marcial-Quino, J., Vanoye-Carlo, A., Serrano-Posada, H., Ortega-Cuellar, D., Gonzalez-Valdez, A., Castillo-Rodriguez, R. A., Hernandez-Ochoa, B., Sierra-Palacios, E., Rodriguez-Bustamante, E., & Arreguin-Espinosa, R. (2016). Glucose-6-phosphate dehydrogenase: Update and analysis of new mutations around the world. *International Journal of Molecular Sciences*, 17(12), 2069.
- Gong, L., Maiteki-Sebuguzi, C., Rosenthal, P. J., Hubbard, A. E., Drakeley, C. J., Dorsey, G., & Greenhouse, B. (2012). Evidence for both innate and acquired mechanisms of protection from *Plasmodium falciparum* in children with sickle cell trait. *Blood*, 119(16), 3808–3814.
- Goshal, D. T. (1979). Study of blood groups, erythrocytic enzymes, serum proteins and hemoglobin variants in the endogamous castes from Cutch (MSc Thesis). University of Bombay.
- Guindo, A., Fairhurst, R. M., Doumbo, O. K., Wellem, T. E., & Diallo, D. A. (2007). X-linked G6PD deficiency protects hemizygous males but not heterozygous females against severe malaria. *PLOS Medicine*, 4(3), e66.

- Gurkan E. (2006). Vaso-occlusive manifestations in a patient with sickle cell– hemoglobin E (Hb SE) disease. *American Journal of Hematology*, 81, 149–156.
- Hafez, M., Amar, E. S., Zedan, M., Hammad, H., Sorour, A. H., el-Desouky, E. S., & Gamil, N. (1986). Improved erythrocyte survival with combined vitamin E and selenium therapy in children with Glucose-6-phosphate dehydrogenase deficiency and mild chronic hemolysis. *Journal of Pediatrics*, 108(4), 558–561.
- Haloui, S., Laouini, N., Sahli, C. A., Daboubi, R., Becher, M., Jouini, L., Kazdaghli, K., Tinsa, F., Cherif, S., Khemiri, M., Fredj, S. H., Othmani, R., Ouali, F., Siala, H., Toumi, N.elH., Barsaoui, S., Bibi, A., & Messaoud, T. (2016). Molecular identification of Gd A- and Gd B- G6PD deficient variants by ARMS-PCR in a Tunisian population. *Annals of Clinical Bioogyl (Paris)*, 74(2), 219–226.
- Hamada, M., Shirakawa, T., Poh-San, L., Nishiyama, K., Uga, S., & Matsuo, M. (2010). Two new variants of G6PD deficiencies in Singapore. *Nepal Medical College Journal*, 12(3), 137–141.
- Harcke, S. J., Rizzolo, D., & Harcke, H. T.. (2019). G6PD deficiency: An update. *Journal of the American Academy of Physician Assistants*, 32(11), 21–26.
- Harewood, J., & Azevedo, A. M. (2023). Alpha thalassemia. In *StatPearls* [Internet]. *StatPearls Publishing*.
- Harteveld, C. L., Achour, A., Arkesteijn, S. J. G., terHuurne, J., Verschuren, M., Bhagwandien-Bisoen, S., Schaap, R., Vijfhuizen, L., el Idrissi, H., & Koopmann, T. T. (2022). The hemoglobinopathies, molecular disease mechanisms and diagnostics. *International Journal of Laboratory Hematology*, 44(Suppl. 1), 28–36.
- Hill, A. V., Allsopp, C. E., Kwiatkowski, D., Anstey, N. M., Twumasi, P., Rowe, P. A., Bennett, S., Brewster, D., McMichael, A. J., & Greenwood, B. M. (1991). Commonwest African HLA antigens are associated with protection from severe malaria. *Nature*, 352(6336), 595–600.

- Hirono, A., Kawate, K., Honda, A., Fujii, H., & Miwa, S. (2002). A single mutation 202G>A in the human Glucose-6-phosphate dehydrogenase gene (G6PD) can cause acute hemolysis by itself. *Blood*, *99*(4), 1498.
- Hockham, C., Ekwattanakit, S., Bhatt, S., Penman, B. S., Gupta, S., Viprakasit, V., & Piel, F. B. (2019). Estimating the burden of α -thalassaemia in Thailand using a comprehensive prevalence database for Southeast Asia. *eLife*, *8*, e40580.
- Horikoshi, N., Hwang, S., Gati, C., Matsui, T., Castillo-Orellana, C., Raub, A. G., Garcia, A. A., Jabbarpour, F., Batyuk, A., Broweleit, J., Xiang, X., Chiang, A., Broweleit, R., Vohringer-Martinez, E., Mochly-Rosen, D., & Wakatsuki, S. (2021). Long-range structural defects by pathogenic mutations in most severe Glucose-6-phosphate dehydrogenase deficiency. *Proceedings of the National Academy of Sciences of the United States of America*, *118*(4), e2022790118.
- Howes, R. E., Piel, F. B., Patil, A. P., Nyangiri, O. A., Gething, P. W., Dewi, M., Hogg, M. M., Battle, K. E., Padilla, C. D., Baird, J. K., & Hay, S. I. (2012). G6PD deficiency prevalence and estimates of affected populations in malaria endemic countries: a geostatistical model-based map. *PLoS Medicine*, *9*(11), e1001339.
- Hsia, Y. E., Miyakawa, F., Baltazar, J., Ching, N. S., Yuen, J., Westwood, B., & Beutler, E. (1993). Frequency of glucose-6-phosphate dehydrogenase (G6PD) mutations in Chinese, Filipinos, and Laotians from Hawaii. *Human genetics*, *92*(5), 470–476.
- Hubbard, S. J., & Thornton, J. M. (1993). ‘NACCESS’, computer program. *Department of Biochemistry and Molecular Biology, University College, London*.
- Hue, N. T., Charlieu, J. P., Chau, T. T., Day, N., Farrar, J. J., Hien, T. T., & Dunstan, S. J. (2009). Glucose-6-phosphate dehydrogenase (G6PD) mutations and haemoglobinuria syndrome in the Vietnamese population. *Malaria Journal*, *8*, 152.
- Hung, N. M., Eto, H., Mita, T., Tsukahara, T., Hombhanje, F. W., Hwaihwanje, I., Takahashi, N., & Kobayakawa, T. (2008). Glucose-6-phosphate dehydrogenase

(G6PD) variants in East Sepik province of Papua New Guinea: G6PD Jammu, G6PD Vanua Lava, and a novel variant (G6PD Dagua). *Tropical Medicine and Health*, 36(4), 163-169.

Hutagalung, J., Soleha, M., Sitorus, N., & Hananta, L. (2019). The Genotyping of Glucose 6 Phosphate Dehydrogenase deficiency (G6PD-d) in Malaria Endemic South Central Timor, East Nusa Tenggara, Eastern Indonesia. IntechOpen. <https://doi.org/10.5772/intechopen.88954>

Hwang, S., Mruk, K., Rahighi, S., Raub, A. G., Chen, C. H., Dorn, L. E., Horikoshi, N., Wakatsuki, S., Chen, J. K., & Mochly-Rosen, D. (2018). Correcting Glucose-6-phosphate dehydrogenase deficiency with a small-molecule activator. *Nature Communications*, 9(1), 4045.

Ishwad, C. S., & Naik, S. N. (1984). A new Glucose-6-phosphate dehydrogenase variant (G6PD Kalyan) found in a Koli family. *Human Genetics*, 66(2-3), 171-175.

Iwai, K., Hirono, A., Matsuoka, H., Kawamoto, F., Horie, T., Lin, K., Tantular, I. S., Dachlan, Y. P., Notopuro, H., Hidayah, N. I., Salim, A. M., Fujii, H., Miwa, S., & Ishii, A. (2001). Distribution of Glucose-6-phosphate dehydrogenase mutations in Southeast Asia. *Human Genetics*, 108(6), 445-449.

Jiang, W., Yu, G., Liu, P., Geng, Q., Chen, L., Lin, Q., Ren, X., Ye, W., He, Y., Guo, Y., Duan, S., Wen, J., Li, H., Qi, Y., Jiang, C., Zheng, Y., Liu, C., Si, E., Zhang, Q., Tian, Q., & Du, C. (2006). Structure and function of Glucose-6-phosphate dehydrogenase deficient variants in Chinese population. *Human Genetics*, 119(5), 463-478.

Johnson, G. J., Vatassery, G. T., Finkel, B., & Allen, D. W. (1983). High-dose vitamin E does not decrease the rate of chronic hemolysis in glucose-6-phosphate dehydrogenase deficiency. *New England Journal of Medicine*, 308(17), 1014-1017.

- Johnson, M. K., Clark, T. D., Njama-Meya, D., Rosenthal, P. J., & Parikh, S. (2009). Impact of the method of G6PD deficiency assessment on genetic association studies of malaria susceptibility. *Plos One*, *4*(9), e7246.
- Kaeda, J. S., Chhotray, G. P., Ranjit, M. R., Bautista, J. M., Reddy, P. H., Stevens, D., Naidu, J. M., Britt, R. P., Vulliamy, T. J., Luzzatto, L., & Mason, P. J. (1995). A new Glucose-6-phosphate dehydrogenase variant, G6PD Orissa (44 Ala–Gly), is the major polymorphic variant in tribal populations in India. *American Journal of Human Genetics*, *57*(6), 1335–1341.
- Kaeda, J. S., Chhotray, G. P., Reddy, P. H., Stevens, D., Britt, R. P., Vulliamy, T., Luzzatto, L., & Mason, P. J. (1995). Molecular genetics of G6PD deficiency in India. *British Society for Haematology* (Abstract).
- Kamble, M., & Chatruvedi, P. (2000). Epidemiology of sickle cell disease in a rural hospital in central India. *Indian Pediatrics*, *37*(4), 391–396.
- Kaplan, M., & Hammerman, C. (2004). Glucose-6-phosphate dehydrogenase deficiency: a hidden risk for kernicterus. *Seminars in perinatology*, *28*(5), 356–364.
- Karna, B., Jha, S. K., & Al Zaabi, E. (2023). Hemoglobin C Disease. In *StatPearls* [Internet]. *StatPearls Publishing*.
- Kawamoto, F., Matsuoka, H., Kanbe, T., Tantular, I. S., Pusarawati, S., Kerong, H. I., Damianus, W., Mere, D., & Dachlan, Y. P. (2006). Further investigations of Glucose-6-phosphate dehydrogenase variants in Flores Island, Eastern Indonesia. *Journal of Human Genetics*, *51*(11), 952–957.
- Khan, K. A., Qureshi, S. U., Khalid, L., & Wahid, K. (2019). A typical presentation of dengue fever in a G6PD deficient patient: A case report. *The Journal of the Pakistan Medical Association*, *69*(10), 1553–1556.
- Khim, N., Benedet, C., Kim, S., Kheng, S., Siv, S., Leang, R., Lek, S., Muth, S., Chea, N., Chuor, C. M., Duong, S., Kerleguer, A., Tor, P., Chim, P., Canier, L., Witkowski,

- B., Taylor, W. R., & Ménard, D. (2013). G6PD deficiency in *Plasmodium falciparum* and *Plasmodium vivax* malaria-infected Cambodian patients. *Malaria Journal*, *12*, 171.
- Kohne, E. (2011). Hemoglobinopathies: Clinical manifestations, diagnosis, and treatment. *Deutsches Arzteblatt International*, *108*(31–32), 532–540.
- Korim, K. M. M., & Arbid, M. S. (2018). Evaluating of β -carotene role in ameliorating of favism-induced disturbances in blood and testis. *Journal of Complementary and Integrative Medicine*, *15*(3), doi:10.1515/jcim-2017-0164
- Kotaka, M., Gover, S., Vand eputte-Rutten, L. V., Au, S. W. N., Lam, V. M. S., & Adams, M. J. (2005). Structural studies of Glucose-6-phosphate and NADP⁺ binding to human Glucose-6-phosphate dehydrogenase. *Acta Crystallographica. Section D, Biological Crystallography*, *61*(5), 495–504.
- Kotepui, M., Uthaisar, K., PhunPhuech, B., & Phiwklam, N. (2016). Prevalence and hematological indicators of G6PD deficiency in malaria-infected patients. *Infectious Diseases of Poverty*, *5*, 36.
- Kumar, P., Yadav, U., & Rai, V. (2016). Prevalence of Glucose-6-phosphate dehydrogenase deficiency in India: An updated meta-analysis. *Egyptian Journal of Medical Human Genetics*, *17*(3), 295–302.
- Kurdi-Haidar, B., Mason, P. J., Berrebi, A., Ankra-Badu, G., al-Ali, A., Oppenheim, A., & Luzzatto, L. (1990). Origin and spread of the Glucose-6-phosphate dehydrogenase variant (G6PD-Mediterranean) in the Middle East. *American Journal of Human Genetics*, *47*(6), 1013–1019.
- Kyriakou, A., & Skordis, N. (2015). Thalassemia, endocrine sequelae. *Reference Module in Biomedical Sciences*, <https://doi.org/10.1016/B978-0-12-801238-3.04488-3>
- Laosombat, V., Sattayasevana, B., Janejindamai, W., Viprakasit, V., Shirakawa, T., Nishiyama, K., & Matsuo, M. (2005). Molecular heterogeneity of Glucose-6-

- phosphate dehydrogenase (G6PD) variants in the south of Thailand and identification of a novel variant (G6PD Songklanagarind). *Blood Cells, Molecules and Diseases*, 34(2), 191–196.
- Laskowski, R. A., & Swindells, M. B. (2011). LigPlot+: multiple ligand-protein interaction diagrams for drug discovery. *Journal of Chemical Information and Modeling*, 51(10), 2778-2786.
- Laskowski, R. A., MacArthur, M. W., Moss, D. S., & Thornton, J. M. (1993). PROCHECK - a program to check the stereochemical quality of protein structures. *Journal of Applied Crystallography*, 26, 283-291.
- Lee, H. Y., Ithnin, A., Azma, R. Z., Othman, A., Salvador, A., & Cheah, F. C. (2022). Glucose-6-phosphate dehydrogenase deficiency and neonatal hyperbilirubinemia: Insights on pathophysiology, diagnosis, and gene variants in disease heterogeneity. *Frontiers in Pediatrics*, 10, 875877.
- Lee, J., Kim, T. I., Kang, J. M., Jun, H., Lê, H. G., Thái, T. L., Sohn, W. M., Myint, M. K., Lin, K., Kim, T. S., & Na, B. K. (2018). Prevalence of glucose-6-phosphate dehydrogenase (G6PD) deficiency among malaria patients in Upper Myanmar. *BMC Infectious Diseases*, 18(1), 131.
- Lee, S. T., Yoo, E. H., Kim, J. Y., Kim, J. W., & Ki, C. S. (2010). Multiplex ligation-dependent probe amplification screening of isolated increased HbF levels revealed three cases of novel rearrangements/deletions in the β -globin gene cluster. *British Journal of Haematology*, 148(1), 154–160.
- Li, Q., Yang, F., Liu, R., Luo, L., Yang, Y., Zhang, L., Liu, H., Zhang, W., Fan, Z., Yang, Z., Cui, L., & He, Y. (2015). Prevalence and molecular characterization of Glucose-6-phosphate dehydrogenase deficiency at the China-Myanmar Border. *Plos One*, 10(7), e0134593.

- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney, P. J. (2001). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, 46(1-3), 3–26.
- Lodha, A., Kamaluddeen, M. S., Kelly, E., & Amin, H. (2008). Clostridium difficile infection precipitating hemolysis in glucose-6-phosphate dehydrogenase-deficient preterm twins causing severe neonatal jaundice. *Journal of Perinatology*, 28(1), 77–78.
- Luzzatto, L., Ally, M., & Notaro, R. (2020). Glucose-6-phosphate dehydrogenase deficiency. *Blood*, 136(11), 1225-1240.
- Luzzatto, L., & Poggi, V. (2009). Glucose-6-Phosphate Dehydrogenase Deficiency. In Nathan and Oskis Hematology of Infancy and Childhood. (7th ed), Philadelphia, PA: Saunders.
- Luzzatto, L., & Seneca, E. (2014). G6PD deficiency: a classic example of pharmacogenetics with on-going clinical implications. *British Journal of Haematology*, 164(4):469-80.
- Luzzatto, L., Mehta, A., & Vulliamy, T. (2001). Glucose 6-phosphate dehydrogenase deficiency. In Scriver, C. R., Beaudet, A. L. Slyand, W. S. D. & Valle, W. S. (Eds.), *The metabolic and molecular bases of inherited disease* (pp. 4517–1553). McGraw-Hill.
- Luzzatto, L., Nannelli, C., & Notaro, R. (2016). Glucose-6-Phosphate Dehydrogenase Deficiency. *Hematology/oncology clinics of North America*, 30(2), 373–393.
- Madan, N., Sharma, S., Sood, S. K., Colah, R., & Bhatia, H. M. (2010). Frequency of β -thalassemia trait and other hemoglobinopathies in northern and western India. *Indian Journal of Human Genetics*, 16(1), 16-25.

- Malaria Genomic Epidemiology Network, & Malaria Genomic Epidemiology Network (2014). Reappraisal of known malaria resistance loci in a large multicenter study. *Nature Genetics*, *46*(11), 1197–1204.
- Manco, L., Gonçalves, P., Antunes, P., Maduro, F., Abade, A., & Ribeiro, M. L. (2007). Mutations and haplotype diversity in 70 Portuguese G6PD-deficient individuals: An overview on the origin and evolution of mutated alleles. *Haematologica*, *92*(12), 1713–1714.
- Mason, P. J., Bautista, J. M., & Gilsanz, F. (2007). G6PD deficiency: The genotype-phenotype association. *Blood Reviews*, *21*(5), 267–283.
- Matsuoka, H., Thuan, D. T., van Thien, H., Kanbe, T., Jalloh, A., Hirai, M., Arai, M., Dung, N. T., & Kawamoto, F. (2007). Seven different glucose-6-phosphate dehydrogenase variants including a new variant distributed in Lam Dong Province in southern Vietnam. *Acta Medica Okayama*, *61*(4), 213–219.
- Matsuoka, H., Wang, J., Hirai, M., Arai, M., Yoshida, S., Kobayashi, T., Jalloh, A., Lin, K., & Kawamoto, F. (2004). Glucose-6-phosphate dehydrogenase (G6PD) mutations in Myanmar: G6PD Mahidol (487G>A) is the most common variant in the Myanmar population. *Journal of Human Genetics*, *49*(10), 544–547.
- Matsuoka, H., Nguon, C., Kanbe, T., Jalloh, A., Sato, H., Yoshida, S., Hirai, M., Arai, M., Socheat, D., & Kawamoto, F. (2005). Glucose-6-phosphate dehydrogenase (G6PD) mutations in Cambodia: G6PD Viangchan (871G>A) is the most common variant in the Cambodian population. *Journal of Human Genetics*, *50*(9), 468–472.
- Mehta, J., Rayalam, S., & Wang, X. (2018). Cytoprotective Effects of Natural Compounds against Oxidative Stress. *Antioxidants (Basel, Switzerland)*, *7*(10), 147.
- Menkin-Smith, L., & Winders, W. T. (2023). *Plasmodium vivax* Malaria. In *StatPearls* [Internet]. *StatPearls Publishing*.

- Menon, V., & Ghaffari, S. (2021). Erythroid enucleation: A gateway into a “bloody” world. *Experimental Hematology*, *95*, 13–22.
- Mesbah-Namin, S. A., Sanati, M. H., Mowjoodi, A., Mason, P. J., Vulliamy, T. J., & Noori-Dalooi, M. R. (2002). Three major Glucose-6-phosphate dehydrogenase deficient polymorphic variants identified in Mazandaran state of Iran. *British Journal of Haematology*, *117*(3), 763–764.
- Metzger, W. G., Mordmüller, B. G., & Kremsner, P. G. (1995). Malaria pigment in leucocytes. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, *89*(6), 637–638.
- Minucci, A., Moradkhani, K., Hwang, M. J., Zuppi, C., Giardina, B., & Capoluongo, E. (2012). Glucose-6-phosphate dehydrogenase (G6PD) mutations database: Review of the “old” and update of the new mutations. *Blood Cells, Molecules and Diseases*, *48*(3), 154–165.
- Minucci, A., De Luca, D., Torti, E., Concolino, P., Maurizi, P., Giardina, B., Zuppi, C., & Capoluongo, E. (2011). Acute hemolytic crisis to concomitant presence of infection and possible altered acetaminophen catabolism in a Filipino child carrying the G6PD Vanua Lava mutation. *Annals of Clinical Biochemistry*, *48*(3), 282–285.
- Mishra, A., Rana, P. S., Mittal, A., & Jayaram, B. (2014). D2N: Distance to the native. *Biochimica et Biophysica Acta*, *1844*(10), 1798–1807.
- Mockenhaupt, F. P., Ehrhardt, S., Gellert, S., Otchwemah, R. N., Dietz, E., Anemana, S. D., & Bienzle, U. (2004). Alpha(+)-thalassemia protects African children from severe malaria. *Blood*, *104*(7), 2003–2006.
- Mockenhaupt, F. P., Ehrhardt, S., Cramer, J. P., Otchwemah, R. N., Anemana, S. D., Goltz, K., Mylius, F., Dietz, E., Eggelte, T. A., & Bienzle, U. (2004). Hemoglobin C and resistance to severe malaria in Ghanaian children. *Journal of Infectious Diseases*, *190*(5), 1006–1009.

- Modell, B., & Darlison, M. (2008). Global epidemiology of haemoglobin disorders and derived service indicators. *Bulletin of the World Health Organization*, 86(6), 480–487.
- Modiano, D., Luoni, G., Sirima, B. S., Simporé, J., Verra, F., Konaté, A., Rastrelli, E., Olivieri, A., Calissano, C., Paganotti, G. M., D'Urbano, L., Sanou, I., Sawadogo, A., Modiano, G., & Coluzzi, M. (2001). Haemoglobin C protects against clinical *Plasmodium falciparum* malaria. *Nature*, 414(6861), 305–308.
- Mohanty, D., Colah, R. B., Gorakshakar, A. C., Patel, R. Z., Master, D. C., Mahanta, J., Sharma, S. K., Chaudhari, U., Ghosh, M., Das, S., Britt, R. P., Singh, S., Ross, C., Jagannathan, L., Kaul, R., Shukla, D. K., & Muthuswamy, V. (2013). Prevalence of β -thalassemia and other haemoglobinopathies in six cities in India: a multicentre study. *Journal of Community Genetics*, 4(1), 33–42.
- Moiz, B., Nasir, A., Khan, S. A., Kherani, S. A., & Qadir, M. (2012). Neonatal hyperbilirubinemia in infants with G6PD c.563C > T Variant. *BMC Pediatrics*, 12, 126.
- Moorthie, S. (2016). Link between G6PD deficiency and malaria. Available at <https://www.phgfoundation.org/news/link-between-g6pd-deficiency-and-malaria>. Accessed on November 18, 2023.
- Mukherjee, M. B., Colah, R. B., Martin, S., & Ghosh, K. (2015). Glucose-6-phosphate dehydrogenase (G6PD) deficiency among tribal populations of India - Country scenario. *Indian Journal of Medical Research*, 141(5), 516–520.
- Mukherjee, M. B., Lu, C. Y., Ducrocq, R., Gangakhedkar, R. R., Colah, R. B., Kadam, M. D., Mohanty, D., Nagel, R. L., & Krishnamoorthy, R. (1997). Effect of alpha thalassaemia on sickle cell anaemia linked to the Arab-India haplotype in India. *American Journal of Hematology*, 55(2), 104–109.

- Murthy, P. S. R. (2011). Health Care System in Tribal Areas – An Insight (With Reference to Andhra Pradesh State, India. Available at SSRN: <https://ssrn.com/abstract=1747341>. Accessed on November 18, 2023.
- National Health Mission. (2016). Prevention and control of haemoglobinopathies in India-thalassaemias, sickle cell disease and other variant haemoglobins. *Ministry of Health and Family Welfare, Government of India*.
- Newman, J. G., Newman, T. B., Bowie, L. J., & Mendelsohn, J. (1979). An examination of the role of vitamin E in Glucose-6-phosphate dehydrogenase. *Clinical Biochemistry*, *12*(5), 149–151.
- Ngo, T. T., Tran, T. H., Ta, T. D., Le, T. P., Nguyen, P. D., Tran, M. A., Bui, T. H., Ta, T. V., & Tran, V. K. (2022). Molecular Characterization and Genotype-Phenotype Correlation of G6PD Mutations in Five Ethnicities of Northern Vietnam. *Anemia*, *2022*, 2653089.
- Nguyen, P. H., Day, N., Pram, T. D., Ferguson, D. J., & White, N. J. (1995). Intraleucocytic malaria pigment and prognosis in severe malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, *89*(2), 200–204.
- Ninokata, A., Kimura, R., Samakkarn, U., Settheetham-Ishida, W., & Ishida, T. (2006). Coexistence of five G6PD variants indicates ethnic complexity of Phuket islanders, Southern Thailand. *Journal of Human Genetics*, *51*(5), 424–428.
- Nkhoma, E. T., Poole, C., Vannappagari, V., Hall, S. A., & Beutler, E. (2009). The global prevalence of Glucose-6-phosphate dehydrogenase deficiency: A systematic review and meta-analysis. *Blood Cells, Molecules and Diseases*, *42*(3), 267–278.
- Ntie-Kang F. (2013). An in silico evaluation of the ADMET profile of the StreptomeDB database. *SpringerPlus*, *2*, 353.

