REFERENCES

- Abdullah, B. M., Mehdi, M. A., Khan, A. R., & Pathan, J. M. (2020). Gas Chromatography-Mass Spectrometry (GC-MS) Analysis of Ajwain (*Trachyspermum ammi*) Seed Extract. *International Journal of Pharmaceutical Quality Assurance*, 11(2), 228–231. http://dx.doi.org/10.25258/ijpqa.11.2.6
- Abraham, J., Chakraborthy, P., Chand, A., Srivastava, S., Yadav, R., & Kingsley, D. (2016). In vitro analysis of antimicrobial compounds from *Alstonia scholaris*. *Asian Journal of Pharmaceutical and Clinical Research*, 9(5), 81–84. https://doi.org/10.22159/ajpcr.2016.v9i5.12173
- Adams, C. M., Caporn, S. J. M., & Hutchinson, T. C. (1990). Crystal occurrence and wax disruption on leaf surfaces of cabbage treated with simulated acid rain. *The New Phytologist*, 114(1), 147–158. https://doi.org/10.1111/j.1469-8137.1990.tb00385.x
- Adamska, A., Rumijowska-Galewicz, A., Ruszczynska, A., Studzińska, M., Jabłońska, A., Paradowska, E., Bulska, E., Munier-Lehmann, H., Dziadek, J., Leśnikowski, Z. J., & Olejniczak, A. B. (2016). Anti-mycobacterial activity of thymine derivatives bearing boron clusters. *European Journal of Medicinal Chemistry*, 121, 71–81. https://doi.org/10.1016/j.ejmech.2016.05.030
- Adnan, M., Nazim Uddin Chy, M., Mostafa Kamal, A. T. M., Azad, M. O. K., Paul, A., Uddin, S. B., Barlow, J. W., Faruque, M. O., Park, C. H., & Cho, D. H. (2019). Investigation of the Biological Activities and Characterization of Bioactive Constituents of *Ophiorrhiza rugosa* var. *prostrata* (D.Don) & Mondal Leaves through In Vivo, In Vitro, and In Silico Approaches. *Molecules*, 24(7), 1367. https://doi.org/10.3390/molecules24071367
- Agarwal, S., Jangir, D. K., Singh, P., & Mehrotra, R. (2014). Spectroscopic analysis of the interaction of lomustine with calf thymus DNA. *Journal of Photochemistry and Photobiology.* B, Biology, 130, 281–286. https://doi.org/10.1016/j.jphotobiol.2013.11.017
- Ahmad, S., Ullah, F., Sadiq, A., Ayaz, M., Imran, M., Ali, I., Zeb, A., Ullah, F., & Shah, M. R. (2016). Chemical composition, antioxidant and anticholinesterase potentials of essential oil of *Rumex hastatus* D. Don collected from the North West of Pakistan. *BMC*

- Complementary and Alternative Medicine, 16, 29. https://doi.org/10.1186/s12906-016-0998-z
- Ahmed, M., Khan, K.-u.-R., Ahmad, S., Aati, H. Y., Ovatlarnporn, C., Rehman, M. S.-u., Javed, T., Khursheed, A., Ghalloo, B. A., Dilshad, R., & Anwar, M. (2022). Comprehensive Phytochemical Profiling, Biological Activities, and Molecular Docking Studies of *Pleurospermum candollei*: An Insight into Potential for Natural Products Development. *Molecules*, 27(13), 4113. https://doi.org/10.3390/molecules27134113
- Airy Shaw, H. K. (1972). The Euphorbiaceae of Siam. *Kew Bulletin*, 26, 191–363. https://doi.org/10.2307/4117717
- Akpuaka, A., Ekwenchi, M. M., Dashak, D. A., & Ahmed, D. (2013). Biological Activities of Characterized Isolates of n-Hexane Extract of *Azadirachta indica* A.Juss (Neem) Leaves. *New York Science Journal*, 6(6), 119–124. http://www.sciencepub.net/newyork
- Alabi, K. A., Lajide, L. & Owolabi, B. J. (2018). Biological activity of oleic acid and its primary amide: Experimental and computational studies. *Journal of Chemical Society of Nigeria*, 43(2), 9–18.
- Alamre, S. F., & Lmtair Algaraawi, N. (2020). Phytochemical profile and anti-fungal activity of stems and leaves methanol extract from the *Juncus maritimus* Linn. Juncaceae family against some dermatophytes fungi. *AIP Conference Proceedings*, 2290(1), 20034. http://dx.doi.org/10.1063/5.0027554
- Ali, M. S., Al Mamun, M. A., Abu Sayeed, M., Rahman, M. S., & Rashid, M. A. (2014). Sedative activity of methanolic extract of *Glochidion multiloculare* (Rottler ex Willd) Voigt leaves. *Pakistan Journal of Biological Sciences: PJBS*, 17(4), 555–559. https://doi.org/10.3923/pjbs.2014.555.559
- Alison, M. H., & James, S. (2010). 1.10 Type I Modular PKS, Editor(s): Hung-Wen (Ben) Liu, Lew Mander, Comprehensive Natural Products II, *Elsevier*, 385–452. https://doi.org/10.1016/B978-008045382-8.00015-0
- Al-Toubi, A. S. S., Al-Sadi, A. M., Al-Mahmooli, I. H., Al-Harrasi, M. M. A., Al-Sabahi, J. N., & Velazhahan, R. (2022). Volatile organic compounds emitted by mycoparasitic fungi *Hypomyces perniciosus* and *Cladobotryum mycophilum* suppress the growth of

- *Agaricus bisporus. Czech Mycology*, 74(2), 141–152. https://doi.org/10.33585/cmy.74203
- Ameya, G., Manilal, A., Sabu, K. R., & Aragie, S. (2022). Bioassay-Guided Phytochemical Analyses and Antimicrobial Potentials of the Leaf Extract of *Clematis hirsuta* Perr. and Guill. Against Some Pathogenic Bacteria and Fungi. *Infection and Drug Resistance*, 15, 6577–6588. https://doi.org/10.2147/IDR.S389699
- Anantachoke, N., Kitphati, W., Mangmool, S., & Bunyapraphatsara, N. (2015).
 Polyphenolic Compounds and Antioxidant Activities of the Leaves of *Glochidion hypoleucum*. Natural Product Communications, 10, 479–82.
 10.1177/1934578X1501000325.
- Are, P. C., Adidala, R. R., & Puchchakayala, G. (2011). Hypoglycemic and Antidiabetic Activity of *Glochidion velutinum* on Streptozotocin-Nicotinamide Induced Type 2 Diabetic Rats. *European Journal of Biological Sciences*, *3*(4), 126–130.
- Arulkumar, A., Rosemary, T., Paramasivam, S., & Rajendran, R. B. (2018). Phytochemical composition, in vitro antioxidant, antibacterial potential and GC-MS analysis of red seaweeds (*Gracilaria corticata* and *Gracilaria edulis*) from Palk Bay, India. *Biocatalysis and Agricultural Biotechnology*, 15, 63–71. https://doi.org/10.1016/j.bcab.2018.05.008
- Asraoui, F., Kounnoun, A., Cadi, H. El, Cacciola, F., Majdoub, Y. O. El, Alibrando, F., Mandolfino, F., Dugo, P., Mondello, L., & Louajri, A. (2021). Phytochemical investigation and antioxidant activity of *Globularia alypum* L. *Molecules*, 26(3), 759. https://doi.org/10.3390/molecules26030759
- Ayodele, A. E., & Olowokudejo, J. D. (2006). The family Polygonaceae in West Africa: Taxonomic significance of leaf epidermal characters. *South African Journal of Botany*, 72(3), 442–459. http://dx.doi.org/10.1016/j.sajb.2005.12.009
- Azam, A. T. M. Z., Abdullah, A. H., Mohammad, G. U., Mohammad, M. M., & Choudhury, M. H. (2012). Antimicrobial, Antioxidant and Cytotoxic Activities of *Glochidion multiloculare* (Roxb. Ex Willd.) Mull. Arg. (Euphorbiaceae). *Dhaka University Journal of Pharmaceutical Sciences*, 11(2), 117–120.

- Bae, I. K., Kim, K., Choi, S. D., Chang, K. S., Lee, H. S., & Lee, S. E. (2017). Mosquito larvicidal activities of naturally occurring compounds derived from *Piper* species. *Applied Biological Chemistry*, 60, 113–117. https://doi.org/10.1007/s13765-017-0256-1
- Bajpai, O, Kumar, A., Srivastava, A. K., Kuhwaha, A. K., Pandey, J., & Chaudhary, L. B. (2015). Tree species of Himalayan Terai region of Uttar Pradesh, India: a checklist. Checklist the Journal of Biodiversity data, 11(4), 1718.
- Baker, A. J. M. (1989). Terrestrial Higher Plants Which Hyperaccumulate Metallic Elements, A Review of Their Distribution, E. https://www.researchgate.net/publication/247713966
- Balakrishnan, N. P., & Chakrabarty T. (2007). The Family Euphorbiaceae in India: A synopsis of its Profile, Taxonomy and Bibliography, M/s Bishen Singh Mahendra Pal Singh, Dehradun, India.
- Balakrishnan, N. P. (1971). *Flora of Jowai*. Volume 2. Botanical Survey of India, Howrah, Calcutta, India.
- Balakrishnan, N. P., Chakrabarty, T., Sanjappa, M., Lakshminarsimhan, P., & Singh, P. (eds.) (2012). Flora of India. Volume 23. Loranthaceae-Daphniphyllaceae Botanical Survey of India, Kolkata, India.
- Balasubramani, G., Ramkumar, R., Krishnaveni, N., Sowmiya, R., Deepak, P., Arul, D., & Perumal, P. (2015). GC-MS analysis of bioactive components and synthesis of gold nanoparticle using *Chloroxylon swietenia* DC leaf extract and its larvicidal activity. *Journal of Photochemistry and Photobiology. B, Biology*, *148*, 1–8. https://doi.org/10.1016/j.jphotobiol.2015.03.016
- Baliyan, S., Mukherjee, R., Priyadarshini, A., Vibhuti, A., Gupta, A., Pandey, R. P., & Chang, C. M. (2022). Determination of Antioxidants by DPPH Radical Scavenging Activity and Quantitative Phytochemical Analysis of *Ficus religiosa*. *Molecules*, 27(4), 1326. https://doi.org/10.3390/molecules27041326
- Barthlott, W., Neinhuis, C., Cutler, D., Ditsch, F., Meusel, I., Theisen, I., & Wilhelmi, H. (1998). Classification and terminology of plant epicuticular wax. *Botanical Journal of the Linnaean Society*, *126*(3), 237–260.

- Bartzatt, R. (2013). Lomustine analogous drug structures for intervention of brain and spinal cord tumors: the benefit of in silico substructure search and analysis. *Chemotherapy Research and Practice*, 2013, 360624. https://doi.org/10.1155/2013/360624
- Behera, B., & Bhattacharya, S. (2016). The importance of assessing heavy metals in medicinal herbs: A quantitative study. *Tang [Humanitas Medicine]*. *6*(1), 3.1–3.4. https://doi.org/10.5667/tang.2015.0029
- Benny, S., Krishnakumar, K., & Sandhya, S. (2019). *Glochidion velutinum*: An overview. *Journal of Bio Innovation*, 8(4), 419–430.
- Bentham, G., & Hooker, J. D. (1862–1883). *Genera Plantarum*. 3 vols. London: various publishers.
- Beulah, G. G., Soris, P. T., & Mohan, V. R. (2018). GC-MS Determination of Bioactive Compounds of *Dendrophthoe falcata* (L.F) Ettingsh: An Epiphytic Plant. *International Journal of Health Sciences and Research*, 8(11), 261–269.
- Bhardwaj, M., Sali, V. K., Mani, S., & Vasanthi, H. R. (2020). Neophytadiene from *Turbinaria ornata* Suppresses LPS-Induced Inflammatory Response in RAW 264.7 Macrophages and Sprague Dawley Rats. *Inflammation*, 43(3), 937–950. https://doi.org/10.1007/s10753-020-01179-z
- Bildziukevich, U., Rárová, L., Šaman, D., & Wimmer, Z. (2018). Picolyl amides of betulinic acid as antitumor agents causing tumor cell apoptosis. *European Journal of Medicinal Chemistry*, *145*, 41–50. https://doi.org/10.1016/j.ejmech.2017.12.096
- Bora, P., & Kumar, Y. (2003). Floristic diversity of Assam Study of Pabitora Wildlife Sanctuary. Daya Publishing House, New Delhi.
- Borthakur, S. K., Bawri, A., Baro, D., & Boro, A. (2018). *Flora of BTAD*. 1st edition. Volume 3. EBH Publishers, India.
- Boulus, S. T., & Beakbane, A. B. (1971). A chemical method for separating leaf epidermis from mesophyll tissue. *U.A.R. Journal Botany*, *14*, 317–322.
- Brahma, P., & Baruah, S. (2023). Extended distribution of an endemic variety *Glochidion zeylanicum* var. *paucicarpum* Chakrab. & N.P. Balakr. (Phyllanthaceae) from Assam, India. *Vegetos*, *37*, 1484–1488. https://doi.org/10.1007/s42535-023-00650-0

- Brintha, S. J., Rajesh, S., Renuka, R. R., Vp, S. K., & Gnanam, R. (2017). Phytochemical analysis and bioactivity prediction of compounds in methanolic extracts of *Curculigo orchioides* Gaertn. *Journal of Pharmacognosy and Phytochemistry*, 6, 192–197.
- Casuga, F. P., Castillo, A. L., & Corpuz, M. J. A. T. (2016). GC–MS analysis of bioactive compounds present in different extracts of an endemic plant *Broussonetia luzonica* (Blanco) (Moraceae) leaves. *Asian Pacific Journal of Tropical Biomedicine*, 6(11), 957–961. https://doi.org/10.1016/j.apjtb.2016.08.015
- Cavazos, P., Gonzalez, D., Lanorio, J., & Ynalvez, R. (2021). Secondary metabolites, antibacterial and antioxidant properties of the leaf extracts of *Acacia rigidula* benth. and *Acacia berlandieri* benth. *SN Applied Sciences*, *3*(5), 522. https://doi.org/10.1007/s42452-021-04513-8
- Chakrabarty, T., & Balakrishnan, N. P. (2018). Indo-Burmese Phyllanthaceae: A Taxonomic Revision, M/s Bishen Singh Mahendra Pal Singh, Dehradun, India.
- Chakrabarty, T., & Gangopadhyay, M. (1995). The genus *Glochidion* (Euphorbiaceae) in Indian Subcontinent. *Journal of Economic and Taxonomic Botany*, 19(1), 173–234.
- Chanda, R., Mohanty, J., Bhuyan, N. R., Kar, P. K., & Nath, L. K. (2007). Medicinal plants used against gastrointestinal tract disorders by the traditional healers of Sikkim Himalayas. *Indian Journal of Traditional Knowledge*, 6(4), 606–610.
- Čižmáriková, R., Habala, L., Valentová, J., & Markuliak, M. (2019). Survey of Pharmacological Activity and Pharmacokinetics of Selected β-Adrenergic Blockers in Regard to Their Stereochemistry. *Applied Sciences*, 9(4), 625. https://doi.org/10.3390/app9040625
- Clark, S. H. (1986). Preservation of Herbarium Specimens: an archive conservator's approach. *Taxon*, *35*(4), 675–682. https://doi.org/10.2307/1221610
- Dahiru, M., Kolawole, S., & Tukur, A. (2022). Antibacterial and Phytochemicals Status of Detarium microcarpum (Guill and Perr) Stem Bark. Natural Resources for Human Health, 2(4), 1–6. https://doi.org/10.53365/nrfhh/145597
- Daimary, R. (2011). Studies in Floristic diversity of Kokrajhar district of Assam with special reference to Chakrasila Wildlife Sanctuary. Ph.D. Thesis, Department of Botany, Gauhati University, Assam, India.

- da Silva, N., Oliveira, M., Filho, H., Pinheiro, L., Kolb, R., & Bruno, O. M. (2017). Automatic Leaf Epidermis Assessment Using Fourier Descriptors in Texture Images. *Bio-Protocol*, 7(23), e2630. https://doi.org/10.21769/bioprotoc.2630
- Das, A. K., Stalin, N., Muthumperumal, C., & Swamy, P. S. (2013). Wild plants used by Muthuvan and Kattunaikkan tribal communities of Palakkayam settlement in Nilambur of Malappuram district, Kerala. *Medicinal Plants*, *5*(2), 82–89. https://doi.org/10.5958/j.0975-6892.5.2.013
- Das, M., & Malipeddi, H. (2014). Phytochemical screening, GC-MS analysis and biological activity of *Ipomoea eriocarpa* leaf extracts. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(4), 592–594.
- Deb, J., Saha, S., & Deb, N. K. (2019). Review on *Glochidion velutinum* Wight (Euphorbiaceae): A medicinal plant. *International Journal of Herbal Medicine*, 7(2), 1–3.
- de Carvalho, P. A. V., Campelo Lopes, I., Silva, E. H. C., Bruzaca, E. E. S., Alves, H. J., Lima, M. I. S., & Tanaka, A. A. (2019). Electrochemical behaviour of anticancer drug lomustine and in situ evaluation of its interaction with DNA. *Journal of Pharmaceutical and Biomedical Analysis*, *176*, 112786. https://doi.org/10.1016/j.jpba.2019.112786
- Deepak, P., Sowmiya, R., Balasubramani, G., & Perumal, P. (2017). Phytochemical profiling of *Turbinaria ornata* and its antioxidant and anti-proliferative effects. *Journal of Taibah University Medical Sciences*, *12*(4), 329–337. https://doi.org/10.1016/j.jtumed.2017.02.002
- Deryabin, D. G., & Tolmacheva, A. A. (2015). Antibacterial and Anti-Quorum Sensing Molecular Composition Derived from Quercus cortex (Oak bark) Extract. *Molecules (Basel, Switzerland)*, 20(9), 17093–17108. https://doi.org/10.3390/molecules200917093
- Duarte-Silva, A. G., Carvalho-Silva, M., & Câmara, P. E. A. S. (2013). Morphology and development of leaf papillae in the Pilotrichaceae. *Acta Botanica Brasilica*, 27(4), 737–742. https://doi.org/10.1590/S0102-33062013000400013
- Duangjan, C., Rangsinth, P., Gu, X., Zhang, S., Wink, M., & Tencomnao, T. (2019). Glochidion zeylanicum leaf extracts exhibit lifespan extending and oxidative stress

- resistance properties in Caenorhabditis elegans via DAF-16/FoxO and SKN-1/Nrf-2 signaling pathways. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*, 64, 153061. https://doi.org/10.1016/j.phymed.2019.153061
- Dulara, B. K., Godara, P., & Barwer, N. (2019). In-vivo and In-vitro phytochemical GC-MS analysis of volatile constituents of *Andrographis paniculata* (Burm.f.) Nees. *The Pharma Innovation*, 8(5), 255–261.
- Eberly, D. (2008). The Area of Intersecting Ellipses. United States of America. Geometric Tools, Redmond WA 98052.
- eFloras. (2008). Missouri Botanical Garden, St. Louis, MO & Harvard University Herbaria, Cambridge, MA. Published on the Internet http://www.efloras.org. (Accessed on 23rd July 2021).
- Elaiyaraja, A., & Chandramohan, G. (2018). Comparative phytochemical profile of *Crinum defixum* Ker-Gawler leaves using GC-MS. *Journal of Drug Delivery and Therapeutics*, 8(4), 365–380. https://doi.org/10.22270/jddt.v8i4.1758
- Elanchezhian, R., Kumar, R. S., Beena, S., & Suryanarayana, M. A. (2007). Ethnobotany of Shompens a primitive tribe of Great Nicobar Island. *Indian Journal of Traditional Knowledge*, 6(2), 342–345.
- Ellis, B., Daly, D., Hickey, L., Johnson, K., Mitchell, J., Wilf, P., & Wing, S. (2009). Manual of Leaf Architecture. The New York Botanical Garden Press, Comstock Publishing Associates a division of Cornell University Press, Ithaca, New York.
- Endress, P. K., Bass, P., & Gregory, M. (2000). Systematic Plant morphology and anatomy-50 years of progress. *Taxon*, 49(3), 401–434.
- Ensikat, H. J., Ditsche-Kuru, P., & Barthlott, W. (2011). Microscopy: Science, Technology, Applications and Education (pp.248-255) Scanning electron microscopy of plant surfaces: simple but sophisticated methods for preparation and examination. Formatex Research Center, Badajoz, Spain.
- Ferguson, N. M. (1956). A textbook of pharmacognosy. New York: Max Millam Company.
- Filartiga, A. L., Klimeš, A., Altman, J., Nobis, M. P., Crivellaro, A., Schweingruber, F., & Doležal, J. (2022). Comparative anatomy of leaf petioles in temperate trees and shrubs:

- the role of plant size, environment and phylogeny. *Annals of Botany*, *129*(5), 567–582. https://doi.org/10.1093/aob/mcac014
- Fontana, A., Spolaore, B., & Polverino de Laureto, P. (2013). The biological activities of protein/oleic acid complexes reside in the fatty acid. *Biochimica et Biophysica Acta*, 1834(6), 1125–1143. https://doi.org/10.1016/j.bbapap.2013.02.041
- Forster, J. R., & Forster, G. (1776). *Characteres Generum Plantarum*. 2nd Edition. Londini, Prostant apud B. White, T. Cadell, & P. Elmsly, 1776.
- Fu, D. J., Zhang, Y. F., Chang, A. Q., & Li, J. (2020). β-Lactams as promising anticancer agents: Molecular hybrids, structure activity relationships and potential targets. *European Journal of Medicinal Chemistry*, 201, 112510. https://doi.org/10.1016/j.ejmech.2020.112510
- Gan, J., Feng, Y., He, Z., Li, X., & Zhang, H. (2017). Correlations between Antioxidant Activity and Alkaloids and Phenols of Maca (*Lepidium meyenii*). *Journal of Food Quality*, 2017(3), 1–10. https://doi.org/10.1155/2017/3185945
- Ganesh, M., & Mohankumar, M. (2017). Extraction and identification of bioactive components in *Sida cordata* (Burm.f.) using gas chromatography-mass spectrometry. *Journal of Food Science and Technology*, *54*(10), 3082–3091. https://doi.org/10.1007/s13197-017-2744-z
- García-Gutiérrez, E., Ortega-Escalona, F., & Angeles, G. (2020). A novel, rapid technique for clearing leaf tissues. *Applications in Plant Sciences*, 8(9), e11391. https://doi.org/10.1002/aps3.11391
- GBIF.org (2021). GBIF- Global Biodiversity Information Facility. Published on the Internet https://www.gbif.org (Accessed on 24th May 2021).
- Gillis, E. P., Eastman, K. J., Hill, M. D., Donnelly, D. J., & Meanwell, N. A. (2015). Applications of Fluorine in Medicinal Chemistry. *Journal of Medicinal Chemistry*, 58(21), 8315–8359. https://doi.org/10.1021/acs.jmedchem.5b00258
- Gliszczyńska, A., Dancewicz, K., Gabryś, B., Świtalska, M., Wietrzyk, J., & Maciejewska, G. (2021). Synthesis of novel phytol-derived γ-butyrolactones and evaluation of their biological activity. *Scientific Reports*, 11(1), 4262. https://doi.org/10.1038/s41598-021-83736-6

- Gogoi, D., Bora, G., Borgohain, R., & Handique, J. G. (2018). Antioxidant Capacity and GC-MS Analysis of Hexane, Ethyl acetate and Methanol extracts of *Ficus bhotanica* A Potential Folklore Medicinal Plant. *International Journal of Pharmacognosy and Phytochemical Research*, 10(5), 201–212. https://doi.org/10.25258/phyto.10.5.5
- Gollo, A. L., Tanobe, V. O. A., de Melo Pereira, G. V., Marin, O., Bonatto, S. J. R., Silva, S., de Barros, I. R., & Soccol, C. R. (2020). Phytochemical analysis and biological activities of in vitro cultured *Nidularium procerum*, a bromeliad vulnerable to extinction. *Scientific Reports*, 10(1), 7008. https://doi.org/10.1038/s41598-020-64026-2
- Gooda Sahib, N., Saari, N., Ismail, A., Khatib, A., Mahomoodally, F., & Abdul Hamid, A. (2012). Plants' metabolites as potential antiobesity agents. *The Scientific World Journal*, 2012, 436039. https://doi.org/10.1100/2012/436039
- Gunavathy, S. K., & Sherine, H. B. (2019). Determination of heavy metals and phytochemical analysis of some selected medicinal plants. *International Journal of Scientific Research in Biological Sciences*, 6(3), 87–96. https://doi.org/10.26438/ijsrbs/v6i3.8796
- Hadi, M. Y., Mohammed, G. J., & Hameed, I. H. (2016). Analysis of bioactive chemical compounds of *Nigella sativa* using gas chromatography-mass spectrometry. *Journal of Pharmacognosy and Phytotherapy*, 8(2), 8–24. http://dx.doi.org/10.5897/JPP2015.0364
- Hana Saeid, Hadeel Al-sayed, & Marwa Bader. (2023). A Review on Biological and Medicinal Significance of Furan. *AlQalam Journal of Medical and Applied Sciences*, 44–58.
- Harborne, J. B. (1973). *Phytochemical Methods, A guide to modern techniques of plant analysis*. Chapman & Hall Ltd., London.
- Harborne, J. B. (1983). *Phytochemical Methods*, A guide to modern techniques of plant analysis. 3rd Edition. Chapman & Hall Ltd., London.
- Hasan, R., Abdul Awoal, M., Shafiqul Islam, M., Raquibul Hasan, M., Islam, T., Iqbal Hussain Shameem, M., & Sohel Rana, M. (2016). Evaluation of cytotoxic potentiality of different extracts of *Glochidion velutinum* Wight's leaves through brine shrimp

- lethality bioassay. Article in World Journal of Pharmaceutical Research, 5(1), 172–179.
- Hata, K., Hori, K., Ogasawara, H., & Takahashi, S. (2003). Anti-leukemia activities of Lup-28-al-20(29)-en-3-one, a lupane triterpene. *Toxicology letters*, *143*(1), 1–7. https://doi.org/10.1016/s0378-4274(03)00092-4
- Hethelyi, E., Tetenyi, P., Dabi, E., & Danos, B. (1987). The Role of Mass Spectrometry in Medicinal Plant Research. *Biomedical and Environmental Mass Spectrometry*, *14*(11), 627–632. https://doi.org/10.1002/bms.1200141110
- Hoffmann, P., Kathriarachchi, H., & Wurdack, K. J. (2006). A phylogenetic classification of Phyllanthaceae (Malpighiales; Euphorbiaceae *sensu lato*). *Kew Bulletin*, 61(1), 37–53.
- Hooker, J. D. (1890). *The Flora of British India*. Volume 5. L Reeve & Co., 5, Henrietta Street, Covent Garden, London.
- Huang, Q., Liu, X., Zhao, G., Hu, T., & Wang, Y. (2018). Potential and challenges of tannins as an alternative to in-feed antibiotics for farm animal production. *Animal Nutrition*, 4(2), 137–150. https://doi.org/10.1016/j.aninu.2017.09.004
- Huang, S., Chen, X., Yan, R., Huang, M., & Chen, D. (2022). Isolation, Identification and Antibacterial Mechanism of the Main Antibacterial Component from Pickled and Dried Mustard (*Brassica juncea* Coss. var. *foliosa* Bailey). *Molecules (Basel, Switzerland)*, 27(8), 2418. https://doi.org/10.3390/molecules27082418
- Hutchinson, J. (1973). *The Families of Flowering Plants*. 3rd Edition. The Clarendon Press, Oxford.
- Igwe, O. U., & Okwunodulu, F. U. (2014). Investigation of bioactive phytochemical compounds from the chloroform extract of the leaves of *Phyllanthus amarus* by GC-MS technique. *International Journal of Chemistry and Pharmaceutical Sciences*, 2(1), 554–560.
- Indian Virtual Herbarium, Botanical Survey of India, Ministry of Environment, Forest and Climate Change (Government of India). Published on the Internet https://bsi.gov.in/page/en/Virtual-Herbarium (Accessed on 25th Feb 2021).