- Nuinoon, M., Krithong, R., Pramtong, S., Sasuk, P., Ngeaiad, C., Chaimusik, S., Kanboonma, J., & Sarakul, O. (2022). Prevalence of G6PD deficiency and G6PD variants amongst the southern Thai population. *PeerJ*, *10*, e14208.
- O'Boyle, N. M., Banck, M., James, C. A., Morley, C., Vandermeersch, T., & Hutchison, G. R. (2011). Open Babel: An open chemical toolbox. *Journal of Cheminformatics*, *3*, 33.
- Olusanya, B. O., Emokpae, A. A., Zamora, T. G., & Slusher, T. M. (2014). Addressing the burden of neonatal hyperbilirubinaemia in countries with significant glucose-6-phosphate dehydrogenase deficiency. *Acta Paediatrica*, *103*(11), 1102–1109.
- Paek, D. S., Nadkarni, M., & Singla, M. (2009). Treatment of MRSA infections in an African-American male with G6PD deficiency. *The Journal of Emergency Medicine*, *37*(3), 273–278.
- Pearson, A. G., Pullar, J. M., Cook, J., Spencer, E. S., Vissers, M. C., Carr, A. C., & Hampton, M. B. (2021). Peroxiredoxin 2 oxidation reveals hydrogen peroxide generation within erythrocytes during high-dose vitamin C administration. *Redox Biology*, *43*, 101980.
- Pei, Y., Liu, H., Yang, Y., Yang, Y., Jiao, Y., Tay, F. R., & Chen, J. (2018). Biological Activities and Potential Oral Applications of N-Acetylcysteine: Progress and Prospects. *Oxidative Medicine and Cellular Longevity*, *2018*, 2835787.
- Persico, M. G., Viglietto, G., Martini, G., Toniolo, D., Paonessa, G., Moscatelli, C., Dono, R., Vulliamy, T., Luzzatto, L., & D'Urso, M. (1986). Isolation of human Glucose-6-phosphate dehydrogenase (G6PD) cDNA clones: primary structure of the protein and unusual 5' non-coding region. *Nucleic Acids Research*, *14*(6), 2511-2522.
- Peters, A. L., & Van Noorden, C. J. F. (2009). Glucose-6-phosphate dehydrogenase deficiency and malaria: Cytochemical detection of heterozygous G6PD deficiency in women. *Journal of Histochemistry and Cytochemistry*, *57*(11),

1003-1011.

- Phompradit, P., Kuesap, J., Chaijaroenkul, W., Rueangweerayut, R., Hongkaew, Y., Yamnuan, R., & Na-Bangchang, K. (2011). Prevalence and distribution of Glucose-6-phosphate dehydrogenase (G6PD) variants in Thai and Burmese populations in malaria endemic areas of Thailand. *Malaria Journal*, *10*, 368.
- Poon, M. C., Hall, K., Scott, C. W., & Prchal, J. T. (1988). G6PD Viangchan: A new glucose 6-phosphate dehydrogenase variant from Laos. *Human Genetics*, *78*(1), 98-99.
- Quereshy, F. A., Gold, E. S., & Powers, M. P. (2000). Hemolytic anemia in a glucose-6-phosphate dehydrogenase-deficient patient triggered by a maxillofacial infection. *Journal of Oral and Maxillofacial Surgery*, *58*(7), 805–807.
- Raftos, J. E., Whillier, S., & Kuchel, P. W. (2010). Glutathione synthesis and turnover in the human erythrocyte: Alignment of a model based on detailed enzyme kinetics with experimental data. *Journal of Biological Chemistry*, *285*(31), 23557–23567.
- Raftos, J. E., Whillier, S., Chapman, B. E., & Kuchel, P. W. (2007). Kinetics of uptake and deacetylation of N-acetylcysteine by human erythrocytes. *International Journal of Biochemistry and Cell Biology*, *39*(9), 1698–1706.
- Rai, V., & Kumar, P. (2012). Epidemiological study of Glucose-6-phosphate dehydrogenase deficiency in Scheduled Caste Population of India. *Journal of Anthropology*, *2012*, 984180.
- Rajkhowa, P., Nath, C., Dutta, A., Misurya, I., Sharma, N., Barman, B., Longkumer, C., Lynrah, K. G., Sarmah, D., & Ruram, A. (2020). Study of Glucose-6-phosphate dehydrogenase (G6PD) deficiency and genotype polymorphism of G6PD B and G6PD (A+/A-) in Patients Treated for *Plasmodium vivax* Malaria in a Tertiary Care Hospital in North East India. *Cureus*, *12*(11), e11463.

- Rao, S., Kar, R., Gupta, S. K., Chopra, A., & Saxena, R. (2010). Spectrum of hemoglobinopathies diagnosed by cation exchange-HPLC and modulating effects of nutritional deficiency anaemias from North India. *Indian Journal of Medical Research*, 132(5), 513–519.
- Rattazzi, M. C. (1968). Glucose-6-phosphate dehydrogenase from human erythrocytes: Molecular weight determination by gel filtration. *Biochemical and Biophysical Research Communications*, 31(1), 16–24.
- Raupp, P., Hassan, J. A., Varughese, M., & Kristiansson, B. (2001). Henna causes life threatening haemolysis in Glucose-6-phosphate dehydrogenase deficiency. *Archives of Disease in Childhood*, 85(5), 411–412.
- Reeves, D. J., Saum, L., & Birhiray, R. (2016). I.V. ascorbic acid for treatment of apparent rasburicase-induced methemoglobinemia in a patient with acute kidney injury and assumed Glucose-6-phosphate dehydrogenase deficiency. *American Journal of Health-system Pharmacy*, 73(9), e238-e242.
- Rehman, A., Shehadeh, M., Khirfan, D., & Jones, A. (2018). Severe acute haemolytic anaemia associated with severe methaemoglobinaemia in a G6PD-deficient man. *BMJ Case Reports*, 2018, bcr2017223369.
- Ren, X., He, Y., Du, C., Jiang, W., Chen, L., & Lin, Q. (2001). A novel mis-sense mutation (G1381A) in the G6PD gene identified in a Chinese man. *Chinese Medical Journal*, 114(4), 399–401.
- Roberts, D. J., & Williams, T. N. (2003). Haemoglobinopathies and resistance to malaria. *Redox Report: Communications in Free Radical Research*, 8(5), 304–310.
- Roy, A., Kucukural, A., & Zhang, Y. (2010). I-TASSER: a unified platform for automated protein structure and function prediction. *Nature Protocols*, 5(4), 725–738.
- Ruwende, C., Khoo, S. C., Snow, R. W., Yates, S. N., Kwiatkowski, D., Gupta, S., Warn, P., Allsopp, C. E., Gilbert, S. C., & Peschu, N. (1995). Natural selection of hemi-

- and heterozygotes for G6PD deficiency in Africa by resistance to severe malaria. *Nature*, 376(6537), 246–249.
- Saddala, M. S., Lennikov, A., & Huang, H. (2020). Discovery of small-molecule activators for Glucose-6-phosphate dehydrogenase (G6PD) using machine learning approaches. *International Journal of Molecular Sciences*, 21(4), 1523.
- Salvador, A., & Savageau, M. A. (2003). Quantitative evolutionary design of glucose 6-phosphate dehydrogenase expression in human erythrocytes. *Proceedings of the National Academy of Sciences of the United States of America*, 100(24), 14463–14468.
- Sanophonasa, A., Cheepsunthorn, C. L., Khaminsou, N., Savongsy, O., Nuchprayoon, I., & Leecharoenkiat, K. (2021). Molecular characterization of G6PD mutations reveals the high frequency of G6PD Aures in the Lao Theung population. *Malaria Journal*, 20(1), 30.
- Santucci, K., & Shah, B. (2000). Association of naphthalene with acute hemolytic anemia. *Academic Emergency Medicine*, 7(1), 42–47.
- Sarkar, B. C. (2021). Tea, Tribes and Doors A geographical perspective. *LAP LAMBERT Academic Publishing*.
- Sarkar, S., Biswas, N. K., Dey, B., Mukhopadhyay, D., & Majumder, P. P. (2010). A large, systematic molecular-genetic study of G6PD in Indian populations identifies a new non-synonymous variant and supports recent positive selection. *Infection, Genetics and Evolution*, 10(8), 1228–1236.
- Sarma, D. K., Mohapatra, P. K., Bhattacharyya, D. R., Chellappan, S., Karuppusamy, B., Barman, K., Senthil Kumar, N., Dash, A. P., Prakash, A., & Balabaskaran Nina, P. (2019, December 10). Malaria in North-East India: Importance and implications in the era of elimination. *Microorganisms*, 7(12), 673.
- Sarnaik, S. A. (2005). Thalassaemia and related haemoglobinopathies. *Indian Journal of*

Pediatrics, 72(4), 319–324.

Sayyed, Z. S., Mudera, V. C., Colah, R. B., & Gupte, S. C. (1994). G6PD Jamnagar: A new class III variant associated with drug induced haemolytic anemia. *Indian Journal of Hematology and Blood Transfusion*, 12, 210–215.

Sayyed, Z. S., Mukherjee, M. B., Mudera, V. C., Colah, R., & Gupte, S. (1992). Characterization of G6PD Rohini—A new class III variant. *Indian Journal of Medical Research*, 96, 96–100.

Schmidtke, P., & Barril, X. (2010). Understanding and predicting druggability. A high-throughput method for detection of drug binding sites. *Journal of Medicinal Chemistry*, 53(15), 5858–5867.

Schrodinger, L., & DeLano, W. (2020). *PyMOL*. Retrieved from <http://www.pymol.org/pymol>

Schwarzer, E., & Arese, P. (1996). Phagocytosis of malarial pigment hemozoin inhibits NADPH-oxidase activity in human monocyte-derived macrophages. *Biochimica et Biophysica Acta*, 1316(3), 169–175.

Shah, I., Jarullah, J., Jaouni, S. A., Jamal, M. S., & Jarullah, B. (2017). Higher prevalence of Glucose-6-phosphate dehydrogenase (G6PD) deficiency in tribal population against urban population: A signal to natural selection. *Biomedical Research*, 28(1), 385-388.

Singh, A., Kaushik, R., Mishra, A., Shanker, A., & Jayaram, B. (2016). ProTSAV: A protein tertiary structure analysis and validation server. *Biochimica et Biophysica Acta*, 1864(1), 11–19.

Singh, M. R., Choudhury, B., & Singh, T. S. (2010). Haemoglobin E Distribution in Four Endogamous Populations of Manipur (India). *Eurasian Journal of Anthropology*, 1(2), 109–117.

- Sinha, S., Black, M. L., Agarwal, S., Colah, R., Das, R., Ryan, K., Bellgard, M., & Bittles, A. H. (2009). Profiling β -thalassaemia mutations in India at state and regional levels: implications for genetic education, screening and counselling programmes. *The HUGO Journal*, 3(1-4), 51–62.
- Sobngwi, E., Gautier, J. F., Kevorkian, J. P., Villette, J. M., Riveline, J. P., Zhang, S., Vexiau, P., Leal, S. M., Vaisse, C., & Mauvais-Jarvis, F. (2005). High prevalence of glucose-6-phosphate dehydrogenase deficiency without gene mutation suggests a novel genetic mechanism predisposing to ketosis-prone diabetes. *Journal of clinical Endocrinology and Metabolism*, 90(8), 4446–4451.
- Sonbol, M. B., Yadav, H., Vaidya, R., Rana, V., & Witzig, T. E. (2013). Methemoglobinemia and hemolysis in a patient with G6PD deficiency treated with rasburicase. *American Journal of Hematology*, 88(2), 152–154.
- Spielberg, S. P., Boxer, L. A., Corash, L. M., & Schulman, J. D. (1979). Improved erythrocyte survival with high-dose vitamin E in chronic hemolyzing G6PD and glutathione synthetase deficiencies. *Annals of Internal Medicine*, 90(1), 53–54.
- Stanton, R. C. (2012). Glucose-6-phosphate dehydrogenase, NADPH, and cell survival. *IUBMB Life*, 64(5), 362–369.
- Steinberg M. H. (2020). Fetal hemoglobin in sickle cell anemia. *Blood*, 136(21), 2392–2400.
- Stincone, A., Prigione, A., Cramer, T., Wamelink, M. M. C., Campbell, K., and Cheung, E., Olin-Sandoval, V., Gruning, N. M., Kruger, A., Alam, M. T., Keller, M. A., Breitenbach, M., Brindle, K. M., Rabinowitz, J. D., & Ralser, M. (2015). The return of metabolism: biochemistry and physiology of the pentose phosphate pathway. *Biological Reviews*, 90(3): 927–963.

- Storey, J., Fleming, A. F., Cornille-Brøgger, R., Molineaux, L., Matsushima, T., & Kagan, I. (1979). Abnormal haemoglobins in the Sudansavanna of Nigeria. IV. Malaria, immunoglobulins and antimalarial antibodies in haemoglobin AC individuals. *Annals of Tropical Medicine and Parasitology*, 73(4), 311–315.
- Sukumar, S., Mukherjee, M. B., Colah, R. B., & Mohanty, D. (2003). Molecular characterization of G6PD Insuli—A novel 989 CGC→CAC (330 Arg→His) mutation in the Indian population. *Blood Cells, Molecules and Diseases*, 30(3), 246–247.
- Sukumar, S., Mukherjee, M. B., Colah, R. B., & Mohanty, D. (2004). Molecular basis of G6PD deficiency in India. *Blood Cells, Molecules and Diseases*, 33(2), 141–145.
- Sukumar, S., Mukherjee, M. B., Colah, R. B., & Mohanty, D. (2005). Two distinct Indian G6PD variants G6PD Jamnagar and G6PD Rohini caused by the same 949 G→A mutation. *Blood Cells, Molecules and Diseases*, 35(2), 193–195.
- Sulistyaningrum, N., Arlinda, D., Hutagalung, J., Sunarno, S., Oktoberia, I. S., Handayani, S., Ekowatiningsih, R., Yusnita, E. A., Prasetyorini, B., Rizki, A., Tjitra, E., Na-Bangchang, K., & Chaijaroenkul, W. (2020). Prevalence of Glucose 6-Phosphate Dehydrogenase Variants in Malaria-Endemic Areas of South Central Timor, Eastern Indonesia. *American Journal of Tropical Medicine and Hygiene*, 103(2), 760-766.
- Tang, T. K., Huang, C. S., Huang, M. J., Tam, K. B., Yeh, C. H., & Tang, C. J. (1992). Diverse point mutations result in glucose-6-phosphate dehydrogenase (G6PD) polymorphism in Taiwan. *Blood*, 79(8), 2135–2140.
- Tanphaichitr, V. S., Hirono, A., Pung-Amritt, P., Treesucon, A., & Wanachiwanawin, W. (2011). Chronic nonspherocytic haemolytic anemia due to Glucose-6-phosphate dehydrogenase deficiency: Report of two families with novel mutations causing G6PD Bangkok and G6PD Bangkok Noi. *Annals of Hematology*, 90(7), 769–775.