- IPNI (2021). International Plant Name Index. Published on the Internet http://www.ipni.org. The Royal Botanic Gardens, Kew, Harvard University Herbaria & Libraries and Australian National Herbarium. (Accessed on 25th September 2021).
- Islam, M. T., Ali, E. S., Uddin, S. J., Shaw, S., Islam, M. A., Ahmed, M. I., Chandra Shill, M., Karmakar, U. K., Yarla, N. S., Khan, I. N., Billah, M. M., Pieczynska, M. D., Zengin, G., Malainer, C., Nicoletti, F., Gulei, D., Berindan-Neagoe, I., Apostolov, A., Banach, M., Yeung, A. W. K., ... Atanasov, A. G. (2018). Phytol: A review of biomedical activities. Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association, 121, 82–94. https://doi.org/10.1016/j.fct.2018.08.032
- Islary, A., Sarmah, J., & Basumatary, S. (2017a). Nutritional Properties, Phytochemicals and In Vitro Antioxidant Assessment of Two Wild Edible Fruits from Assam of North-East India. *Journal of Pharmacy and Nutrition Sciences*, 7, 55–63.
- Islary, A., Sarmah, J., & Basumatary, S. (2017b). Nutritional value, phytochemicals and antioxidant properties of two wild edible fruits (*Eugenia operculata* Roxb. and *Antidesma bunius* L.) from Assam, North-East India. *Mediterranean Journal of Nutrition and Metabolism*, 10(1), 29–40. https://doi.org/10.3233/MNM-16119
- Jain, S. K., & Rao, R. R. (1977). *A handbook of Field and Herbarium Technique*. Today & Tomorrow Publication, New Delhi.
- Jalpa, K., & Vijaykumar, R. (2023). Extraction and Identification of Phytochemicals in N-hexane and Methanol Extracts of *Cocos nucifera* L. Leaves. *Advances in Zoology and Botany*, 11(3), 213–219. http://dx.doi.org/10.13189/azb.2023.110307
- Jassbi, A. R., Mirzaei, Y., Firuzi, O., Chandran, J. N., & Schneider, B. (2016). Bioassay guided purification of cytotoxic natural products from a red alga *Dichotomaria obtusata*. *Revista Brasileira de Farmacognosia*, 26(6), 705–709. http://dx.doi.org/10.1016/j.bjp.2016.06.008
- Jawarkar, S. V., & Kane, S. R. (2017). Phytochemical and Anthelmintic Investigation of Leaves of *Glochidion ellipticum* Linn. *European Journal of Experimental Biology*, 7, 2–9.

- Jawarkar, S. V., & Mohite, S. K. (2016). In vitro-anticancer activity, activity of ethanol extract of *Glochidion ellipticum*. *International Journal of Recent Scientific Research*, 7(8), 13222–13225.
- Jawarkar, S. V. (2015). *Phytochemical investigation and Pharmacological screening on Glochidion elipticum family Euphorbiaceae*. Ph.D Thesis. Department of Pharmaceutical Sciences. Shri Jagdish Prasad Jhabarmal Tibrewala University, Vidyanagari, Jhunjhunu, Rajasthan.
- Junlatat, J., & Sripanidkulchai, B. (2016). Anti-Inflammatory And Anti-Oxidant Activities of *Glochidion daltonii* Branch Extract. *Jurnal Farmasi Dan Ilmu Kefarmasian Indonesia*, 3(2), 39. http://dx.doi.org/10.20473/jfiki.v3i22016.39-43
- Kabir, S., & Chowdhury, A. M. S., Rashid, M. A., & Hasan, C. M. (2013). Antimicrobial and Cytotoxic Activities of the Extracts of *Glochidion multiloculare*. *Journal of Science Foundation*, 10. 10.3329/jsf.v10i1.16273.
- Kabir, S., Zahan, R., Mohammad, A., Chowdhury, S., Haque, M. R., & Rashid, M. A. (2015). Antitumor, Analgesic and Anti-inflammatory Activities of Glochidion multiloculare (Rottler ex Willd) Voigt. Bangladesh Pharmaceutical Journal, 18(2), 14–28.
- Kabir, S. & Haque, M. R., Chowdhury, A. M. S., Rashid, M. A., & Hasan, C. M. (2021).
 Triterpenoids Isolated from Stem Bark of *Glochidion lanceolarium* (Roxb.), Voigt.
 Journal of Science Foundation, 18(1), 13–18. 10.3329/jsf.v18i1.51278.
- Kandasamy, S., Sahu, S. K., & Kandasamy, K. (2012). In Silico Studies on Fungal Metabolite against Skin Cancer Protein (4,5-Diarylisoxazole HSP90 Chaperone). *ISRN Dermatology*, 2012, 626214. https://doi.org/10.5402/2012/626214
- Kanjilal, U. N., Kanjilal, P. C., Dey, R. N., & Das, A. (1940). *Flora of Assam*. Volume 4. Prabasi Press, Upper Circular Road, Calcutta.
- Karthikeyan, S. C., Velmurugan, S., Donio, M. B., Michaelbabu, M., & Citarasu, T. (2014). Studies on the antimicrobial potential and structural characterization of fatty acids extracted from Sydney rock oyster *Saccostrea glomerata*. *Annals of*

- Kathriarachchi, H., Samuel, R., Hoffmann, P., Mlinarec, J., Wurdack, K. J., Ralimanana, H., Stuessy, T. F., & Chase, M. W. (2006). Phylogenetics of tribe Phyllantheae (Phyllanthaceae; Euphorbiaceae *sensu lato*) based on nrITS and plastid matK DNA sequence data. *American Journal of Botany*, 93(4), 637–655. https://doi.org/10.3732/ajb.93.4.637
- Kato, M., Takimura, A., Kawakita, A. (2003) An obligate pollination mutualism and reciprocal diversification in the tree genus *Glochidion* (Euphorbiaceae). *Proceedings of the National Academy of Sciences, USA, 100*(9), 5264–5267.
- Katz, D. H., Marcelletti, J. F., Khalil, M. H., Pope, L. E., & Katz, L. R. (1991). Antiviral activity of 1-docosanol, an inhibitor of lipid-enveloped viruses including herpes simplex. *Proceedings of the National Academy of Sciences of the United States of America*, 88(23), 10825–10829. https://doi.org/10.1073/pnas.88.23.10825
- Kedare, S. B., & Singh, R. P. (2011). Genesis and development of DPPH method of antioxidant assay. *Journal of Food Science and Technology*, 48(4), 412–422. https://doi.org/10.1007/s13197-011-0251-1
- Khajuria, V., Gupta, S., Sharma, N., Kumar, A., Lone, N. A., Khullar, M., Dutt, P., Sharma,
 P. R., Bhagat, A., & Ahmed, Z. (2017). Anti-inflammatory potential of hentriacontane in LPS stimulated RAW 264.7 cells and mice model. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 92, 175–186. https://doi.org/10.1016/j.biopha.2017.05.063
- Khalid, A., Algarni, A. S., Homeida, H. E., Sultana, S., Javed, S. A., Rehman, Z. U., Abdalla, H., Alhazmi, H. A., Albratty, M., & Abdalla, A. N. (2022).
 Phytochemical, Cytotoxic, and Antimicrobial Evaluation of *Tribulus terrestris* L., *Typha domingensis* Pers., and *Ricinus communis* L.: Scientific Evidences for Folkloric Uses. *Evidence-Based Complementary and Alternative Medicine:* eCAM, 2022, 6519712. https://doi.org/10.1155/2022/6519712
- Khan, M. S., Yusufzai, S. K., Ying, L. Y., & Zulnashriq, W. (2018). GC-MS Based Chemical Profiling and Evaluation of Antioxidant Potential of Leaves and Stems of *Althernanthera sessilis* Red from Sabah, Malaysia. *International Journal of*

- Pharmacy and Pharmaceutical Sciences, 10(7), 4–9. http://dx.doi.org/10.22159/ijpps.2018v10i7.25204
- Khodarahmi, G., Asadi, P., Hassanzadeh, F., & Khodarahmi, E. (2015). Benzofuran as a promising scaffold for the synthesis of antimicrobial and antibreast cancer agents: A review. *Journal of Research in Medical Sciences: the official journal of Isfahan University of Medical Sciences*, 20(11), 1094–1104. https://doi.org/10.4103/1735-1995.172835
- Kibungu, W. C., Fri, J., Clarke, A. M., Otigbu, A., & Akum Njom, H. (2021). Seasonal Variation in Antimicrobial Activity of Crude Extracts of *Psammaplysilla* sp. 1 from Phillips Reef, South Africa. *International Journal of Microbiology*, 2021, 7568493. https://doi.org/10.1155/2021/7568493
- Kiem, P. V., Thu, V. K., Yen, P. H., Nhiem, N. X., Tung, N. H., Cuong, N. X., Minh, C. V., Huong, H. T., Hyun, J. H., Kang, H. K., & Kim, Y. H. (2009). New Triterpenoid Saponins from *Glochidion eriocarpum* and Their Cytotoxic Activity. *Chemical and Pharmaceutical Bulletin*, 57(1), 102–105.
- Klein, E., DeBonis, S., Thiede, B., Skoufias, D. A., Kozielski, F., & Lebeau, L. (2007).

 New chemical tools for investigating human mitotic kinesin Eg5. *Bioorganic & Medicinal Chemistry*, *15*(19), 6474–6488.

 https://doi.org/10.1016/j.bmc.2007.06.016
- Koch, K., & Barthlott, W. (2006). Plant Epicuticular Waxes: Chemistry, Form, Self-Assembly and Function. *Natural Product Communications*, *1*(11), 1067–1072.
- Kong, M. J., & Hong, S. P. (2019). Leaf micromorphology of the *Persicaria* sect. *Cephalophilon* (Polygonaceae) and its systematic re-evaluation. *Phytotaxa*, 391(3), 167–184. https://doi.org/10.11646/phytotaxa.391.3.1
- Kong, H. S., Musa, K. H., Mohd-Kasim, Z., & Abdullah Sani, N. (2019). Qualitative and quantitative phytochemical analysis and antioxidant properties of leaves and stems of *Clinacanthus nutans* (Burm. f.) Lindau from two herbal farms of Negeri Sembilan, Malaysia. *ASM Science Journal*, 12, 1–13. http://dx.doi.org/10.32802/asmscj.2019.87

- Kulikowska, E., Kierdaszuk, B., & Shugar, D. (2004). Xanthine, xanthosine and its nucleotides: solution structures of neutral and ionic forms, and relevance to substrate properties in various enzyme systems and metabolic pathways. *Acta Biochimica Polonica*, *51*(2), 493–531. http://dx.doi.org/10.18388/abp.2004_3587
- Kumar, J. U. S., Chaitanya, M. J. K., Semotiuk, A. J., & Krishna, V. (2019). Indigenous knowledge on medicinal plants used by ethnic communities of South India. *Ethnobotany Research and Applications*, 18, 11–12.
- Kumar, P., & Sati, S. C. (2021). Chemical composition, antioxidant and antimicrobial activities of Himalayan *Fraxinus micrantha* Lingelsh leaf extract. *Natural Product Research*, *35*(20), 3519–3523. https://doi.org/10.1080/14786419.2019.1710706
- Kumar, S., Koh, J., Kim, H., Gupta, M. K., & Dutta, P. K. (2012). A new chitosan-thymine conjugate: synthesis, characterization and biological activity. *International Journal of Biological Macromolecules*, *50*(3), 493–502. https://doi.org/10.1016/j.ijbiomac.2012.01.015
- Kumar, V., Singh, S., Srivastava, B., Kumari Patial, P., Kondalkar, S. A., & Bharthi, V. (2019). Volatile and semi-volatile compounds of *Tephrosia purpurea* and its medicinal activities: Experimental and computational studies. *Biocatalysis and Agricultural Biotechnology*, 20, 101222. https://doi.org/10.1016/j.bcab.2019.101222.
- Kushwaha, P., Yadav, S. S., Singh, V., & Dwivedi, L. K. (2019). GC-MS analysis of bioactive compounds in methanolic extract of *Ziziphus mauritiana* fruit. *International Journal of Pharmaceutical Sciences* & *Research*, 10(6), 2911–2916. http://dx.doi.org/10.13040/IJPSR.0975-8232.10(6).2911-16
- Kwofie, M. A., & Gupta, M. (2021). Phenanthrene: A versatile molecule; A review. *Plant Archives*, 21(1), 368–378. http://dx.doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.051
- Lalfakzuala, R., Lalramnghinglova, H. & Kayang, H. (2007). Ethnobotanical usage of plants in western Mizoram. *Indian Journal of Traditional Knowledge*, 6(3), 486–493.