- Tantular, I. S., & Kawamoto, F. (2021). Distribution of G6PD deficiency genotypes among Southeast Asian populations. *Tropical Medicine and Health*, 49(1), 97.
- Taylor, S. M., Parobek, C. M., & Fairhurst, R. M. (2012). Impact of haemoglobinopathies on the clinical epidemiology of malaria: A systematic review and meta-analysis. *Lancet Infectious Diseases*, 12(6), 457–468.
- Taylor, S. M., Parobek, C. M., and Fairhurst, R. M. (2012). Haemoglobinopathies and the clinical epidemiology of malaria: A systematic review and meta-analysis. *Lancet Infectious Diseases*, 12(6), 457–468.
- Temel, Y., Bengu, A. Ş., Akkoyun, H. T., Akkoyun, M., & Ciftci, M. (2017). Effect of astaxanthin and aluminumchloride on erythrocyte G6PD and 6PGD enzyme activities in vivo and on erythrocyte G6PDin vitro in rats. *Journal of Biochemical and Molecular Toxicology*, 31(10), e21954 60.
- Tian, W., Chen, C., Lei, X., Zhao, J., & Liang, J. (2018). CASTp 3.0: computed atlas of surface topography of proteins. *Nucleic Acids Research*, 46(W1), W363–W367.
- Tripathy, V., & Reddy, B. M. (2007). Present status of understanding on the G6PD deficiency and natural selection. *Journal of Postgraduate Medicine*, 53(3), 193–202.
- Trott, O., & Olson, A. J. (2010). AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2), 455–461.
- Ursu, O., Rayan, A., Goldblum, A., & Oprea, T.I. (2011), Understanding drug-likeness. *WIREs Computational Molecular Science*, 1, 760-781.
- van Wijk, R., Huizinga, E. G., Prins, I., Kors, A., Rijksen, G., Bierings, M., & van Solinge, W. W. (2004). Distinct phenotypic expression of two de novo missense mutations affecting the dimer interface of glucose-6-phosphate dehydrogenase. *Blood Cells, Molecules & Diseases*, 32(1), 112–117.

- Vaz, F. E., Thakur, C. B., Banerjee, M. K., & Gangal, S. G. (2000). Distribution of beta-thalassemia mutations in the Indian population referred to a diagnostic center. *Hemoglobin*, 24(3), 181–194.
- Vela-Amieva, M., Alcántara-Ortigoza, M. A., Gonzalez-Del Angel, A., Belmont-Martínez, L., López-Candiani, C., & Ibarra-González, I. (2021). Genetic spectrum and clinical early natural history of glucose-6-phosphate dehydrogenase deficiency in Mexican children detected through newborn screening. *Orphanet Journal of Rare Diseases*, 16(1), 103.
- Verma, C., Wijnen, J. T. H., & Khan, P. M. (1987). G6PD Punjab, a dialysis sensitive variant of human glucose-6-phosphate dehydrogenase. *Journal of Genetics*, 66(1), 17–24.
- Vives-Corrons, J. L., Kuhl, W., Pujades, M. A., & Beutler, E. (1990). Molecular Genetics of the glucose-6-phosphate dehydrogenase (G6PD) Mediterranean variant and description of a new G6PD mutant, G6PD Andalus^{1361A}. *American Journal of Human Genetics*, 47(3), 575–579.
- Volney, G., Tatusov, M., Yen, A. C., & Karamyan, N. (2018). Naphthalene toxicity: methemoglobinemia and acute intravascular hemolysis. *Cureus*, 10, e3147.
- Vulliamy, T. J., Dursot, M., Battistuzzi, G., Estrada, M., Foulkes, N. S., Martini, G., Calabro, V., Poggi, V., Giordanot, R., Town, M., Luzzatto, L., & Persico, M. G. (1988). Diverse point mutations in the human Glucose-6-phosphate dehydrogenase gene cause enzyme deficiency and mild or severe haemolytic anemia. *Proceedings of the National Academy of Sciences*, 85(14), 5171–5175.
- Vulliamy, T. J., Kaeda, J. S., Ait-Chafa, D., Mangerini, R., Roper, D., Barbot, J., Mehta, A. B., Athanassiou-Metaxa, M., Luzzatto, L., & Mason, P. J. (1998). Clinical and haematological consequences of recurrent G6PD mutations and a single new

- mutation causing chronic nonspherocytic haemolytic anaemia. *British Journal of Haematology*, 101(4), 670–675.
- Vulliamy, T., Luzzatto, L., Hirono, A., & Beutler, E. (1997). Hematologically important mutations: Glucose-6-phosphate dehydrogenase. *Blood Cells, Molecules and Diseases*, 23(2), 302–313.
- Wagner, G., Bhatia, K., & Board, P. (1996). Glucose-6-phosphate dehydrogenase deficiency mutations in Papua New Guinea. *Human Biology*, 68(3), 383–394.
- Wallner, B., & Elofsson, A. (2003). Can correct protein models be identified?. *Protein Science*, 12(5):1073-1086.
- Wang, J., Luo, E., Hirai, M., Arai, M., Manan, E. A. S. A., Isa, Z. M., Hidayah, N. I., & Matsuoka, H. (2008). Nine different glucose-6-phosphate dehydrogenase (G6PD) variants in a Malaysian population with Malay, Chinese, Indian and Orang Asli (Aboriginal Malaysian) backgrounds. *Acta Medicinæ Okayama*, 62, 327–332.
- Weatherall, D. J. (2001). Genetic disorders of haemoglobin. In Hoffbrand, A. V., Lewis, S. M., Tuddenham, E. G. D. (Eds.), *Postgraduate Haematology*. (4th ed., pp. 91–119). London, UK: Arnold Publishers.
- Weatherall, D. J. (2010). The inherited diseases of hemoglobin are an emerging global health burden. *Blood*, 115(22), 4331-4336.
- Weatherall, D. J. (2016). The thalasseмии: Disorders of globin synthesis. In Kaushansky, K., Lichtman, M. A., Prchal J. T. *et al.* (Eds.), *Williams hematology*. (9th ed., pp. 725–758). McGraw-Hill.
- Weatherall, D., Akinyanju, O., Fucharoen, S., Olivieri, N., and Musgrove, P. (2006). In Jamison, D. T., Breman, J. G., Measham A. R., *et al.* (Eds.). *Inherited disorders*

of hemoglobin. *Disease control priorities in developing countries*. (2nd ed.) International Bank for Reconstruction and Development/the World Bank; Oxford University Press.

Weatherall, D. J. (2008). Genetic variation and susceptibility to infection: The red cell and malaria. *British Journal of Haematology*, *141*(3), 276–286.

Weatherall, D. J., & Clegg, J. B. (2001). Inherited haemoglobin disorders: An increasing global health problem. *Bulletin of the World Health Organization*, *79*(8), 704–712.

WHO Working Group. (1989). Glucose-6-phosphate dehydrogenase deficiency. *Bulletin of the World Health Organization*, *67*(6), 601–611.

Wiederstein, M., & Sippl, M. J. (2007). ProSA-web: interactive web service for the recognition of errors in three - dimensional structures of proteins. *Nucleic Acids Research*, *35*(Web Server issue), W407–W410.

William, T. J. (1927). The error of determination of toxicity. *Proceedings of the Royal Society of London Series B*, *101*, 483–514.

Williams, T. N., & Weatherall, D. J. (2012). World distribution, population genetics, and health burden of the hemoglobinopathies. *Cold Spring Harbor Perspectives in Medicine*, *2*(9), a011692.

Williams, T. N., Wambua, S., Uyoga, S., Macharia, A., Mwacharo, J. K., Newton, C. R., and Maitland, K. (2005). Both heterozygous and homozygous alpha thalassemias protect against severe and fatal *Plasmodium falciparum* malaria on the coast of Kenya. *Blood*, *106*(1), 368–371.

Wong, F. L., Ithnin, A., Othman, A., & Cheah, F. C. (2017). Glucose-6-phosphate dehydrogenase (G6PD)-deficient infants: Enzyme activity and gene variants as risk factors for phototherapy in the first week of life. *Journal of Paediatrics and Child Health*, *53*(7), 705–710.

- World Health Organization (2016). Testing for G6PD deficiency for safe use of primaquine in radical cure of *P. vivax* and *P. ovale* malaria Policy brief. *WHO Document Production Services, Geneva, Switzerland*.
- World Health Organization (2021). Regional desk review of haemoglobinopathies with an emphasis on thalassaemia and accessibility and availability of safe blood and blood products as per these patients' requirement in South-East Asia under universal health coverage. *New Delhi: World Health Organization, Regional Office for South-East Asia*.
- Xiao, Wusheng, Wang, R.-S., Hand y, D. E., & Loscalzo, J. (2018). NAD(H) and NADP(H) redox couples and cellular energy metabolism. *Antioxidants & Redox Signaling*, 28(3), 251–272.
- Xu, W., Westwood, B., Bartsocas, C. S., Malcorra-Azpiazu, J. J., Indrak, K., & Beutler, E. (1995). Glucose-6 phosphate dehydrogenase mutations and haplotypes in various ethnic groups. *Blood*, 85(1), 257–263.
- Yan, T., Cai, R., Mo, O., Zhu, D., Ouyang, H., Huang, L., Zhao, M., Huang, F., Li, L., Liang, X., & Xu, X. (2006). Incidence and complete molecular characterization of Glucose-6-phosphate dehydrogenase deficiency in the Guangxi Zhuang autonomous region of southern China: Description of four novel mutations. *Haematologica*, 91(10), 1321–1328.
- Yang, J., Yan, R., Roy, A., Xu, D., Poisson, J., & Zhang, Y. (2015). The I-TASSER Suite: protein structure and function prediction. *Nature Methods*, 12(1), 7–8.
- Yang, Y., & Zhou, Y. (2008). Specific interactions for ab initio folding of protein terminal regions with secondary structures. *Proteins*, 72(2), 793–803.
- Youngster, I., Arcavi, L., Schechmaster, R., Akayzen, Y., Popliski, H., Shimonov, J., Beig, S., & Berkovitch, M. (2010). Medications and glucose-6-phosphate dehydrogenase deficiency: an evidence-based review. *Drug Safety*, 33(9), 713–726.

Zamvar, V., McClean, P., Odeka, E., Richards, M., & Davison, S. (2005). Hepatitis E virus infection with non-immune hemolytic anemia. *Journal of Pediatric Gastroenterology and Nutrition*, 40(2), 223–225.

Zhang Y. (2008). I-TASSER server for protein 3D structure prediction. *BMC Bioinformatics*, 9, 40.

PUBLICATIONS

CASE REPORT

Open Access

Compound heterozygosity for hemoglobin S and hemoglobin E in a family of Proto-Australoid origin: a case report



Noymi Basumatary¹, Dipankar Baruah², Paresh Kumar Sarma³ and Jatin Sarmah^{4*}

Abstract

Background: Hemoglobin S and E are commonly occurring hemoglobin variants among distinctly separate tribal populations of Central and Northeast India, respectively. Combined heterozygosity for hemoglobin S and E or hemoglobin SE disease is a benign clinical condition with rare incidence. Reports of approximately 46 hemoglobin SE cases are available worldwide. We conducted a screening program to study the prevalence of hemoglobin variants among the tribal population working in the tea estates of Northeast India. A total of 551 subjects were screened, and complete blood count was performed. Based on their hematological profiles, hemoglobin typing was done for 218 subjects.

Case presentation: We describe a case of an adolescent male of Munda tribe diagnosed as double heterozygous for hemoglobin S and E. On screening of the nuclear family of the subject, the mother was found to have hemoglobin E disease and father as hemoglobin S trait. Both siblings of the subject were diagnosed as hemoglobin E trait.

Conclusion: This is the first case of compound heterozygous for hemoglobin S and E to be reported from the tea tribes of Assam, India.

Keywords: Proto-Australoid, Hemoglobinopathy, Hemolytic anemia, Sickle cell disease, Tea tribe

Background

Hemoglobinopathies are monogenic disorders characterized by abnormal hemoglobin structure [1]. Among the hemoglobin variants, the most commonly occurring and clinically significant variants are hemoglobin S (Hb S), hemoglobin C (Hb C), hemoglobin E (Hb E), and thalassemia [2]. In context to its occurrence, Hb E is the second most common abnormal variant of hemoglobin in the world and most common variant in Southeast Asia [3]. Central-West Africa, East Asia, and India experience higher occurrence of sickle cell disease in comparison

with other parts of the world. Hemoglobinopathies are a cause of both economic and psychosocial burden [4]. Sickle cell disease shows an autosomal recessive inheritance resulting from A > T mutation in the sixth residue of the β -globin chain. Hb E results from a Glu→Lys mutation in the 26th amino acid.