- Lalrinkimi, & Lallianthanga, R. K. (2019). Documentation of tree species within Mizoram Science Centre, Berawtlang, Aizawl, India with notes on their ethnomedicinal values. *Science Vision*, *19*(3), 63–78.
- Lersten, N. R., & Curtis, J. D. (2001). Idioblasts and other unusual internal foliar secretory structures in Scrophulariaceae. *Plant Systematics and Evolution*, 227(1), 63–73.
- Li, P. T., & Gilbert, M. G. (2008). *Glochidion. In*: Wu, Z.Y., Raven, P.H. & Hong, D.Y. (Eds.) *Flora of China*, vol. 11. Science Press & Missouri Botanical Garden Press, Beijing & St. Louis, pp. 193–202.
- Liang, P., Sang, H., & Sun, Z. (2006). Cloud point extraction and graphite furnace atomic absorption spectrometry determination of manganese (II) and iron (III) in water samples. *Journal of Colloid and Interface Science*, 304(2), 486–490. https://doi.org/10.1016/j.jcis.2006.09.006
- Liu, K., Zhang, X., Xie, L., Deng, M., Chen, H., Song, J., Long, J., Li, X., & Luo, J. (2021).
 Lupeol and its derivatives as anticancer and anti-inflammatory agents: Molecular mechanisms and therapeutic efficacy. *Pharmacological Research*, 164, 105373.
 https://doi.org/10.1016/j.phrs.2020.105373
- Liu, Y., Nielsen, M., Staerk, D., & Jäger, A. K. (2014). High-resolution bacterial growth inhibition profiling combined with HPLC-HRMS-SPE-NMR for identification of antibacterial constituents in Chinese plants used to treat snakebites. *Journal of Ethnopharmacology*, 155(2), 1276–1283. https://doi.org/10.1016/j.jep.2014.07.019
- Liu, Y., Yang, K., Jia, Y., Shi, J., Tong, Z., & Wang, Z. (2021). Thymine Sensitizes Gram-Negative Pathogens to Antibiotic Killing. *Frontiers in Microbiology*, *12*, 622798. https://doi.org/10.3389/fmicb.2021.622798
- Lognay, G., Marlier, M., Seck, D., & Haubruge, É. (2000). The occurrence of 2-hydroxy-6-methoxybenzoic acid methyl ester in *Securidaca longipedunculata* Fresen root bark. *Biotechnology*, *Agronomy and Society and Environment*, 4(2), 107–110.
- Lopez-Romero, J. C., González-Ríos, H., Borges, A., & Simões, M. (2015). Antibacterial Effects and Mode of Action of Selected Essential Oils Components against

- Escherichia coli and Staphylococcus aureus. Evidence-Based Complementary and Alternative Medicine: eCAM, 2015, 795435. https://doi.org/10.1155/2015/795435
- Machana S., Weerapreeyakul N., Barusrux S., Nonpunya A., Sripanidkulchai B., & Thitimetharoch T. (2011). Cytotoxic and apoptotic effects of six herbal plants against the human hepatocarcinoma (HepG2) cell line. *Chinese Medicine*, 6, 39.
- Mahajan, A., Singh, H., Singh, A., Agrawal, D. K., Arora, A., Chundawat, T. S. (2022). Trifluoromethylated Quinolone-Hydantoin Hybrids: Synthesis and Antibacterial Evaluation. *Sci*, *4*(*3*), 30. https://doi.org/10.3390/sci4030030
- Maher, T., Kabbashi, N. A., Mirghani, M. E. S., Alam, M. Z., Daddiouaissa, D., Abdulhafiz, F., Reduan, M. F. H., Omran, J. I., Abdul Razab, M. K. A., & Mohammed, A. (2021). Optimization of Ultrasound-Assisted Extraction of Bioactive Compounds from *Acacia Seyal* Gum Using Response Surface Methodology and Their Chemical Content Identification by Raman, FTIR, and GC-TOFMS. *Antioxidants*, 10, 1612. https://doi.org/10.3390/antiox10101612
- Maitra, S., De, A., Das, B., Roy, S. N., Chakraborty, R., Samanta, A., & Bhattacharya, S.
 (2019). Seasonal Variation of Phyto-Constituents of Tea Leaves Affects
 Antiproliferative Potential. *Journal of the American College of Nutrition*, 38(5),
 415–423. https://doi.org/10.1080/07315724.2018.1538829
- Mallikadevi, T., Paulsamy, S., Jamuna, S., & Karthika, K. (2012). Analysis for phytoceuticals and bioinformatics approach for the evaluation of therapeutic properties of whole plant methanolic extract of *Mukia maderaspatana*—A traditional medicinal plant in western districts of Tamil Nadu. India. *Asian Journal of Pharmaceutical and Clinical Research*, 5(4), 163–168.
- Mallikarjuna, B. (2012). Studies on in vitro morphogenesis, phytochemistry and bioactivity of Glochidion velutinum Wt., a medicinal plant. Ph.D Thesis. Department of Botany. Sri Venkateswara University, Tirupati-517 502 (A.P), India.
- Marchioni, I., Najar, B., Ruffoni, B., Copetta, A., Pistelli, L., & Pistelli, L. (2020).

 Bioactive Compounds and Aroma Profile of Some Lamiaceae Edible
 Flowers. *Plants* (*Basel*, *Switzerland*), 9(6), 691.

 https://doi.org/10.3390/plants9060691

- Martins, F., Santos, S., Ventura, C., Elvas-Leitão, R., Santos, L., Vitorino, S., Reis, M., Miranda, V., Correia, H. F., Aires-de-Sousa, J., Kovalishyn, V., Latino, D. A., Ramos, J., & Viveiros, M. (2014). Design, synthesis and biological evaluation of novel isoniazid derivatives with potent antitubercular activity. *European Journal of Medicinal Chemistry*, 81, 119–138. https://doi.org/10.1016/j.ejmech.2014.04.077
- Mazumder, K., Nabila, A., Aktar, A., & Farahnaky, A. (2020). Bioactive Variability and In Vitro and In Vivo Antioxidant Activity of Unprocessed and Processed Flour of Nine Cultivars of Australian *lupin* Species: A Comprehensive Substantiation. *Antioxidants* (*Basel*, *Switzerland*), 9(4), 282. https://doi.org/10.3390/antiox9040282
- Mc Ateer, C. H., Balasubramanian, M., & Murugan, R. (2008). Pyridines and their Benzo Derivatives: Applications. In Comprehensive Heterocyclic Chemistry III; Elsevier: Cambridge, MA, USA, pp. 309–336.
- Melville, R. (1976). The terminology of Leaf Architecture. *Taxon*, 25, 549–561.
- Mena, T. P., Sutrisno, S., & Marfu'ah, S. (2020). Antibacterial activity of free fatty acids, potassium soap, and fatty acids methylesters from VCO (virgin coconut oil). *IOP Conference Series: Materials Science and Engineering*, 833(1): 012023. http://dx.doi.org/10.1088/1757-899X/833/1/012023
- Metcalfe, C. R., & Chalk, L. (1950). *Anatomy of Dicotyledons Vol.II.* Clarendon Press, Oxford, England.
- Metcalfe, C. R., & Chalk, L. (1979). Anatomy of Dicotyledonous. 2nd Edition, Clarendon Press, Oxford, 456–473.
- Miao, Y. H., Hu, Y. H., Yang, J., Liu, T., Sun, J., & Wang, X. J. (2019). Natural source, bioactivity and synthesis of benzofuran derivatives. *RSC advances*, 9(47), 27510–27540. https://doi.org/10.1039/c9ra04917g
- Mishra, C., & Tripathi, I. P. (2015). Phytochemical Screening of Some Medicinal Plants of Chitrakoot Region Phytochemical Screening of Some Medicinal Plants of Chitrakoot Region. *Indian Journal of Applied Research*, *5*(12), 56–60. https://www.researchgate.net/publication/312384165

- Mohammad, S. M., Janaki, C. S., Rao, M. R. K., Prabhu, K., Deepa, K., Franklin, & Vijayalakshmi, N. (2022). The Gas Chromatography-Mass Spectroscopy Analysis of One Unani Drug, "Majoon Falasifa". *Journal of Research in Medical and Dental Science*, 10(9), 129–132.
- Mohanta, Y. K., Biswas, K., Jena, S. K., Hashem, A., Abd Allah, E. F., & Mohanta, T. K. (2020). Anti-biofilm and Antibacterial Activities of Silver Nanoparticles Synthesized by the Reducing Activity of Phytoconstituents Present in the Indian Medicinal Plants. Frontiers in Microbiology, 11. https://doi.org/10.3389/fmicb.2020.01143
- Morah, F. N. I., Emehige, E. P., & Mowang, M. M. (2017). Chemical composition and antimicrobial activity of *Nauclea latifolia* leaf essential oil. *International Journal of Chemical and Biochemical Sciences*, 11, 44–50.
- More, K., Tayade, S., Gawande, P., Manik, S. & Shelke, D. (2022). Antioxidant and antimicrobial potential of *Canavalia gladiata* (Jacq.) DC. leaves and seeds: GC-MS based metabolic profiling. *Indian Journal of Natural Products and Resources*, 13(2), 163–169. https://doi.org/10.56042/ijnpr.v13i2.47499
- Musialik, M., Kuzmicz, R., Pawlowski, T. S., & Litwinienko, G. (2009). Acidity of hydroxyl groups: An overlooked influence on antiradical properties of flavonoids. *Journal of Organic Chemistry*, 74(7), 2699–2709. https://doi.org/10.1021/jo802716v
- Nabi, M., Tabassum, N., & Ganai, B. A. (2022). Phytochemical screening and antibacterial activity of *Skimmia anquetilia* N.P. Taylor and Airy Shaw: A first study from Kashmir Himalaya. *Frontiers in Plant Science*, *13*, 937946. https://doi.org/10.3389/fpls.2022.937946
- Naine, S. J., Devi, C. S., Mohanasrinivasan, V., Doss, C. G., & Kumar, D. T. (2016). Binding and molecular dynamic studies of sesquiterpenes (2R-acetoxymethyl-1,3,3-trimethyl-4t-(3-methyl-2-buten-1-yl)-1t-cyclohexanol) derived from marine *Streptomyces* sp. VITJS8 as potential anticancer agent. *Applied Microbiology and Biotechnology*, 100(6), 2869–2882. https://doi.org/10.1007/s00253-015-7156-2
- Nandhini, R. S., Nithya, R. N., & Vidhya, K. (2021). GC-MS analysis of Phytochemical compounds in different extracts of *Curculigo orchiodes*. *Research Journal of*

- Pharmacy and Technology, 14, 4355–4360. http://dx.doi.org/10.52711/0974-360X.2021.00756
- Narzary, H., Islary, A., & Basumatary, S. (2016). Phytochemicals and antioxidant properties of eleven wild edible plants from Assam, India. *Mediterranean Journal of Nutrition and Metabolism*, 9(3), 191–201. https://doi.org/10.3233/MNM-16116
- Nasr, Z., El-shershaby, H., Sallam, K., Abed, N., Abd- El ghany, I., & sidkey, N. (2022).
 Evaluation of Antimicrobial Potential of Tetradecane Extracted from *Pediococcus acidilactici* DSM: 20284 CM Isolated from Curd Milk. *Egyptian Journal of Chemistry*, 65(3), 705-713. doi: 10.21608/ejchem.2021.92658.4385
- Ngobeni, B., Mashele, S. S., Malebo, N. J., van der Watt, E., & Manduna, I. T. (2020).