Among the different types of hemoglobinopathies, prevalence of Hb S and Hb E in India is 4.3% and 10.9% respectively [5]. The burden of hemoglobinopathies in India is so high that it has become a major public health issue in some parts of the country [3]. In India, prevalence of Hb S among the tribals of central, southern, and western part has been reported [6]. In the eastern and northeastern part, Hb E is prevalent [7]. Sickle cell disease, particularly, has turned into a major health concern in states such as Chhattisgarh, Maharashtra, Gujarat,

*Correspondence: jatinsarmahindia@gmail.com

⁴ Department of Biotechnology and Co-ordinator, DBT (Govt. of India) sponsored Bioinformatics Infrastructure Facility, Bodoland University, Kokrajhar, Assam, India

Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Research Article

PREVALENCE AND ALTERATION IN HAEMATOLOGICAL PARAMETERS OF GLUCOSE-6-PHOSPHATE DEHYDROGENASE DEFICIENT PROTO-AUSTRALOID POPULATION OF MALARIA ENDEMIC HIMALAYAN BELT OF ASSAM, INDIA

Noymi Basumatary¹, Dipankar Baruah², Paresh Kumar Sarma², Manab Deka³, Jatin Sarmah^{1*}

Received 27.02.2021, revised 24.04.2021

ABSTRACT: Glucose-6-phosphate dehydrogenase (G6PD) deficiency is a common genetic disorder in malaria endemic regions and high among tribal population. To address the issue, the present study was framed to find out the prevalence of G6PD deficiency among the Proto-Australoid tribal population of malaria endemic Himalayan belt of Indo-Bhutan border areas of Assam and haematological changes in the target population. Screening for G6PD deficiency was done in 1436 normal individuals, out of which 6.62 percent (n=95) were found as deficient. Prevalence was higher in males (68.4%) compared to the females (31.6%). Complete Blood Count (CBC) was done in all samples. Further, analysis was performed to study the changes in the mean values of haematological parameters (both RBC and WBC indices) of G6PD normal and deficient subjects as well as between severe G6PD deficient and intermediate subjects. Subsequently association of haematological parameters with G6PD as well as between G6PD deficiency and gender was also studied. RBC indices viz., Hb, RBC and MCHC showed significant positive correlation with G6PD. No significant correlation was seen with WBC parameters.

Key words: Glucose-6-Phosphate Dehydrogenase, Proto-Australoid population, Malaria, Haemolytic anemia, Hematology.

INTRODUCTION

The enzyme Glucose-6-Phosphate Dehydrogenase (G6PD) is a catalyst in the conversion of glucose-6-phosphate into 6-phosphogluconate, a rate limiting step of pentose phosphate pathway (Stanton 2012). The NADPH produced in this step controls the supply of reduced glutathione (GSH) to the Red Blood Corpuscles (RBC). This in turn saves the RBCs from oxidative stress (Au *et al.* 2000, Eferth *et al.* 2005). More than 400 million people of the world are affected by the deficiency of this enzyme (Nkhoma *et al.* 2009). G6PD deficiency causes premature breakdown of RBCs which results in haemolytic anemia. Generally, G6PD deficient individuals do not show any symptoms or suffer from any harmful effects, but exposure to certain factors like consumption of fava beans or certain anti-malarial drugs may trigger haemolytic anemia (Mehta *et al.* 2000). More than 400 mutants of G6PD have been reported on the

basis of biochemical characterization and about 220 mutations are identified at DNA level (Gomez-Manzo *et al.* 2016). These mutations may result in changes in the protein structure thereby causing a decrease in its activity (Gomez-Manzo *et al.* 2014). The World Health Organization (WHO) has classified these mutations as Class I, II, III, IV and V based on the severity of the deficiency. Both class I and class II mutations show less than 10% enzyme activity causing chronic haemolytic anemia and periodic haemolysis respectively, and class III mutations exhibits 10-60% enzyme activity. The other two classes *i.e.*, IV and V show mild effect on the enzyme activity (WHO 1989).

The World Health Organization has established a population specific prevalence of G6PD deficiency in India ranging between 0-10%, with a higher prevalence among the tribal population (Tripathy and Reddy 2007). It has been estimated that a minimum of 3,90,000 children

¹Department of Biotechnology, Bodoland University, Kokrajhar, Assam, India,

²Fakhruddin Ali Ahmed Medical College and Hospital, Barpeta, Assam, India,

³Arya Vidyapeeth College, Guwahati, Assam, India.

*Corresponding author; e-mail: jatinsarmahindia@gmail.com

Carrier identification of Hemoglobin disorders among adolescents of Northeast India: Necessity of genetic counselling intervention

Noymi Basumatary

*Department of Biotechnology
Bodoland University, Kokrajhar, Assam, India*

Dipankar Baruah

*Department of Pathology
Gauhati Medical College and Hospital, Guwahati, Assam, India*

Paresh Kumar Sarma

*Department of Medicine
Fakhruddin Ali Ahmed Medical College and Hospital, Barpeta, Assam, India*

Jatin Sarmah*

*Department of Biotechnology
Bodoland University, Kokrajhar, Assam, India*

ABSTRACT

Among hereditary hemoglobin disorders, Sickle cell disease and β -thalassemia requires serious considerations since lifelong blood transfusion is required for management of such patients. Certain advanced interventions are developed; however, such interventions are very costly and require strict observance, thereby generating the need of low cost but effectual strategy. Carrier screening and genetic counselling for these disorders have been proven to be effective in certain countries. Thus, the present work was carried out to understand the burden of carriers of Hb disorders among the adolescents of Assam, Northeast India. We attempt to highlight the need for introduction of genetic counselling to minimize the genetic load in future generations. Hemoglobin S and hemoglobin E were the major hemoglobin variants encountered in the study population. Prevalence of hemoglobin disorder carriers was observed to be 10.7% out of the total population. Among the total carriers, 20.2% were sickle cell carriers while 76.06% were Hb E carriers. Others were β thalassemia and compound heterozygous S and E. Thus, an extensive carrier screening program in the region is much needed to identify the carriers among the pre-marital age group population and to educate them through genetic counselling about the risks associated with such diseases.

KEYWORDS

Hemoglobinopathy, thalassemia, carrier screening, genetic counselling, Northeast India.

I. INTRODUCTION

Hemoglobinopathies and thalassemias are the most commonly occurring monogenic disorders that affect a person's hemoglobin. This group of disorders exhibit autosomal recessive pattern of inheritance [1]. Every year around 0.3-0.4 million children are born with such disorders in both low and middle-income countries [2]. These

Available online at www.sciencedirect.com

ScienceDirect

journal homepage: www.elsevier.com/locate/mjafi

Correspondence/Letter to the Editor

Asymptomatic hemoglobin SE compound heterozygous hemoglobinopathy in an Aryan male from Northeast India

Dear Editor,

Hemoglobin S (Hb S) is a commonly encountered hemoglobinopathy among the tribal of anthropologically distinct Proto-Australoid population of Central, Southern and Western part of India.¹ The tea estate employees of Assam were from mainland India, who were brought by the British rulers during the last part of the nineteenth century. Again, Hemoglobin E (Hb E) is common among the Mongoloid population of north-eastern India.² Compound heterozygosity for Hemoglobin S and E (Hb SE) is a clinically rare, silent, and benign condition; such cases have lower chances of being diagnosed. They are detected occasionally when the person is being tested for other complaints. Although rare, the Hb SE cases may suffer from intravascular sickling, vaso-occlusion, bone necrosis, and sudden acute chest syndrome. The clinicians involved in sickle cell disease management and pathologists engaged in clinical forensic medicine services may take these rare cases and symptoms into consideration for referral etc. To the best of our knowledge, the case presented here is the first report revealing the existence of compound heterozygosity for Hb S and E (Hb SE) in a person of Aryan origin.

During a screening program for different types of hemoglobinopathies among the tea estate daily wage laborers of Udalguri district, Assam of Northeast India, we detected the case of Hb SE trait in a clinically normal 45-year-old male of the Aryan race (Fig. 1a). We screened all members of the nuclear family. His wife and 21-year-old daughter were detected with Hb S trait, and his 13-year-old son was detected with transfusion-dependent Hb S disease (Fig. 1b, c, and d). Hematological profiles, namely Hb%, MCV, MCH, and MCHC of all members of the family, are presented in Table 1. Earlier generations of the case had natural death without any recorded medical history of hemoglobinopathies. As per the records from the District Marriage Registry Office, there was no racial

intermarriage of both the parents and their earlier three generations. Since the present family members could not provide information about the presence of hemoglobinopathy among their earlier generations, we could not prepare the family pedigree.

We could not find any report on the occurrence of Hb SE from the Aryan community. The Hb SE case presented here was of Aryan stock and was asymptomatic. The hematological profile revealed normal Hb% and MCV, raised RBC count, and lower MCH and MCHC. Hb typing showed 2.9% Hb F, 4.5% Hb A₂, 68.6% Hb S, and 24.0% Hb E. A few earlier cases reported from India also had normal Hb%, MCV, MCH, and MCHC levels.³ Although Hb SE is a clinically benign condition, sudden death after exercise was reported in a 12-year-old American Hb SE patient. In this case, approximately after 1 h and 45 min of exercise, the patient collapsed; hemodynamic instability and cardiac arrest were reported as per postmortem report, which is assumed to be due to tissue ischemia caused by intravascular sickling. He also had records of ventricular septal cardiac defect at birth, mild asthma, and fractures of the radius and ulna 17 months prior to death.⁴ Earlier, another case of Hb SE in a 15-year-old Teli community male was reported from central part of India. He was suffering from recurrent upper respiratory tract infections, weakness, and tiredness.⁵

Again, a marriage between Hb SE trait father and Hb S trait mother had Hb S disease son and Hb S trait daughter. The subject's wife was from Proto-Australoid race (tea tribe). She had normal Hb% and RBC, lower MCV, MCH, and MCHC levels. The son was dependent on periodic blood transfusions. He had low Hb%, RBC count, and MCHC levels. The daughter had low Hb%, RBC, MCV, MCH, and MCHC levels, which is indicative of microcytic hypochromic anemia due to borderline low serum iron (65 microgram/dl). This may be due to multiple factors, including menstrual loss, low dietary intake, etc.



The First Report of Three Glucose-6-Phosphate Dehydrogenase (G6PD) Variants: Mediterranean, Orissa and Kalyan-Kerala from Northeast India

Noymi Basumatary¹ · Dipankar Baruah² · Paresh Kumar Sarma³ ·
Kishore Kumar Wary⁴ · Jatin Sarmah¹

Received: 17 March 2023 / Accepted: 30 July 2023

© The Author(s), under exclusive licence to Indian Society of Hematology and Blood Transfusion 2023

Dear Editor,

The most common enzymopathy prevalent in malaria-endemic regions is glucose-6-phosphate dehydrogenase (G6PD) deficiency. This deficiency affects over 400 million people worldwide. The global prevalence of G6PD deficiency is geographically associated with malaria endemic areas. The Northeastern region of India is malaria endemic, accounting for a large proportion of India's malaria incidences each year [1]. G6PD deficiency is a result of mutations in the DNA encoding G6PD enzyme. The significance of this enzyme lies in its involvement in the generation of NADPH, an antioxidant, during the process of the pentose phosphate pathway. Through subsequent processes, NADPH provides protection to the RBCs against oxidative damage. Mutations in the G6PD gene cause the G6PD enzyme to lose its stability and activity, thereby making deficient RBCs more prone to oxidative damage. G6PD deficiency is widespread across Africa, Asia, the Mediterranean, and the Middle East. Approximately 7.5% of the world population is carriers of G6PD deficiency, and 2.9% are deficient according to the World Health Organization (WHO) report. Moreover, a higher frequency of G6PD deficiency (5.5%) was reported among the tribal/scheduled caste population of India [2, 3]. Molecular

genetic analysis has revealed about 220 different mutations in the G6PD gene. In India, the most commonly occurring variant is G6PD Mediterranean, followed by G6PD Orissa, and Kalyan-Kerala. However, these variants have not been reported earlier from the Northeastern region of India, which is predominantly inhabited by Mongoloid and Proto-Australoid population and few from Aryan stock. On the other hand, variants like G6PD A⁺, Mahidol and Acores are previously reported from this part of the country [4, 5].

This is the first report to document the evidence of G6PD Mediterranean, Orissa, and Kalyan-Kerala from the tribal population of Northeast India. The variants were detected in a study conducted in four districts of Assam viz., Kokrajhar, Chirang, Baksa and Udalguri which form the Bodoland Territorial Region (BTR) and share its boundaries with Bhutan. The tribes included in the study were Bodo, Rabha, Garo and the tea tribe. G6PD deficiency was detected in 6.2% of the total population with highest number of cases from the tea tribe population. The variants were detected using Polymerase Chain Reaction–Restriction Fragment Length Polymorphism (PCR–RFLP) technique. Restriction endonucleases MboII, HaeIII and MnlI were used for Mediterranean, Orissa, and Kalyan-Kerala respectively [6, 7]. India has a diverse population admixture of various ethnic groups each of which differs in their physical, cultural and genetic background. Approximately 8.6% of the country's population is tribal. The Northeast India alone has more than 200 different ethnic groups belonging to Mongoloid and Proto-Australoid origin. Individuals of Mongoloid and Proto-Australoid descents were found to harbor the variants described in this report. G6PD Mediterranean and Kalyan-Kerala variants were detected in both Mongoloids and Proto-Australoids. While G6PD Orissa was observed exclusively in the Proto-Australoids. All the detected cases of G6PD deficiency were asymptomatic.

✉ Jatin Sarmah
jatinsarmahindia@gmail.com

¹ Department of Biotechnology, Bodoland University, Kokrajhar, Assam, India

² Department of Pathology, Gauhati Medical College and Hospital, Guwahati, Assam, India

³ Department of Medicine, Dhubri Medical College and Hospital, Dhubri, Assam, India

⁴ Department of Pharmacology and Regenerative Medicine, University of Illinois, Chicago, USA

A Case Report of Co-occurrence of Hemoglobinopathy EE and Glucose-6-phosphate Dehydrogenase A+ Variant

Noymi Basumatary¹, Dipankar Baruah², Paresh Kumar Sarma³, Jatin Sarmah¹

¹Department of Biotechnology, Bodoland University, Kokrajhar, ²Department of Pathology, Gauhati Medical College and Hospital, Guwahati, ³Department of Medicine, Dhubri Medical College and Hospital, Dhubri, Assam, India

Abstract

Hemoglobinopathies and glucose-6-phosphate dehydrogenase (G6PD) deficiency are two genetic disorders prevalent in malaria-endemic regions. There are conflicting reports on the co-occurrence of G6PD deficiency and hemoglobinopathies. The present study was conducted to explore the co-occurrence (if any) of the two disorders among ethnic populations of Proto-Australoid and Mongoloid origin races. Out of 2310 subjects screened, only one case was detected with both disorders. The case described here is an asymptomatic female of 52 years detected with both hemoglobinopathy EE (HbEE) and G6PD A+ variant. Lower levels of hemoglobin (Hb) %, mean corpuscular hemoglobin (MCH), MCH concentration, platelets, and white blood cell count, and high red cell distribution width and fetal Hb% were recorded. Family screening revealed both daughters as HbAE and normal G6PD. The husband had HbAA and normal G6PD. Out of six siblings of the case subject, two were detected as HbEE and four as HbAE; G6PD was normal.

Keywords: Glucose-6-phosphate dehydrogenase A+, glucose-6-phosphate dehydrogenase deficiency, hemoglobinopathies, hemolysis, polymerase chain reaction–restriction fragment length polymorphism

INTRODUCTION

Hemoglobinopathies and glucose-6-phosphate dehydrogenase (G6PD) deficiency are two genetically inherited disorders prevalent in malaria-endemic regions of the world. Although generally asymptomatic, G6PD-deficient individuals may experience hemolytic anemia when exposed to certain factors that act as hemolytic triggers.^[1] Among these triggers, primaquine, an antimalarial drug, induces severe hemolysis in G6PD-deficient people.^[2] Hemoglobin E (HbE) is a nontransfusion-dependent form of hemoglobinopathy, caused by Glu→Lys replacement at the 26th codon of the β -globin gene. Homozygous HbE is characterized by mild anemia with hypochromic microcytic red blood cells. HbE is the most prevalent Hb variant in North-east India.^[3] Although there are limited studies on G6PD deficiency from this region, the prevalence of G6PD deficiency is reported to be 5.4%.^[4] There are two schools of viewpoints on the association between G6PD deficiency and hemoglobinopathies. According to one school, a greater co-occurrence of G6PD deficiency and hemoglobinopathies was observed in regions with high frequencies of both genes. The second school of workers

reported that G6PD deficiency and hemoglobinopathies are genetically independent disorders that assort independently.^[5]

METHODS

Ethical consideration

Ethical agreement and approval for the study were obtained from the Institutional Ethics Committee of Bodoland University, Kokrajhar, Assam, India, vide Reference No.: IEC/BU/ICMR/2019-2 dated May 10, 2019.

Type of sampling and reasons for selection

Random sampling was done using STANDARD G6PD Analyzer (SD Biosensors) for identification of subjects with abnormality in either Hb or G6PD value, or both.

Address for correspondence: Prof. Jatin Sarmah,

Department of Biotechnology, Bodoland University, Kokrajhar, Assam, India.
E-mail: jatinsarmahindia@gmail.com

ORCID: <https://orcid.org/0000-0001-8939-0495>

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Basumatary N, Baruah D, Sarma PK, Sarmah J. A case report of co-occurrence of hemoglobinopathy EE and glucose-6-phosphate dehydrogenase A+ variant. *Biomed Biotechnol Res J* 2023;7:646-8.

Submitted: 05-Aug-2023;

Revised: 28-Sep-2023;

Accepted: 04-Oct-2023;

Published: 23-Dec-2023.

Access this article online

Quick Response Code:



Website:

<https://journals.lww.com/BBRJ>

DOI:

10.4103/bbrj.bbrj_228_23



ELSEVIER

Contents lists available at ScienceDirect

Gene Reports

journal homepage: www.elsevier.com/locate/genrep

Identification of glucose-6-phosphate dehydrogenase variants by utilizing polymerase chain reaction – Restriction fragment length polymorphism based method

Noymi Basumatary^a, Dipankar Baruah^b, Paresh Kumar Sarma^c, Jatinder Sarmah^{a,*}

^a Department of Biotechnology, Bodoland University, Assam, India

^b Department of Pathology, Gauhati Medical College & Hospital, Assam, India

^c Department of Medicine, Dhubri Medical College & Hospital, Assam, India

ARTICLE INFO

Edited by Ziqing Li

Keywords:

G6PD
PCR-RFLP
Restriction endonucleases
G6PD variants
Malaria
Primaquine

ABSTRACT

Glucose-6-phosphate dehydrogenase (G6PD) deficiency is a widespread genetically inherited enzyme disorder caused by mutation in the *G6PD* gene located in X chromosome. Although sequencing of the gene is considered as the standard method for detection of mutations, but the process is expensive, laborious, time-consuming and not possible to perform in low resource settings. Thus, the present work was aimed to identify the G6PD variants using Polymerase Chain Reaction – Restriction Fragment Length Polymorphism (PCR-RFLP) based method among the ethnic groups of malaria endemic Northeastern part of India. STANDARD G6PD Analyzer was utilized for screening to identify the G6PD deficient malarial patients. Molecular analysis of the deficient samples was done using PCR-RFLP technique. A total of 17 variants were explored using different restriction endonucleases. Five variants viz., Orissa (63.8%), Kalyan-Kerala/Jamnagar/Rohini (9.02%), Mediterranean (8.3%), A⁺ (11.8%) and Mahidol (1.38%) were detected among the patients. G6PD Orissa was the major variant among the study population. In malaria endemic regions like Northeast India, a mass screening program for identification of G6PD variants needs to be adopted.

1. Introduction

Malaria is one of the infectious diseases that cause high morbidity and mortality rates in the world's tropical regions. According to the World Health Organization (WHO), there were an estimated 249 million malarial cases worldwide in 2022 (WHO, 2023). In India, various species of malarial parasites such as *Plasmodium malariae*, *P. vivax*, *P. ovale* and *P. falciparum* are prevalent. In case of *P. vivax* infection, the persistent liver stages of the parasite (hypnozoites) may lead to disease relapse weeks or months after the initial infection (White, 2011). To prevent such relapses, the only way is inhibiting the reactivation of hypnozoites. The 8-aminoquinolines, including primaquine, stands as the sole anti-malarial drug that can prevent reactivation of the hypnozoites (von Seidlein et al., 2013). However, its use poses haemolysis risk, especially in patients with Glucose-6-phosphate dehydrogenase (G6PD) deficiency (Baird et al., 2016). As per the WHO, a recommended safe dose of primaquine is 0.75 mg base/kg body weight weekly for eight weeks to prevent *P. vivax* and *P. ovale* malaria relapse in G6PD deficient

patients, requiring close medical supervision. Again, in situations where the G6PD status is unknown and testing is unavailable, prescribing primaquine should involve a careful evaluation of risks and benefits of administering primaquine (WHO, 2016).

G6PD deficiency, the most common hereditary enzyme disorder in malaria-endemic regions, impacts over 400 million people of the world (Nkhoma et al., 2009). While G6PD deficient patients are partially protected against malaria, experiencing lower susceptibility to the disease, they face a significantly higher risk of haemolysis upon treatment with anti-malarial drugs like primaquine (Peters and Van Noorden, 2009). The presumed mechanism behind the protection is the reduced replication of the malarial parasite in the G6PD deficient red blood cells (RBCs). Due to either being phagocytosed or a shortened lifespan of G6PD deficient RBCs, the malarial parasite is unable to complete its life cycle within them. This observation of malaria protection, however, is not uniformly consistent across all G6PD defective genotypes (Guindo et al., 2007; Badoum et al., 2019; Johnson et al., 2009). G6PD deficiency occurs as a result of mutations in the G6PD enzyme encoding gene, *G6PD*

* Corresponding author.

E-mail address: jatindersarmahindia@gmail.com (J. Sarmah).

<https://doi.org/10.1016/j.genrep.2024.101911>

Received 30 September 2023; Received in revised form 5 March 2024; Accepted 20 March 2024

Available online 22 March 2024

2452-0144/© 2024 Elsevier Inc. All rights reserved.

**Eurasian Conference on
'Science, Engineering & Technological Innovations'**

Date: 20 & 21 November, 2021. (Online mode)

Jointly organized by

Kryvyi Rih National University, Ukraine, Europe
Automation, Computer Science and Technology Department
Research Culture Society

&
Co-Sponsored by : Scientific Research Association.



Certificate of Participation and Presentation

Ref.No: RCS/ECSETI-21/PPP/006

This is to Certify that

Noymi Basumatary

has participated and presented a Paper titled

A study on acceptability of genetic counselling intervention for hereditary hemoglobin disorders among adolescents of Northeast India

in the 'Eurasian Conference on Science, Engineering & Technological Innovations' dated 20 - 21 November, 2021.



Prof. Natalia Morkun

ECSETI-2021 Conference Chair

Head, Automation, Computer Science and Technology
Department, Kryvyi Rih National University, Ukraine.

www.knu.edu.ua

Dr. C. M. Patel

ECSETI-2021 Conference Chair

Director, Research Culture Society.
President, Scientific Research Association.

www.researchculturesociety.org

**CERTIFICATE
OF APPRECIATION**

**International Conference on
BIOTECHNOLOGY FOR ENVIRONMENT & HEALTH**

This certificate is presented to

Noymi Basumatary

from

Bodoland University

for the best oral presentation for the paper titled "Prevalence of Glucose-6-Phosphate Dehydrogenase (G6PD) deficiency and comparison of hematological parameters among G6PD deficient and normal Proto- Australoid tribal population of malaria endemic Indo-Bhutan border areas of Assam" in the International Conference on Biotechnology for Environment & Health (ICBEH), organized by School of Bio Sciences and Technology, Vellore Institute of Technology along with Association of Biotechnology and Pharmacy, from 25 - 27 November 2021.

K M Gothandam
Convenor, ICBEH

V Pragasam
Chairperson, ICBEH

K R S Sambasiva Rao
General Secretary, ABAP

**NATIONAL CONFERENCE
ON
SCIENCE & TECHNOLOGY FOR SUSTAINABLE DEVELOPMENT
(STSD-2022)**

Organized jointly by
SCIENCE COLLEGE, KOKRAJHAR & VIJNANA BHARATI, NESM

In collaboration with
IASST, Guwahati, India & NECTAR, Shillong, India








Certificate of Presentation
This is to certify that
Ms. Noymi Basumatary
of
Bodoland University, Kokrajhar
has presented a paper entitled "Global burden of Glucose-6-phosphate dehydrogenase (G6PD) deficiency- a lesser known asymptomatic genetic disorder: a pilot study from Bodoland Territorial Region (BTR), Assam." in the National Conference on "Science & Technology for Sustainable Development (STSD-2022)" held on 9th-10th September, 2022 at Science College, Kokrajhar, Assam, India.




Principal
Science College, Kokrajhar


President
Vijnana Bharati, NESM


Chairperson
National Conference
STSD-2022


Convener
National Conference
STSD-2022


Convener
National Conference
STSD-2022


Organizing Secretary
National Conference
STSD-2022







**NATIONAL SEMINAR
ON
RECENT ADVANCES IN BIOLOGICAL SCIENCES: BIODIVERSITY & HUMAN WELFARE**
November 18-19, 2022

Sponsored by: DST SERB, Govt. of India & ASTEC
Organized by: Department of Zoology, Darrang College, Tezpur, Assam
In collaboration with: Zoological Society of Assam & IQAC-Darrang College, Tezpur

CERTIFICATE OF PARTICIPATION

This is to certify that Dr./ Mr./Ms. *Noymi Basumatary*.....
of *Bodoland University*..... has participated/presented a paper titled
Understanding the structural An in silico analysis.....in the DST SERB,
Govt. of India sponsored National seminar on "Recent Advances in Biological Sciences: Biodiversity & Human Welfare" held from 18th to 19th November, 2022 at Darrang College, Tezpur Assam, India.