 Disruption of microbial cell morphology by *Buxus macowanii*. *BMC Complementary Medicine and Therapies*, 20(1), 266. https://doi.org/10.1186/s12906-020-03049-5
- Nhiem, N. X., Thu, V. K., Kiem, P. V., Minh, C. V., Tai, B. H., Quang, T. H., Cuong, N. X., Yen, P. H., Boo, H. J., Kang, J., Kang, H. K., & Kim, Y. H. (2012). Cytotoxic Oleane-type Triterpene Saponins from *Glochidion eriocarpum*. Archives of Pharmacal Research, 35(1), 19–26.
- Nistor, G., Trandafirescu, C., Prodea, A., Milan, A., Cristea, A., Ghiulai, R., Racoviceanu, R., Mioc, A., Mioc, M., Ivan, V., & Şoica, C. (2022). Semisynthetic Derivatives of Pentacyclic Triterpenes Bearing Heterocyclic Moieties with Therapeutic Potential. *Molecules* (*Basel*, *Switzerland*), 27(19), 6552. https://doi.org/10.3390/molecules27196552
- Nithyadevi, J., & Sivakumar, R. (2015). Phytochemical screening and GC-MS, FT-IR analysis of methanolic extract leaves of *Solanum torvum* Sw. *International Journal of Research Studies in Biosciences*, *3*(9), 61–66.
- Ogunjobi, K. M., Abdulwahab, S. O., Gakenou, O. F., Thompson, O. E., & Olorunfemi, O. (2020). Qualitative and quantitative evaluation of the phytochemical constituents of three wood species in Ogun state, Nigeria. *Tropical Plant Research*, 7(3), 627–633. https://doi.org/10.22271/tpr.2020.v7.i3.078

- Oliveira, L. C. C., Rodrigues, F. A. A., dos Santos Barbosa, C. R., dos Santos, J. F. S., Macêdo, N. S., de Sousa Silveira, Z., Coutinho, H. D. M., & da Cunha, F. A. B. (2022). Antibacterial Activity of the Pyrogallol against *Staphylococcus aureus* Evaluated by Optical Image. *Biologics*, 2(2), 139–150. https://doi.org/10.3390/biologics2020011
- Olivia, N. U., Goodness, U. C., & Obinna, O. M. (2021). Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of *Hibiscus asper* leaves. *Future Journal of Pharmaceutical Sciences*, 7(1). https://doi.org/10.1186/s43094-021-00208-4
- Otsuka, H., Hirata, E., Shizato, T., & Takeda, Y. (2000). Isolation of Lignan Glucosides and Neolignan sulfate from the leaves of *Glochidion zeylanicum* (Gaertn.) A. Juss. *Chemical and Pharmaceutical Bulletin*, 48(7), 1084–1086.
- Otsuka, H., Hirata, E., Shinzato, T., & Takeda, Y. (2003). Stereochemistry of megastigmane glucosides from *Glochidion zeylanicum* and *Alangium premnifolium*. *Phytochemistry*, 62, 763–768.
- Ozdemir, G., Horzum, Z., Sukatar, A., & Karabay-Yavasoglu, N. U. (2006). Antimicrobial Activities of Volatile Components and Various Extracts of *Dictyopteris membranaceae* and *Cystoseira barbata* from the Coast of Izmir, Turkey. *Pharmaceutical Biology*, 44(3), 183–188. https://doi.org/10.1080/13880200600685949
- Ozsen Batur, O. & atlı, Özlem, & Ismail, K. (2019). Biotransformation of oleic acid and antimicrobial and anticancer activities of its biotransformation extracts. *Bulgarian Chemical Communications*, 51(2), 200–205. http://dx.doi.org/10.34049/bcc.51.2.4831
- Padma, M., Ganesan, S., Jayaseelan, T., Azhagumadhavan, S., Sasikala, P., Senthilkumar, S., & Mani, P. (2019). Phytochemical screening and GC–MS analysis of bioactive compounds present in ethanolic leaves extract of *Silybum marianum* (L). *Journal of Drug Delivery and Therapeutics*, 9(1), 85–89. https://doi.org/10.22270/jddt.v9i1.2174
- Padma, R., Parvathy, N. G., Renjith, V., & Kalpana, P. R. (2013). Quantitative estimation of tannins, phenols and antioxidant activity of methanolic extract of *Imperata*

- cylindrica. International Journal of Research in Pharmaceutical Sciences, 4(1), 73–77.
- Painuli, S., Rai, N., & Kumar, N. (2016). GC-MS analysis of methanolic extract of leaves of *Rhododendron arboreum*. Asian Journal of Pharmaceutical and Clinical Research, 9(1), 101–104.
- Pathan, A. K., Bond, J., & Gaskin, R. E. (2010). Sample preparation for SEM of plant surfaces. *Materials Today*, 12(1), 32–43. https://doi.org/10.1016/S1369-7021(10)70143-7
- Paul, S. H., Usman, A. A., Gana, I. N., Manase, A., Adeniyi, O. D., & Olutoye, M. A. (2018). Comparative Study of Mineral and Nutritional Composition of a Multifunctional Flora Composite Formulated from Seven Medicinal Plants and their Applications to Human Health. *Engineering Technology Open Access*, 1(5), 555572. DOI: 10.19080/ETOAJ.2018.01.555572
- Pauletti, P. M., Araújo, A. R., Young, M. C., Giesbrecht, A. M., & Bolzani, V. D. (2000). nor-Lignans from the leaves of *Styrax ferrugineus* (Styracaceae) with antibacterial and antifungal activity. *Phytochemistry*, *55*(6), 597–601. https://doi.org/10.1016/s0031-9422(00)00225-9
- Payne, W. W. (1978). A glossary of Plant hair Terminology. *Brittonia*, 30(2), 239–255.
- Pequerul, A., Pérez, C., Madero, P., Val, J., & Monge, E. (1993). A rapid wet digestion method for plant analysis. In *Optimization of Plant Nutrition* (pp. 3–6). Springer Netherlands. https://doi.org/10.1007/978-94-017-2496-8_1
- Ponnudurai, G., & Peter Paul, J. J. (2020). GC-MS Analysis of Methanolic Extract of Colpomenia Sinuosa (Mertens Ex Roth) Derb. Et Sol. From Manapad in the South East Coast of Tamil Nadu, India. *Asian Journal of Pharmaceutical Research and Development*, 8(4), 41–43. https://doi.org/10.22270/ajprd.v8i4.761
- POWO (2021). Plants of the Word Online. Facilitated by the Royal Botanical Gardens, Kew. Published on the Internet http://www.plantsoftheworldonline.org/ (Accessed on 5th May 2021).
- Pratama, O. A., Sri Tunjung, W. A., Sutikno, S., & Daryono, B. S. (2019). Bioactive compound profile of melon leaf extract (*Cucumis melo* L. 'Hikapel') infected by

- downy mildew. *Biodiversitas*, 20(11), 3448–3453. https://doi.org/10.13057/biodiv/d201143
- Preetha, S. S. (2007). *Flora of Kerala family Euphorbiaceae*. Ph.D Thesis. Department of Post graduate Studies and Research in Botany. Sanatana Dharma College, Sanatanapuram, Alappuzha-688003. University of Kerala.
- Pruesapan, K., Telford, I. R., Bruhl, J. J., & van Welzen, P. C. (2012). Phylogeny and proposed circumscription of *Breynia*, *Sauropus* and *Synostemon* (Phyllanthaceae), based on chloroplast and nuclear DNA sequences. *Australian Systematic Botany*, 25(5), 313–330. http://dx.doi.org/10.1071/SB11005
- Puapairoj, P., Naengchomnong, W., Kijjoa, A., Pinto, M. M., Pedro, M., Nascimento, M. S. J., Silva, A. M. S., & Herz, W. (2005). Cytotoxic activity of lupane-type triterpenes from *Glochidion sphaerogynum* and *Glochidion eriocarpum* two of which induce apoptosis. *Planta Medica*, 71(3), 208–213. https://doi.org/10.1055/s-2005-837818
- Raaman, N. (2006). Phytochemical Techniques. New Delhi, India.
- Radford, A. E., Dickson, W. C., Massey, J. R., & Bell, C. L. (1974). *Vascular Plant Systematic*. Harper and Raw Publisher, New York.
- Radha, Kumar, M., Puri, S., Pundir, A., Bangar, S. P., Changan, S., Choudhary, P.,
 Parameswari, E., Alhariri, A., Samota, M. K., Damale, R. D., Singh, S., Berwal,
 M. K., Dhumal, S., Bhoite, A. G., Senapathy, M., Sharma, A., Bhushan, B., &
 Mekhemar, M. (2021). Evaluation of nutritional, phytochemical, and mineral composition of selected medicinal plants for therapeutic uses from cold desert of
 Western Himalaya. *Plants*, 10(7), 1429. https://doi.org/10.3390/plants10071429
- Rahaman, M. M., Hassan, S. M. H., Martorell, M., Sharifi-Rad, J., & Islam, M. T. (2020). Ascorbic acid interaction with phytol: a modulatory effects on the anti-pyretic activity of paracetamol in Swiss albino mice. *Clinical Phytoscience*, *6*, 54. https://doi.org/10.1186/s40816-020-00200-0
- Rajkumari, R., Singh, P. K., Das, A. K., & Dutta, B. K. (2013). Ethnobotanical investigation of wild edible and medicinal plants used by the Chiru Tribe of Manipur, India. *Pleione*, 7(1), 167–174.

- Ralte, L., Khiangte, L., Thangjam, N. M., Kumar, A., & Singh, Y. T. (2022). GC-MS and molecular docking analyses of phytochemicals from the underutilized plant, *Parkia timoriana* revealed candidate anti-cancerous and anti-inflammatory agents. *Scientific reports*, 12(1), 3395. https://doi.org/10.1038/s41598-022-07320-2
- Rao, N. V. R., Nadendla, R., Arunkumar, M., Chowdary, B. S., Reddy, B. S., Nirojini, P. S., & Begum, S. S. (2013). Antidiabetic activity of *Glochidion velutinum* L. extract in Alloxan-induced type II diabetic rats. *International Journal of Phytopharmacology*, 4(5), 311–314.
- Rana, T. S., Nair, K. N., & Upreti, D. K. (2014). Plant Taxonomy and Biosystematics: Classical and Modern Methods. New India Publishing Agency, New Delhi-110034, India.
- Rasingam, L., Chorghe, A. R., Prasanna, P. V., & Sankara Rao, M. (2014). *Glochidion tirupathiense* (Phyllanthaceae) A new species from Seshachalam Biosphere reserve of Andhra Pradesh, India. *Taiwania*, 59(1), 9–12. https://doi.org/10.6165/tai.2014.59.9
- Rathod, V., & Rajurkar, N. S. (2017). Phytochemical screening and antioxidant activity of *Glochidion ellipticum. Journal of Applicable Chemistry*, 6(2), 219–226.
- Rawat, P., Bachheti, R. K., Kumar, N., Rai, N. (2018). Phytochemical analysis and evaluation of in vitro immunomodulatory activity of *Rhododendron arboreum* leaves. *Asian Journal of Pharmaceutical and Clinical Research*, *11*(8), 123–128. https://doi.org/10.22159/ajpcr.2018.v11i8.25372
- Razzaq, A., Shahid, S., Akram, M., Ashraf, M., Iqbal, S., Hussain, A., Azam Zia, M., Qadri, S., Saher, N., Shahzad, F., Shah, A. N., Rehman, A. U., & Jacobsen, S. E. (2021). Stomatal State Identification and Classification in Quinoa Microscopic Imprints through Deep Learning. *Complexity*, 2021. https://doi.org/10.1155/2021/9938013
- Reddy, S. R., Reddy, K. N., Pattanaik, C., Raju, V. S., & Autonagar, J. (2006).
 Ethnobotanical Observations on some Endemic Plants of Eastern Ghats, India.
 Ethnobotany Leaflet, 10, 82–91.

- Reza, A. S. M. A., Haque, M. A., Sarker, J., Nasrin, M. S., Rahman, M. M., Tareq, A. M., Khan, Z., Rashid, M., Sadik, M. G., Tsukahara, T., & Alam, A. K. (2021). Antiproliferative and antioxidant potentials of bioactive edible vegetable fraction of *Achyranthes ferruginea* Roxb. in cancer cell line. *Food Science & Nutrition*, 9(7), 3777–3805. https://doi.org/10.1002/fsn3.2343
- Saeid, H., Al-sayed, H., & Bader, M. (2023). A Review on Biological and Medicinal Significance of Furan. *AlQalam Journal of Medical and Applied Sciences*, 6(1), 44–58. https://doi.org/10.5281/zenodo.7650255
- Safara, S., Harighi, B., Bahramnejad, B., & Ahmadi, S. (2022). Antibacterial Activity of Endophytic Bacteria Against Sugar Beet Root Rot Agent by Volatile Organic Compound Production and Induction of Systemic Resistance. *Frontiers in Microbiology*, 13, 921762. https://doi.org/10.3389/fmicb.2022.921762
- Saha, M., & Bandyopadhyay, P. K. (2020). In vivo and in vitro antimicrobial activity of phytol, a diterpene molecule, isolated and characterized from *Adhatoda vasica* Nees. (Acanthaceae), to control severe bacterial disease of ornamental fish, Carassius auratus, caused by *Bacillus licheniformis* PKBMS₁₆. *Microbial Pathogenesis*, 141, 103977. https://doi.org/10.1016/j.micpath.2020.103977
- Sahu, J. K. (2019). Development of Sustainable Rural Livelihood Options Utilizing Locally Available Bio-Resources Through Transformative Rural Technologies in the Indian Himalayan Regions of Himachal Pradesh and Sikkim 2019-20. Principal Investigator Centre for Rural Development and Technology Indian Institute of Technology Delhi Hauz Khas, New Delhi 110 016.
- Salim, A. S. (2018). In Vitro Induction of Callus from Different Explants of *Terminalia arjuna* (Roxb.) Wight and Arn. and Detection of its Active Secondary Metabolites Using GC-MS Analysis. *Plant Archives*, *18*(2), 2519–2527.
- Sandhya, S., Chaitanya, R. S. N. A. K. K., Vinod, K. R., Rao, K. N. V., Banji, D., Sudhakar, K., & Swetha, R. (2010). An updated review on the genus *Glochidion* plant. *Archives of Applied Science Research*, 2(2), 309–322.
- Santos, P. L., Matos, J. P. S. C. F., Picot, L., Almeida, J. R. G. S., Quintans, J. S. S., & Quintans-Júnior, L. J. (2019). Citronellol, a monoterpene alcohol with promising pharmacological activities A systematic review. *Food and Chemical Toxicology:*

- An International Journal Published for the British Industrial Biological Research Association, 123, 459–469. https://doi.org/10.1016/j.fct.2018.11.030
- Saravanan, D., Kasisankar, V., & Asharani, I. V. (2013). GC-MS analysis of phytocomponents in the leaves of *Actinodaphne madraspatana* Bedd. International Journal Research in Pharmaceutical Sciences, 4, 469–473
- Saupi, N., Zakaria, M. H., & Bujang, J. S. (2009). Analytic chemical composition and mineral content of yellow velvet leaf (*Limnocharis flava* L. Buchenau)'s edible parts. *Journal of Applied Sciences*, 9(16), 2969–2974. https://doi.org/10.3923/jas.2009.2969.2974
- Savithramma, N., Yugandhar, P., Haribabu, R., & Prasad, K. S. (2014). Validation of Indigenous Knowledge of Yanadi Tribe and Local Villagers of Veyilingalakona-A Sacred Grove of Andhra Pradesh, Indian. *Journal of Pharmaceutical Sciences and Research*, 6, 382–388.
- Saxena, V., Mishra, G., Saxena, A., Kamlesh, K. R., & Vishwakarma. (2013). A comparative study on quantitative estimation of tannins in *Terminalia chebula*, *Terminalia belerica*, *Terminalia arjuna* and *Saraca indica* using spectrophotometer. *Asian Journal of Pharmaceutical and Clinical Research*, 6(7), 148–49.
- Senguttuvan, J., Paulsamy, S., & Karthika, K. (2014). Phytochemical analysis and evaluation of leaf and root parts of the medicinal herb, *Hypochaeris radicata* L. for in vitro antioxidant activities. *Asian Pacific Journal of Tropical Biomedicine*, 4, S359–S367. https://doi.org/10.12980/APJTB.4.2014C1030
- Serafim, A., Company, R., Lopes, B., Rosa, J., Cavaco, A., Castela, G., Castela, E., Olea, N., & Bebianno, M. J. (2012). Assessment of essential and nonessential metals and different metal exposure biomarkers in the human placenta in a population from the South of Portugal. *Journal of Toxicology and Environmental Health Part A:***Current** Issues, 75(13–15), 867–877.

 https://doi.org/10.1080/15287394.2012.690704
- Shah, S. L., Bashir, K., Rasheed, H. M., Rahman, J. U., Ikram, M., Shah, A. J., Majrashi, K. A., Alnasser, S. M., Menaa, F., & Khan, T. (2022). LC-MS/MS-Based Metabolomic Profiling of Constituents from *Glochidion velutinum* and its activity

- against cancer cell lines. *Molecules (Basel, Switzerland)*, 27(24), 9012. https://doi.org/10.3390/molecules27249012
- Shahzad, K., Zafar, M., Khan, A. M., Mahmood, T., Abbas, Q., Ozdemir, F. A., Ahmad, M., & Sultana, S. (2022). Characterization of anatomical foliar epidermal features of herbaceous flora of Tilla Jogian, Pakistan by using light microscopy techniques.

 *Microscopy** Research** and *Technique*, *85(1), 135–148.

 https://doi.org/10.1002/jemt.23890
- Shantabi, L., Jagetia, G. C., Ali, M. A., Singh, T. T., and Devi, S. V. (2014). Antioxidant potential of *Croton caudatus* leaf extract In vitro. *Translational Medicine and Biotechnology*, 2(6), 1–15. https://www.researchgate.net/publication/304792401
- Sharma, D., Rani, R., Chaturvedi, M., & Yadav, J. P. (2018). Antibacterial capacity and identification of bioactive compounds by GC/MS of *Allium cepa*. *International Journal of Pharmacy and Pharmaceutical Sciences*, 10(2), 116–121. http://dx.doi.org/10.22159/ijpps.2018v10i2.23698
- Sharma, J. V., Malleswari, G. N., Rao, J. V., Muralibalaram, V., & Sangeetha, R. (2010).
 Antimicrobial Activity of Root Extracts of *Glochidion zeylanicum*. *International Journal of Chemical Sciences*, 8, 1088–1090.
- Sharma, J. V. C., Shekar, B. C., Chakraborty, R., Chanda, H., Rao, J. V. & Kumar, H. S., (2011). Anticancer activity of aqueous extract of roots of *Glochidion zeylanicum* (Gaertn.) A. Juss. *Journal of Pharmaceutical and Biomedical Sciences*, 6(6), 1–4.
- Sharma, K., Assefa, A. D., Kim, S., Ko, E. Y., Lee, E. T., & Park, S. W. (2014). Evaluation of total phenolics, flavonoids and antioxidant activity of 18 Korean onion cultivars: A comparative study. *Journal of the Science of Food and Agriculture*, *94*(8), 1521–1529. https://doi.org/10.1002/jsfa.6450
- Sharma, N., Palia, P., Chaudhary, A., Shalini, Verma, K., & Kumar, I. (2020). A Review on Pharmacological Activities of Lupeol and its Triterpene Derivatives. *Journal of Drug Delivery and Therapeutics*, 10(5), 325-332. https://doi.org/10.22270/jddt.v10i5.4280
- Shukla, R. K., Painuly, D., Porval, A., & Shukla, A. (2014). Proximate analysis, nutritional value, phytochemical evaluation, and biological activity of *Litchi chinensis* Sonn.

- leaves. *Journal of Herbs, Spices and Medicinal Plants*, 20(2), 196–208. https://doi.org/10.1080/10496475.2013.848830
- Silva Pereira, R. L., Campina, F. F., Costa, M. D. S., Pereira da Cruz, R., Sampaio de Freitas, T., Lucas Dos Santos, A. T., Cruz, B. G., Maciel de Sena Júnior, D., Campos Lima, I. K., Xavier, M. R., Rodrigues Teixeira, A. M., Alencar de Menezes, I. R., Quintans-Júnior, L. J., Araújo, A. A. S., & Melo Coutinho, H. D. (2021). Antibacterial and modulatory activities of β-cyclodextrin complexed with (+)-β-citronellol against multidrug-resistant strains. *Microbial Pathogenesis*, *156*, 104928. https://doi.org/10.1016/j.micpath.2021.104928
- Silva, R. O., Sousa, F. B., Damasceno, S. R., Carvalho, N. S., Silva, V. G., Oliveira, F. R., Sousa, D. P., Aragão, K. S., Barbosa, A. L., Freitas, R. M., & Medeiros, J. V. (2014). Phytol, a diterpene alcohol, inhibits the inflammatory response by reducing cytokine production and oxidative stress. *Fundamental & Clinical Pharmacology*, 28(4), 455–464. https://doi.org/10.1111/fcp.12049
- Siswadi, S., & Saragih, G. S. (2021). Phytochemical analysis of bioactive compounds in ethanolic extract of *Sterculia quadrifida* R. Br. *In: AIP Conference Proceedings*, 2353(1), 030098. http://dx.doi.org/10.1063/5.0053057
- Skowyra, M., Gallego, M. G., Segovia, F., & Almajano, M. P. (2014). Antioxidant properties of *Artemisia annua* extracts in model food emulsions. *Antioxidants*, *3*(1), 116–128. https://doi.org/10.3390/antiox3010116
- Soetan, K. O., Olaiya, C. O., & Oyewole, O. E. (2010). The importance of mineral elements for humans, domestic animals and plants: A review. *African Journal of Food Science*, 4(5), 200–222. http://www.academicjournals.org/ajfs
- Solihani, N. S., Noraini, T., Azahana, A., & Nordahlia, A. S. (2015). Leaf micromorphology of some *Phyllanthus* L. species (Phyllanthaceae). *AIP Conference Proceedings*, 1678. https://doi.org/10.1063/1.4931207
- Soni, V., Jha, A. K., Dwivedi, J., & Soni, P. (2018). Qualitative and quantitative determination of phytoconstituents in some antifertility herbs. *Indian Journal of Pharmaceutical Sciences*, 80(1), 79–84. http://dx.doi.org/10.4172/pharmaceutical-sciences.1000332

- Sorlozano-Puerto, A., Albertuz-Crespo, M., Lopez-Machado, I., Gil-Martinez, L., Ariza-Romero, J. J., Maroto-Tello, A., Baños-Arjona, A., & Gutierrez-Fernandez, J. (2020). Antibacterial and Antifungal Activity of Propyl-Propane-Thiosulfinate and Propyl-Propane-Thiosulfonate, Two Organosulfur Compounds from *Allium cepa*: In Vitro Antimicrobial Effect via the Gas Phase. *Pharmaceuticals (Basel, Switzerland)*, *14*(1), 21. https://doi.org/10.3390/ph14010021
- Srivastava, R., & Kulshreshtha, D. K. (1988). Triterpenoids from *Glochidion heyneanum*. *Phytochemistry*, 27(11), 3575–3578.
- Stace, C. A. (1969). The Significance of the Leaf Epidermis in the Taxonomy of the Combretaceae III. The Genus *Combretum* in America. *Brittonia*, 21(2), 130–143.
- Suárez-Quiroz, M. L., Taillefer, W., López Méndez, E. M., González-Ríos, O., Villeneuve, P., & Figueroa-Espinoza, M. C. (2013). Antibacterial activity and antifungal and anti-mycotoxigenic activities against *Aspergillus flavus* and *A. ochraceus* of green coffee chlorogenic acids and dodecyl chlorogenates. *Journal of Food Safety*, 33(3), 360–368. http://dx.doi.org/10.1111/jfs.12060
- Subramanian, R., Gayathri, S., Rathnavel, C., & Raj, V. (2012). Analysis of mineral and heavy metals in some medicinal plants collected from local market. *Asian Pacific Journal of Tropical Biomedicine*, 2(1 SUPPL.). https://doi.org/10.1016/S2221-1691(12)60133-6
- Sun, Y., Wang, C., Du, G., Deng, W., Yang, H., Li, R., Xu, Q., & Guo, Q. (2022). Two Nematicidal Compounds from *Lysinimonas* M4 against the Pine Wood Nematode, *Bursaphelenchus xylophilus*. *Forests*, 13(8), 1191. https://doi.org/10.3390/f13081191
- Supreeth, M., Chandrashekar, M. A., Sachin, N., & Raju, N. S. (2016). Effect of chlorpyrifos on soil microbial diversity and its biotransformation by *Streptomyces* sp. HP-11. *3 Biotech*, *6*(2), 147. https://doi.org/10.1007/s13205-016-0462-2
- Sunkara, S., Tripuramallu, S., & Reddy Maram, V. R. (2009). Antioxidant activity of *Glochidion zeylanicum* leaves extract. *Research Journal of Pharmacognosy and Phytochemistry*, 1(3), 224–226.

- Suryowati, T., Forman, E., Maheshwari, H., Diani, Y. H., & Kusuma, R. A. (2023). Bioactive compounds, antidiabetic and antimicrobial potential of pinang seeds extract (*Areca catechu* 1). *Jurnal Aisyah: Jurnal Ilmu Kesehatan*, 8(3). http://dx.doi.org/10.30604/jika.v8i3.2060
- Tadavi, S. C., & Bhadane, V. V. (2014). Leaf Architecture in some Euphorbiaceae. *Indian Journal of Applied & Pure Biology*, 29(2), 343–360.
- Taj, T., Sultana, R. S., Shahin, H., Chakraborty, M., Mohammed, G., & Ahmed, M. I. (2021). Phytol: Aphytoconstituent, its chemistry and pharmacological actions. GIS-Zeitschrift füGeoinformatik (Geoinfomatics and digital earthinitiatives), 8(1), 395–406.
- Tanaka, R., Kinouchi, Y., Wada, S.-I., Tokuda, H., Tanaká, R., Stuttgart´, S., York´doi, S., & York´doi, Y. (2004). Potential Anti-Tumor Promoting Activity of Lupane-Type Triterpenoids from the Stem Bark of *Glochidion zeylanicum* and *Phyllanthus flexuosus*. *Planta Medica*, 70(12), 1234–6. <a href="https://doi.org/10.1055/s-2004-835858'ISSN835858'835858'ISSN
- Tanod, W. A., Yanuhar, U., Wahyudi, D., & Risjani, Y. (2019). DPPH scavenging property of bioactives from soft corals origin Palu Bay, Central Sulawesi, Indonesia. *In IOP Conference Series: Earth and Environmental Science*, 236(1), 012121. IOP Publishing. https://doi.org/10.1088/1755-1315/236/1/012121
- Tchounwou, P. B., Yedjou, C. G., Patlolla, A. K., & Sutton, D. J. (2012). Heavy metal toxicity and the environment. *EXS*, 101, 133–164. https://doi.org/10.1007/978-3-7643-8340-4-6
- Thakur, H. A., & Patil, D. A. (2011). The foliar epidermal studies in some hitherto unstudied Euphorbiaceae. *Current Botany*, 2(4), 22–30.
- Thakur, H. A., & Patil, D. A. (2014). Foliar Epidermal Studies of Plants in Euphorbiaceae. *Taiwania*, 59, 59–70. http://dx.doi.org/10.6165/tai.2014.59.59
- Thang, N. V., Thu, V. K., Nhiem, N. X., Dung, D. T., Quang, T. H., Tai, B. H., Anh, H. L. T., Yen, P. H., Ngan, N. T. T., Hoang, N. H., & Kiem, P. V. (2017). Oleanane-type Saponins from *Glochidion hirsutum* and their cytotoxic activities. *Chemistry*

https://doi.org/10.1002/cbdv.201600445

- The Angiosperm Phylogeny Group. (1998). An Ordinal Classification for the Families of Flowering Plants. *Annals of the Missouri Botanical Garden*, 85(4), 531–553. https://doi.org/10.2307/2992015
- The Herbarium Catalogue, Royal Botanic Gardens, Kew. (2020). Published on the Internet http://www.kew.org/herbcat (Accessed on 23rd May 2020).
- The Plant List (2013). Version 1.1. Published on the Internet http://www.theplantlist.org/
 (Accessed on 20th April 2021).
- The World's Herbaria (2018). A Summary Report Based on Data from Index Herbariorum Issue 3.0, published January 10, 2019, Barbara M. Thiers Editor, Index Herbariorum.
- Thiers B (2013). [continuously updated] Index Herbarium: a global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. The New York Botanical Garden, New York. http://sweetgum.nybg.org/ih/ (Accessed on 25th May 2020).
- Thomas, R. A., & Krishnakumari, S. (2015). Proximate analysis and mineral composition of *Myristica fragrans* seeds. *Journal of Pharmacognosy and Phytochemistry*, *3*(6), 39–42.
- Thu, V. K., Van Thang, N., Nhiem, N. X., Tai, B. H., Nam, N. H., Kiem, P. V., Minh, C. V., Anh, H. L. T., Kim, N., Park, S., & Kim, S. H. (2015). Oleanane-type saponins from *Glochidion glomerulatum* and their cytotoxic activities. *Phytochemistry*, 116, 213–220. https://doi.org/10.1016/j.phytochem.2015.05.001
- Tian, Y., Fan, R., Yin, Z., Huang, Y., Huang, D., Yuan, F., Yin, A., Tang, G., Pu, R., & Yin, S. (2023). Glochodpurnoid B from *Glochidion puberum* Induces Endoplasmic Reticulum Stress-Mediated Apoptosis in Colorectal Cancer Cells. *Molecules (Basel, Switzerland)*, 28(2), 511. https://doi.org/10.3390/molecules28020511
- Togashi, N., Shiraishi, A., Nishizaka, M., Matsuoka, K., Endo, K., Hamashima, H., & Inoue, Y. (2007). Antibacterial activity of long-chain fatty alcohols against