Dr. Palash Moni Saikia
President, Organizing Committee
& Principal, Darrang College


Dr. Jogen Ch. Kalita
General Secretary, ZSA
Zoological Society of Assam


Dr. Swapnalee Kalaty
Coordinator, IQAC
Darrang College, Tezpur


Dr. Chittaranjan Baruah
Convener

APPENDICES

Appendix – I

Ethical clearance certificate



INSTITUTIONAL ETHICS COMMITTEE
Bodoland University, Kokrajhar, Assam, India.
PIN-783370. Phone No:-03661-277183

Ref. No:-IEC/BU/ICMR/2019-2

Date:-10.05.2019

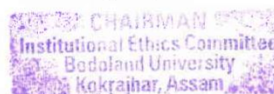
CERTIFICATE

The Meeting of the Institutional Ethics Committee was held on 10th May, 2019. The Committee considered the research proposal entitled “**A study on Glucose-6-Phosphate Dehydrogenase (G6PD) variants in Sickle Cell Anaemic subjects among the tea tribes of malaria endemic Indo-Bhutan border districts of India**” submitted by Dr. Jatin Sarmah, Associate Professor, Department of Biotechnology under Faculty of Science and Technology, Bodoland University, Kokrajhar, Assam and presented by his research scholar Miss Noymi Basumatary for her research work leading to Ph. D. degree in the Department of Biotechnology, Bodoland University. **The study is sponsored by Lady Tata Memorial Trust Scholarship to Miss Noymi Basumatary.**

The committee did not find anything objectionable / unethical *vis-a-vis* human subjects in this research proposal. The proposed research work is, therefore, awarded ethical clearance.


(PRADIP KUMAR PATRA)

Chairman,
Institutional Ethics Committee,
Bodoland University,
Kokrajhar, Assam.



Appendix – II

Consent form

Proposed work on Glucose-6-Phosphate Dehydrogenase (G6PD) variants in sickle cell anaemic subjects among the tea tribes of malaria endemic Indo-Bhutan border districts of India.

Principal Investigator/ Guide: Dr. Jatin Sarmah
Department of Biotechnology
Bodoland University
Kokrajhar, Assam

Informed Consent for Genetic Analysis

By signing below, I hereby authorize Dr. Jatin Sarmah to obtain
blood sample from [Redacted]
(Nature) (Patient Name) ✓
of Udalguri, Assam, India.
(Address)
for the genetic analysis of Saemoglobinopathies and G6PD deficiency.
(Disease)

It has been explained to me and I have understood that:

- * DNA testing, which is done on a small sample of blood or other tissue, looks at one or more parts of the particular gene associated with above disease. Genes contain the information that guides proper functioning of the body. A change in genetic information can result in an abnormal gene that doesn't work properly.
- * Most often DNA testing directly detects the most common disease-causing changes in a gene, the test result is highly accurate (~98%). In other cases, an indirect method called linkage analysis is used which may produce an (3-5%) uncertainty in predicting carrier status or diagnosis due to naturally occurring rearrangements in the DNA (recombination). Rare variations in individuals can also cause uncertainty in the results. In other words, the test is not 100% accurate.
- * The linked markers may not be informative in some families and hence, this DNA test can not provide results for the family, or for some members of that family.
- * Based on the results from initial tests, subsequent analysis may be performed on the sample for better understanding of the disease.
- * If the DNA testing does not show a known genetic change, the probability that the person is a carrier or is affected is reduced. However, there is still a small chance to be a carrier or to be affected because the current testing cannot find all the possible changes within a gene.
- * The accuracy of DNA analysis is entirely dependent on clinical diagnosis made elsewhere and Bodoland University cannot be responsible for erroneous clinical diagnosis or sample related problems made at other centres.
- * Unknown genetic changes are different in different populations. Providing the laboratory with accurate information about family history and ethnic background will make the interpretation of the test results more accurate.
- * The results of this testing will be disclosed ONLY to the patient/relative specified by the patient/doctor named above and to associated medical personnel.
- * A part of this sample not used for diagnostic testing may be stored and used for medical research or education as long as any names and other identifying information have been removed. In some cases, it may be possible to reanalyse the leftover samples in the future using new and improved methods.
- * The clinical information/video/photographic material resulting from the tests may be used for publication.

[Redacted]
Signature

Relationship to patient: self/parent/guardian

[Redacted]
Name Date

[Redacted]
Signature of witness

[Redacted]
Name Date

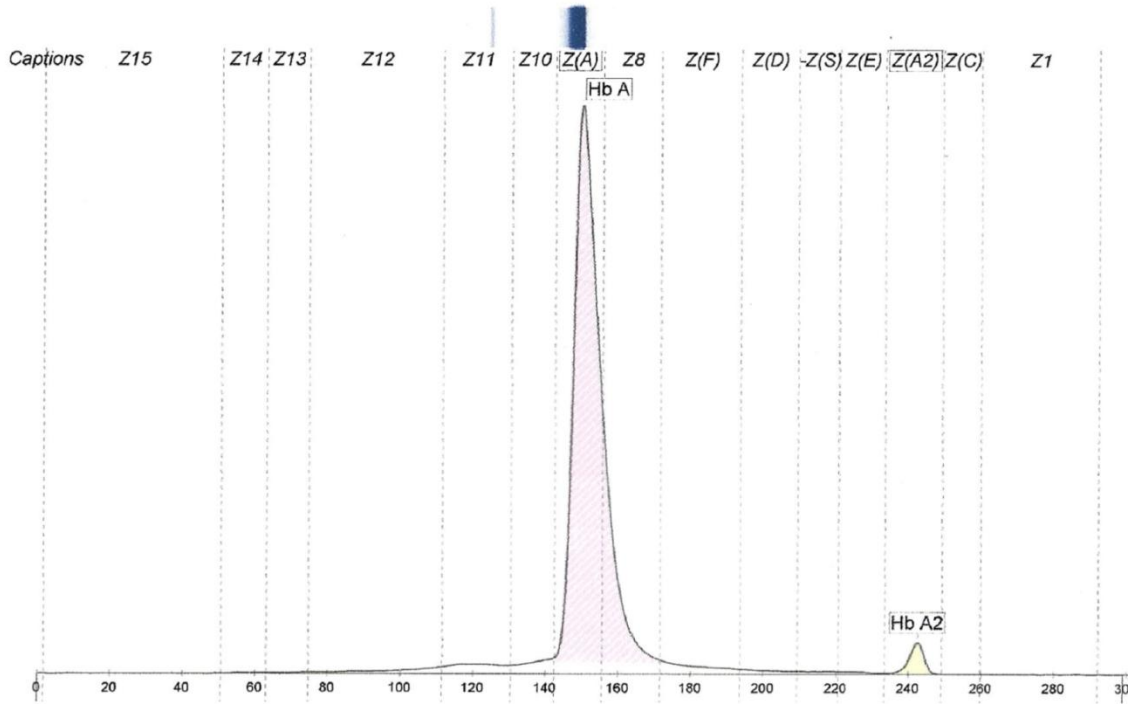
Appendix – III

CBC report

<u>Investigation</u>	<u>Result</u>	<u>Unit</u>	<u>Bio. Ref. Interval</u>
Haemoglobin (Hb)	: 10.5	gm%	13 - 17
Total Leucocyte Count	: 8200	Cells/Cumm	4000 - 11000
<u>DIFFERENTIAL COUNT</u>			
Polymorphs	: 40	%	35 - 70
Lymphocytes	: 49	%	20 - 40
Monocytes	: 08	%	02 - 08
Eosinophils	: 03	%	01 - 06
Basophils	: 00	%	00 - 02
Platelet Count	: 192	10 ³ uL	150 - 400
Total RBC Count	: 4.6	Million /Cumm	4.5 - 5.5
PCV	: 35	%	45 - 54
MCV	: 74	fl	76 - 98
MCH	: 22	pg	27 - 32
MCHC	: 30	gm/dl	32 - 36
RDW	: 15	%	11 - 14

Appendix – IV

Hb-typing report showing normal Hb type



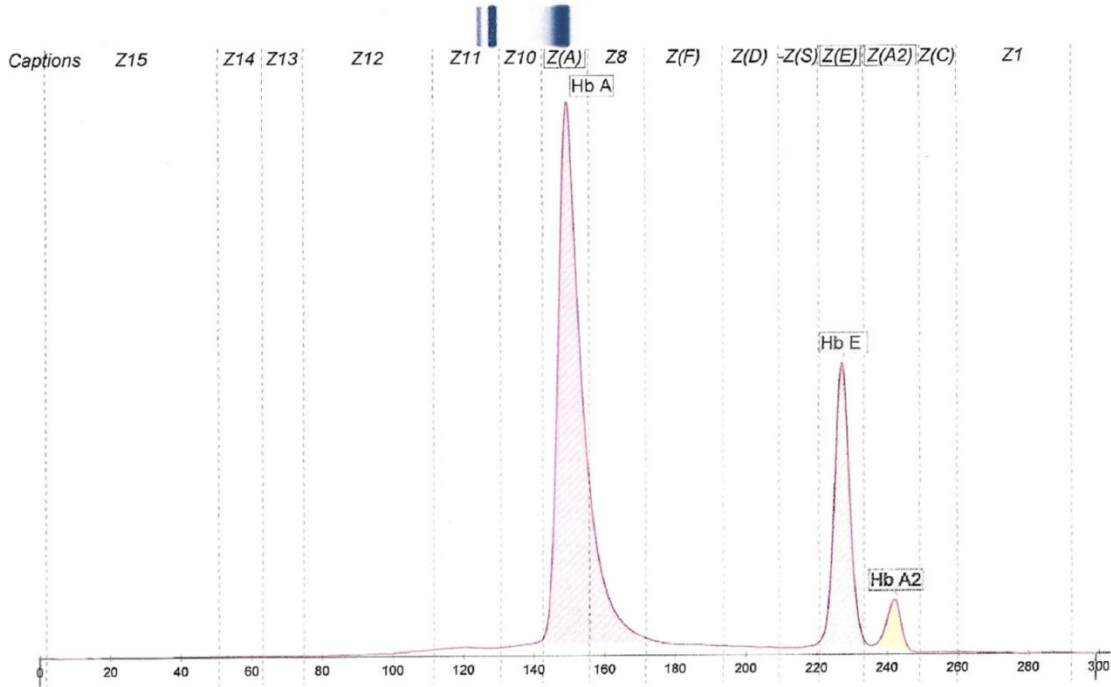
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb A	97.2	96.8 - 97.8
Hb A2	2.8	2.2 - 3.2

Comment:- Normal chromatogram.No hemoglobinopathy detected.

Appendix – V

Hb-typing report showing HbE trait



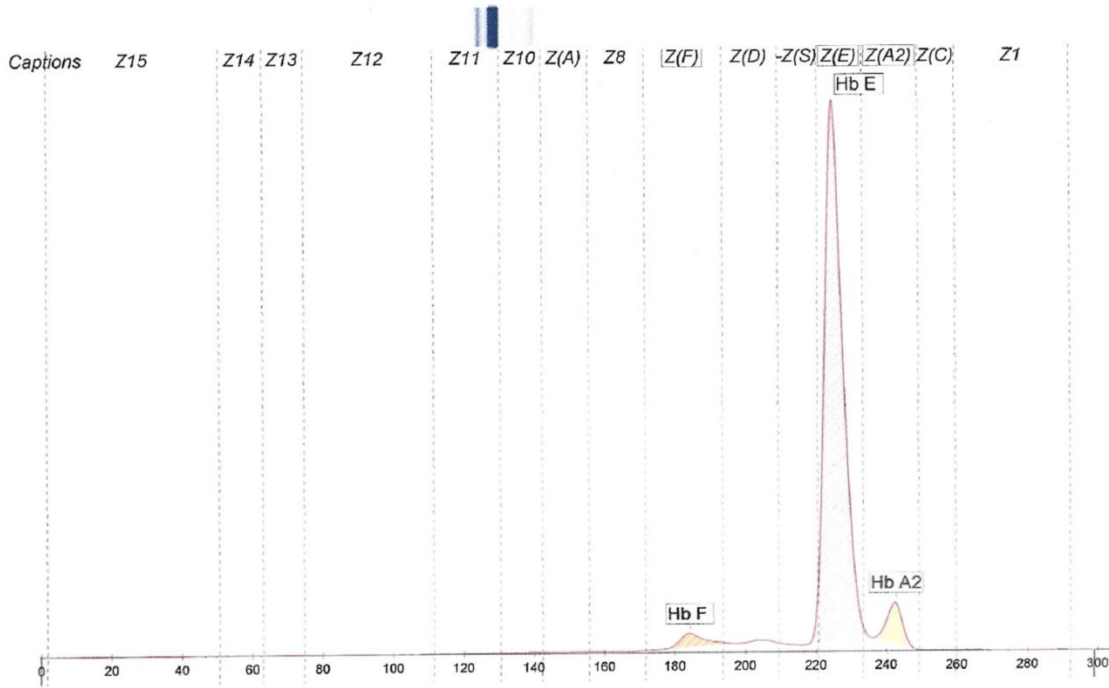
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb A	72.4	
Hb E	23.8	
Hb A2	3.8	

Comment:- Chromatogram is suggestive of Hb E heterozygous.(E Trait)