- Staphylococcus aureus. Molecules (Basel, Switzerland), 12(2), 139–148. https://doi.org/10.3390/12020139
- Toor, R. H., Khan, Z. N., Tariq, M., Tassaduq, R., Gardner, Q. A., Waheed-Uz-Zaman, W., Lian, J. B., Stein, J. L., Stein, G. S., & Shakoori, A. R. (2020). Bioactivity-Guided Isolation and Identification of Anti-adipogenic Constituents from the n-Butanol Fraction of *Cissus quadrangularis*. *Critical reviews in eukaryotic gene expression*, 30(6), 519–541. https://doi.org/10.1615/CritRevEukaryotGeneExpr.2020036843
- Traiperm, P., Chow, J., Nopun, P., Staples, G., & Swangpol, S. C. (2017). Identification among morphologically similar *Argyreia* (Convolvulaceae) based on leaf anatomy and phenetic analyses. *Botanical Studies*, 58(1), 1–14. https://doi.org/10.1186/s40529-017-0178-6
- Trease, E. C., & Evans, W. C. (2009). *Pharmacognosy*. Sixteenth Edition. Edinburgh London New York Philadelphia St Louis Sydney Toronto.
- Van Cotthem, W. R. J. (1970). A classification of stomatal types. *Botanical Journal of the Linnean Society*, 63(3), 235–246. https://doi.org/10.1111/j.1095-8339.1970.tb02321.x
- Vandana, & Deora, G. S. (2020). Preliminary phytochemical screening and GC-MS analysis of methanolic leaf extract of *Tephrosia falciformis* Ramaswami from Indian Thar Desert. *International Journal of Pharmaceutical Sciences & Research*, 11(6), 3040–3046. doi: 10.13040/IJPSR.0975-8232.11(6).3040-46.
- van Dronkelaar, C., van Velzen, A., Abdelrazek, M., van der Steen, A., Weijs, P. J. M., & Tieland, M. (2018). Minerals and Sarcopenia; The Role of Calcium, Iron, Magnesium, Phosphorus, Potassium, Selenium, Sodium, and Zinc on Muscle Mass, Muscle Strength, and Physical Performance in Older Adults: A Systematic Review. *Journal of the American Medical Directors Association*, 19(1), 6–11.e3. https://doi.org/10.1016/j.jamda.2017.05.026
- van Welzen, P. C., Pruesapan, K., Telford, I. R. H., Esser, H. J., & Bruhl, J. J. (2014). Phylogenetic reconstruction prompts taxonomic changes in *Sauropus*, *Synostemon* and *Breynia* (Phyllanthaceae tribe Phyllantheae). *Blumea: Journal of Plant*

- *Taxonomy and Plant Geography*, *59*(2), 77–94. https://doi.org/10.3767/000651914X684484
- Vasco, A., Thadeo, M., Conover, M., & Daly, D. C. (2014). Preparation of Samples for Leaf architecture studies, a method for mounting cleared leaves. *Application in Plant Sciences*, 2(9), 1400038.
- Venn-Watson, S., & Schork, N. J. (2023). Pentadecanoic Acid (C15:0), an Essential Fatty Acid, Shares Clinically Relevant Cell-Based Activities with Leading Longevity-Enhancing Compounds. *Nutrients*, *15*(21), 4607. https://doi.org/10.3390/nu15214607
- Vinayaka, K. S., Hallur, R. L. S., & Prashith, K. T. R. (2022). Preliminary phytochemical analysis and in vitro antioxidant activity of *Glochidion ellipticum* Wight (Phyllanthaceae). *Biomedicine* (*India*), 42(1), 148–153. https://doi.org/10.51248/.v42i1.654
- Voigt, J. O. (1841). Hortus Suburbanus Calcuttensis: A catalogue of the plants which have been cultivated in the Hon. East India Company's Botanical Garden, Calcutta, and in the Serampore Botanical Garden. Calcutta Bishops College Press, Calcutta.
- Wal, P., Wal, A., Sharma, G., & Rai, A. K. (2011). Biological activities of Lupeol. Systematic Reviews in Pharmacy, 2(2), 96–103. https://doi.org/10.4103/0975-8453.86298
- Webster, G. L. (2014). Euphorbiaceae. In: Kubitzki K (Eds) Flowering Plants. Eudicots. The Families and Genera of Vascular Plants 11. Springer, Berlin, Heidelberg, 51–216. https://doi.org/10.1007/978-3-642-39417-1_10
- WFO (2023). World Flora Online. Published on the Internet http://www.worldfloraonline.org. (Accessed on 22nd June 2023).
- Wiart, Christophe (2022). Medicinal Plants in the Asia Pacific for Zoonotic Pandemics Volume 2. CRC Press, Taylor & Francis Group, LLC.
- Wiraswati, H. L., Fauziah, N., Pradini, G. W., Kurnia, D., Kodir, R. A., Berbudi, A., Arimdayu, A. R., Laelalugina, A., Supandi, & Ma'ruf, I. F. (2023). *Breynia cernua*: Chemical Profiling of Volatile Compounds in the Stem Extract and Its Antioxidant,

- Antibacterial, Antiplasmodial and Anticancer Activity In Vitro and In Silico. *Metabolites*, 13(2), 281. https://doi.org/10.3390/metabo13020281
- Wrona, M., Pezo, D., Rovito, M. A., Vera, P., Nerín, C., & Asensio, E. (2022). Application of Untargeted Metabolomics to Determine Volatile Compounds from the Spanish Plant *Arctostaphylos uva-ursi* Used as Tea. *Separations*, *9*(*3*), 68. https://doi.org/10.3390/separations9030068
- Xiao, H. T., Hao, X. Y., & Yang, X. W. (2007). Bisabolane-Type Sesquiterpenoids from the Rhizomes of *Glochidion coccineum*. *Helvetica Chimica Acta*, *90*, 164–170. https://doi.org/10.1002/hlca.200790011
- Xiao, H. T., Wang, Y. H., Hao, X. Y., Yang, X. S. & Hao, X. J. (2008a). Triterpenes from Glochidion coccineum and their Cytotoxicity in Vitro. Lishizhen Medicine and Materia Medica Research, 19, 1931–1932.
- Xiao, H. T., He, H. P., Peng, J., Wang, Y. H., Yang, X. W., Hu, X. J., Hao, X. Y., & Hao, X. J. (2008b). Two new norbisabolane sesquiterpenoid glycosides from *Glochidion coccineum*. *Journal of Asian Natural Products Research*, 10, 1–5. https://doi.org/10.1080/10286020701189393
- Xu, Z. L., Hang, Y. Y., & Lei, L. G. (2020). Phyllanthaceae. In: Li DZ (Ed.) The Families and Genera of Chinese Vascular Plants (Vol. II). Science Press, Beijing, 1238– 1248.
- Yao, G., Song, Z. Q., Xue, B. E., Shi, S., Li, Y. L., & Luo, S. X. (2020). Taxonomic revision of the genus *Glochidion* (Phyllanthaceae) in Taiwan, China. *PhytoKeys*, *159*, 137–159. https://doi.org/10.3897/phytokeys.159.54839
- Yao, G., Song, Z., Wu, Y., Kim Thanh, N. T., Li, Y., Shi, S., & Luo, S. (2018). Taxonomic studies of *Glochidion* (Phyllanthaceae) from the Indo-China Peninsula (II): The identities of *G. anamiticum* and *G. annamense*. *PhytoKeys*, 114, 1–9. https://doi.org/10.3897/phytokeys.114.30725
- Yao, G., & Zhang, D. (2015). Taxonomic notes on *Glochidion acuminatum* and *G. triandrum* (Phyllanthaceae). *Phytotaxa*, 236(1), 79–85. https://doi.org/10.11646/phytotaxa.236.1.7

- Yasa, S. R., Poornachandra, Y., Kumar, C. G., & Penumarthy, V. (2017). Synthesis, Characterization, Antimicrobial and Anti-Biofilm Activity of a New Class of 11-Bromoundecanoic Acid-Based Betaines. *Medicinal Chemistry Research*, 26(10), 2592–2601. https://doi.org/10.1007/s00044-017-1958-y.
- Yi, J., Wu, J. G., Wu, Y. Bin, & Peng, W. (2016). Antioxidant and anti-proliferative activities of flavonoids from *Bidens pilosa* L var. *radiata* Sch Bip. *Tropical Journal of Pharmaceutical Research*, 15(2), 341–348. https://doi.org/10.4314/tjpr.v15i2.17
- Zhu, C., Hu, Y., Mao, H., Li, S., Li, F., Zhao, C., Luo, L., Liu, W., & Yuan, X. (2021). A Deep Learning-Based Method for Automatic Assessment of Stomatal Index in Wheat Microscopic Images of Leaf Epidermis. Frontiers in plant science, 12, 716784. https://doi.org/10.3389/fpls.2021.716784

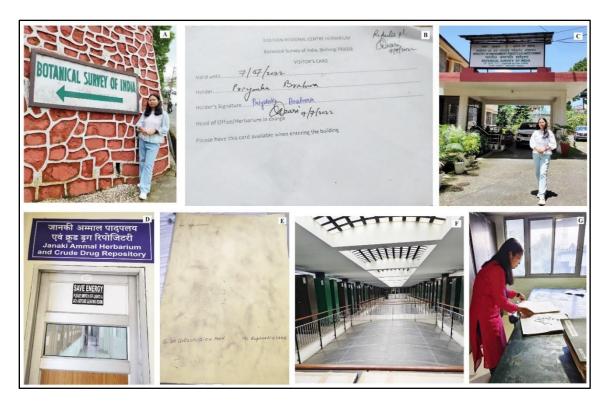


Plate 41. Type specimen study in different Herbaria (A-C). BSI, Shillong, Meghalaya, (D-E). CSIR IIIM, Jammu and (F-G). NBRI, Lucknow