Appendix – VI

Hb-typing report showing HbE disease



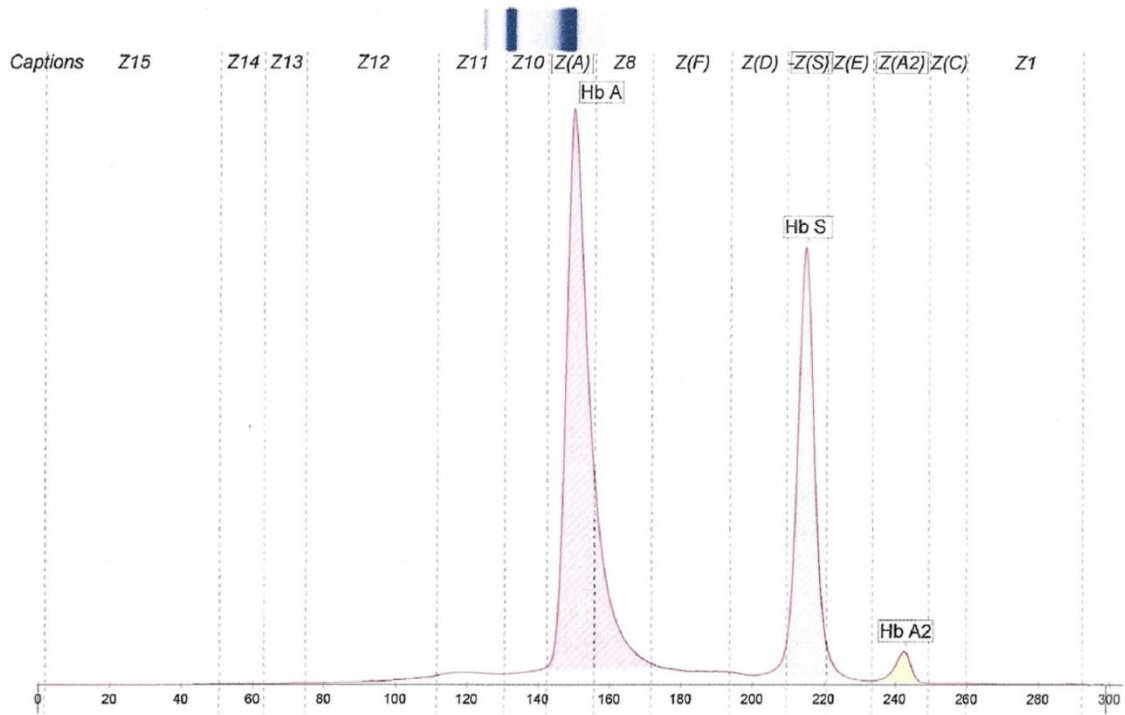
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb F	2.9	
Hb E	91.6	
Hb A2	5.5	

Comment:- Chromatogram is suggestive of Hb E homozygous.(E disease)

Appendix – VII

Hb-typing report showing HbS trait



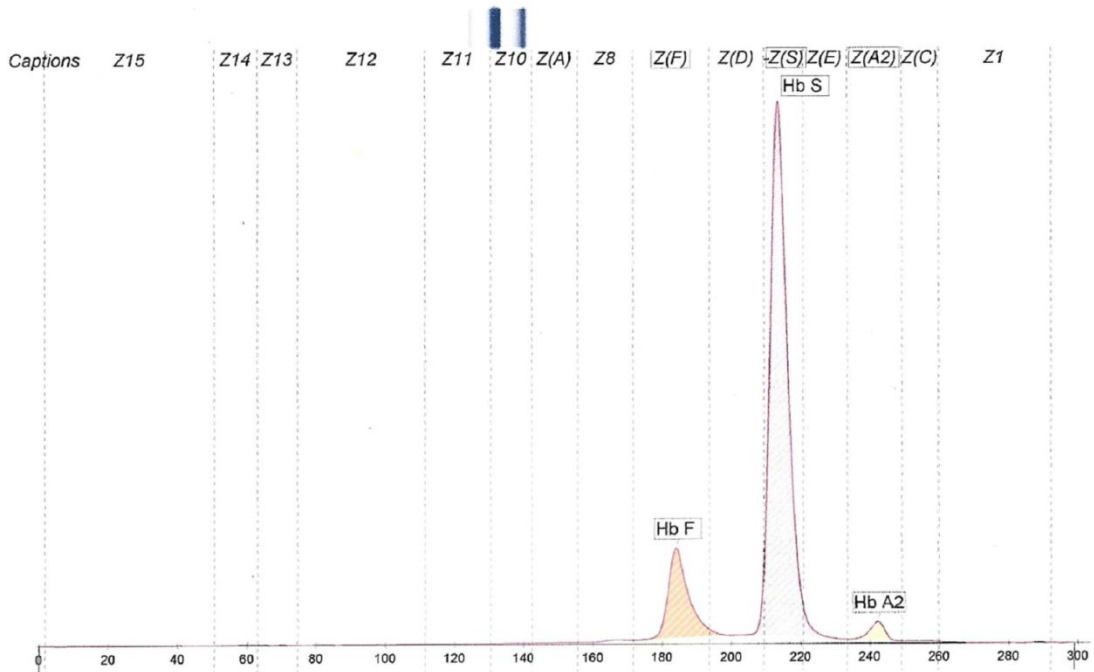
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb A	63.3	
Hb S	34.5	
Hb A2	2.2	

Comment:- Chromatogram is suggestive of Hb S heterozygous.(S Trait)

Appendix – VIII

Hb-typing report showing HbS disease



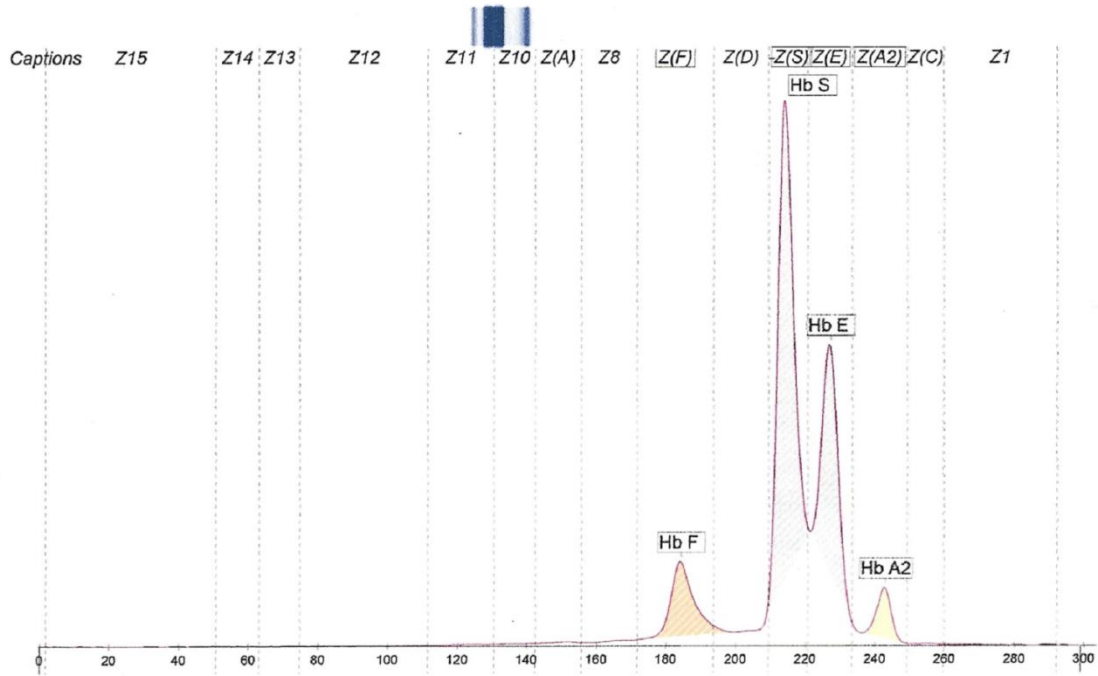
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb F	16.1	
Hb S	81.5	
Hb A2	2.4	

Comment:- Chromatogram is suggestive of Hb S homozygous.(S disease)

Appendix – IX

Hb-typing report showing compound heterozygous HbE and HbS trait



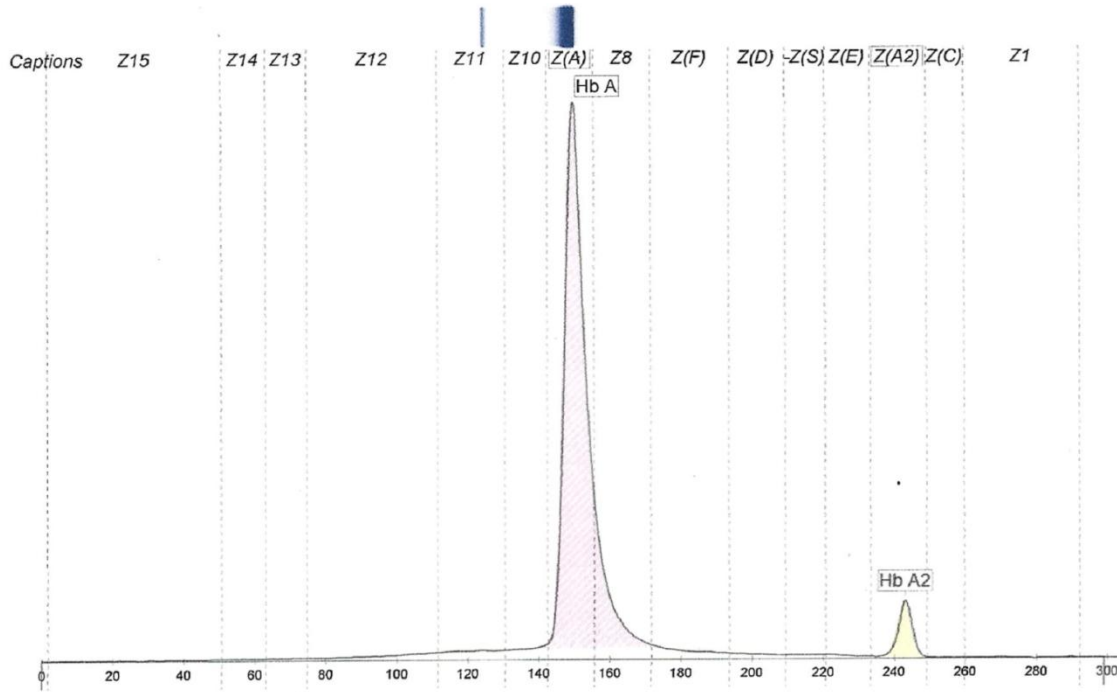
Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb F	12.7	
Hb S	55.2	
Hb E	26.7	
Hb A2	5.4	

Comment:- Chromatogram is suggestive of double heterozygous for Hb E and Hb S trait.

Appendix – X

Hb-typing report showing beta-thalassaemia trait



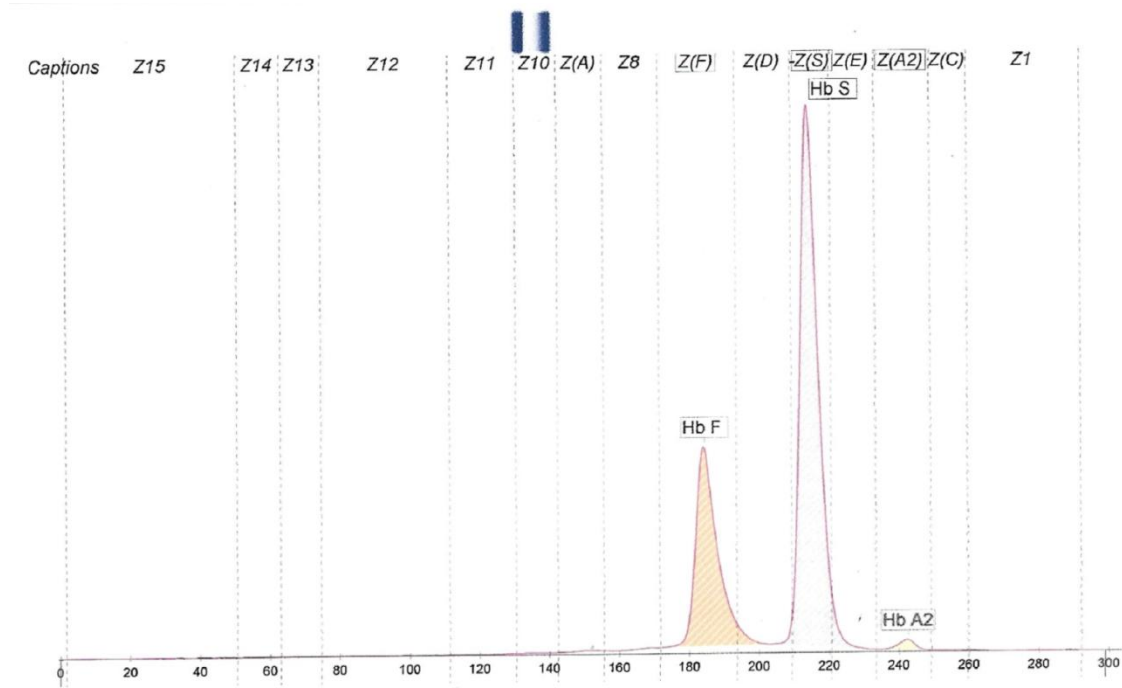
Hemoglobin Electrophoresis

Name	%		Normal Values %
Hb A	94.3	<	96.8 - 97.8
Hb A2	5.7	>	2.2 - 3.2

Comment:- Chromatogram is suggestive of beta thalassaemia trait.

Appendix – XI

Hb-typing report showing high HbF



Hemoglobin Electrophoresis

Name	%	Normal Values %
Hb F	30.7	
Hb S	68.2	
Hb A2	1.1	

Comment:- Chromatogram is suggestive of Hb S homozygous.(S disease)