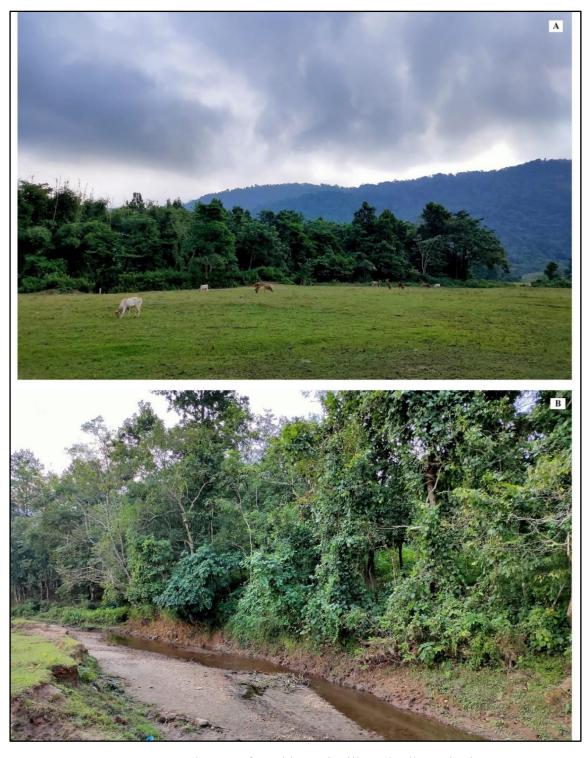


Plate 42. (A-B). Landscape of Baokhungri Hill, Kokrajhar District, Assam

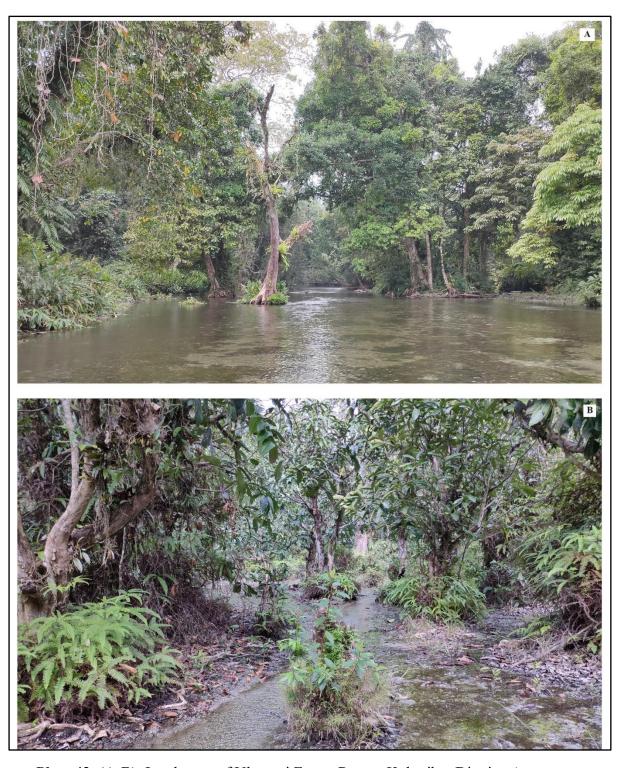


Plate 43. (A-B). Landscape of Ultapani Forest Range, Kokrajhar District, Assam

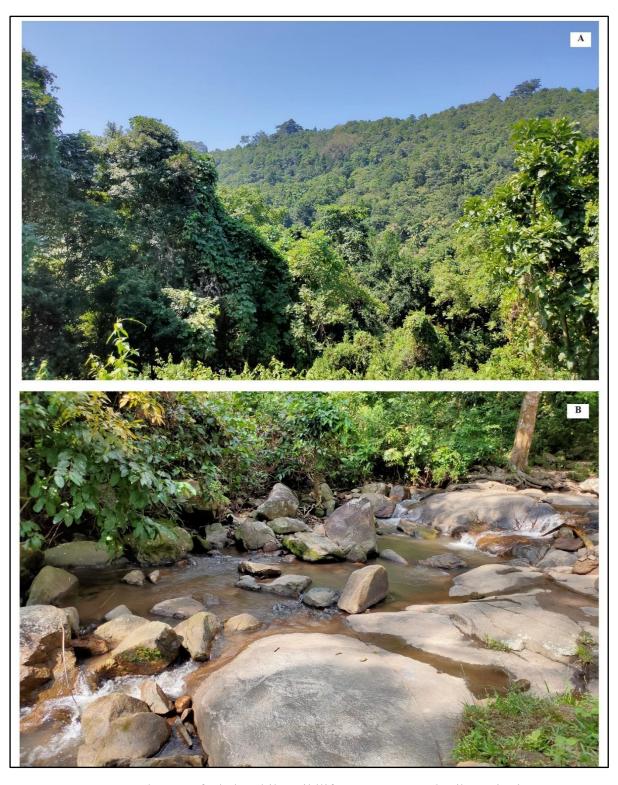


Plate 44. (A-B). Landscape of Chakrashila Wildlife Sanctuary, Kokrajhar District, Assam



Plate 45. (A-B). Landscape of Raimona National Park, Kokrajhar District, Assam



Plate 46. (A-B). Landscape of Owabari, Daolur Dwisa, Kokrajhar District, Assam

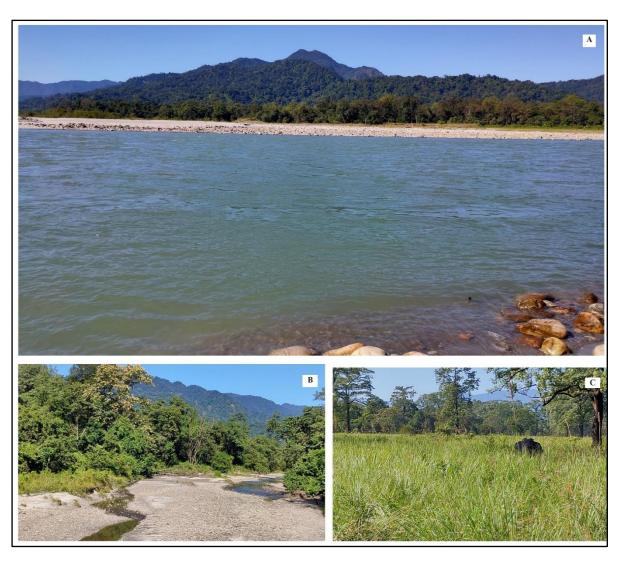


Plate 47. (A-C). Landscape of Manas National Park, Baksa District, Assam



Plate 48. (A-B). Landscape of Orang National Park, Udalguri District, Assam



Plate 49. (A-B). Landscape of Barnodi Wildlife Sanctuary, Udalguri District, Assam (C-D). Inside forest view of Nunai Reserve Forest, Udalguri District, Assam



Plate 50. (A-C). Landscape of Karbi Anglong District, Assam



Plate 51. (A-C). Landscape of Kaziranga National Park, Golaghat District, Assam

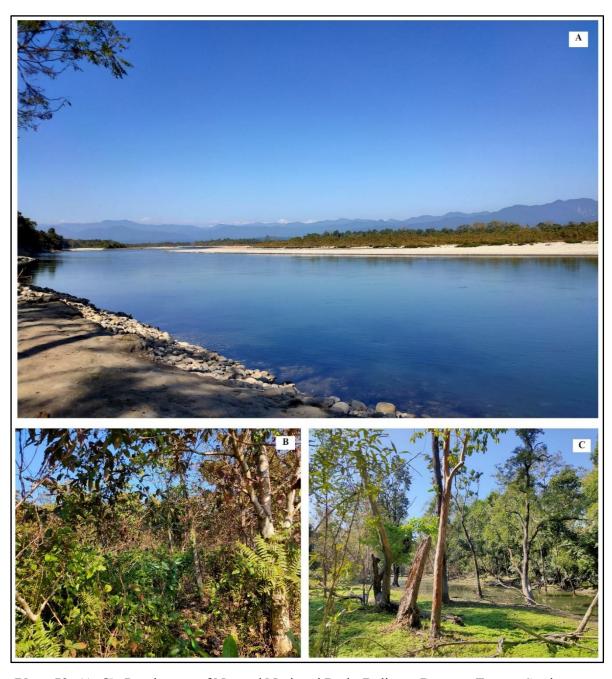


Plate 52. (A-C). Landscape of Nameri National Park, Balipara Reserve Forest, Sonitpur District, Assam



Plate 53. (A-B). Landscape of Cachar District, Assam (C-D). Dimahasao District, Assam



Plate 54. Different habitats of *Glochidion* spp. A. Grassland B. Moist shady area C. Hilly area D. Sal Forest E. Primary Forest F. Road side area G. Moist semi-evergreen forest H. Stream Side area

APPENDIX 1

PUBLICATIONS

- Brahma, P., & Baruah, S. (2023). Investigation of phytochemical constituents, GC-MS, DPPH free radical scavenging assay, and mineral contents of *Glochidion* sphaerogynum (Mull. Arg.) Kurz bark extract. Plant Science Today, 10(2), 98–105. https://doi.org/10.14719/pst.2019 (Figure 34)
- 2. **Brahma, P., & Baruah, S. (2023).** Phytochemical analysis and determination of mineral elements in two species of genus *Glochidion J.R.* Forst. & G. Forst, pp. 43–60. In: P. S. Singh (ed.). *Traditional Medicinal Plants (Volume 7)*. AkiNik publications. http://dx.doi.org/10.22271/ed.book.2290 (**Figure 35**)
- 3. **Brahma, P., & Baruah, S. (2023).** Extended distribution of an endemic variety *Glochidion zeylanicum* var. *paucicarpum* Chakrab. & N.P. Balakr. (Phyllanthaceae) from Assam, India. *Vegetos*. https://doi.org/10.1007/s42535-023-00650-0 (**Figure 36**)
- 4. **Brahma, P., & Baruah, S. (2023).** Comparative morphological and ethnobotanical assessment of certain taxa of genus *Glochidion* (Phyllanthaceae) from Assam, India. *Journal of Threatened Taxa*, 15(12), 24409–24419. https://doi.org/10.11609/jot.8696.15.12.24409-24419 (**Figure 37**)
- Brahma, P., & Baruah, S. (2024). Foliar epidermal micromorphology of genus Glochidion J.R.Forst. & G.Forst. (Phyllanthaceae) by using light and electron microscopy. Current Botany, 15, 47–57. https://doi.org/10.25081/cb.2024.v15.8609 (Figure 38)





RESEARCH ARTICLE

Investigation of phytochemical constituents, GC-MS, DPPH free radical scavenging assay, and mineral contents of *Glochidion* sphaerogynum (Mull. Arg.) Kurz bark extract

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Brahma P, Baruah S. Investigation of phytochemical constituents, GC-MS, DPPH free radical scavenging assay, and mineral contents of Glochidion sphaerogynum (Mull. Arg.) Kurz bark extract. Plant Science Today. 2023; 10(2): 98– 105. https://doi.org/10.14719/pst.2019

Abstract

The aim of the present study was to assess phytochemical constituents, chemical composition, DPPH free radical scavenging assay and mineral contents of Glochidion sphaerogynum (Mull.Arg.) Kurz bark extract. Standard procedures were used to test preliminary phytochemical constituents as well as quantitative analysis for total alkaloid, flavonoid, saponin, tannin, phenolic and terpenoid content. The extract was examined using gas chromatography-mass spectrometry (GC-MS) to know the biologically active compound. In-vitro antioxidant potential was investigated using DPPH free radical scavenging assay and the ICso value for the antioxidant activity of bark extract was 37.4479 µg/mL. The qualitative phytochemical investigation revealed the presence of important phytochemical constituents as well as considerable amounts of total alkaloid, flavonoid, saponin, tannin, phenolic and terpenoid content. GC-MS revealed the presence of biologically active compounds like 1,1,6-Trimethyl-3-methylene-2-(3,6,10,13,14-pentamethyl-3-ethenylpentadec-4-enyl)cyclohexane; Benzenepropanoic acid, 3,5-bis(1,1-dimethyl ethyl) -4-hydroxy-, methyl ester; Neophytadiene etc which would be important for medicinal industries. Mineral contents were determined by using Atomic Absorption Spectrometry (AAS). The result revealed the presence of a good concentration of Na (10.552±0.343 ppm) and Ca (8.973±0.310 ppm) elements followed by K, Fe, Mg and Mn and very less concentrations of heavy metals such as Cd, Cr and Pb indicated the species was devoid of harmful metals.

Keywords

Glochidion sphaerogynum, phytochemical, GC-MS, DPPH, mineral contents

Introduction

The Plant produces many phytochemical constituents that protect them against insects, pathogens, and herbivores (1). Many of these phytochemical constituents also have different biological activities and protect human beings against various diseases (2, 3). According to WHO (World Health Organization), 80% of people rely on indigenous plant-based medicines for curing different diseases (4). Plant produces antioxidants as secondary metabolites, which are substances that act in the cell to counteract free radicals and reactive oxygen species (ROS) and control oxidative damage in the body that become scientifically compelling compounds as a consequence of many pharmacological activities (5, 6). One of the reliable analytical methods for identifying the different chemical components in a plant sample is gas chromatography-mass spectrometry (GC-MS) which plays a significant role in the analysis of phytochemical compounds (7-9).

Plant Science Today, ISSN 2348-1900 (online)

Figure 34. Front page of a published paper

Chapter - 3

Phytochemical Analysis and Determination of Mineral Elements in Two Species of genus *Glochidion J.R.* Forst. & G. Forst.

Priyanka Brahma and Sanjib Baruah

Abstract

Glochidion multiloculare (Rottler ex. Willd.) Voigt and G. ellipticum Wight are two medicinally known species of the family Phyllanthaceae. Traditionally all the parts of the plants have been used by many tribes in India for medicinal purposes. Based on the traditional knowledge the current study was conducted to determine the presence of their phytoconstituents with their yield extract quantitatively as well as mineral components present in these two species. Phytochemical analysis has been carried out by following standard methods and the determination of mineral elements has been carried out by atomic absorption spectroscopy (AAS). The result of the phytochemical analysis showed the presence of important phytoconstituents as well as good extraction yield in both species. In both species, the highest concentration of calcium elements has been observed. It shows both plants have nutritional as well as medicinal value.

Keywords: Glochidion multiloculare (Rottler ex. Willd.) Voigt, G. ellipticum Wight, phytochemical, mineral

1. Introduction

Phytochemicals are chemical compounds that are found in the various parts of the plant and these phytoconstituents may be alkaloids, flavonoids, steroids, tannin, terpenoids, phenol, etc. and they possess various types of biological activities such as antioxidants, antimicrobial, antibacterial, cytotoxic activities, etc. These compounds play a major role in protecting plants and humans against harmful diseases [1]. Plants not only contain chemical compounds they are also known for their mineral composition which is beneficial for the daily needs of human beings. Essential elements play an important role in the human body but they can cause harmful disorders if the concentration of the elements is higher beyond the trace limits. The human

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Figure 35. Front page of a published paper



RESEARCH ARTICLES



Extended distribution of an endemic variety *Glochidion zeylanicum* var. *paucicarpum* Chakrab. & N.P. Balakr. (Phyllanthaceae) from Assam, India

Priyanka Brahma¹ • Sanjib Baruah¹

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Abstract

Glochidion zeylanicum var. paucicarpum Chakrab. & N.P. Balakr., an endemic variety of family Phyllanthaceae, was recorded for the first time in Assam. The species was collected while working on the taxonomic study of the genus Glochidion in Assam. After a rational evaluation of published work on distribution, this variety was previously recorded only from Andaman and Nicobar Islands, Bay of Bengal, India and reported as an endemic variety under the species G. zeylanicum (Gaertn.) A.Juss. But the present study revealed that its occurrence in Kokrajhar district set extended distribution records for Assam including northeast India. The present communication deals with thorough taxonomic descriptions, phenology, habitat and ecology, distribution, threat status, and photographs, including a distribution map of the taxa for better taxonomic identity. Although the present study area recorded less population of the plant, the plant has been commonly used as medicine by the local community. Based on the threat status in the present work, the plant can be assigned to an endangered category. The present communication recorded an extended occurrence of the plant in Assam, so further population study of the plant in other regions of India is also needed.

Keywords Glochidion · Phyllantheae · New distribution · Endemic · Assam

Introduction

The genus Glochidion J.R.Forst. & G.Forst., belonging to the tribe Phyllantheae was one of the large genera of the family Phyllanthaceae with about 320 species and distributed mainly in tropical Asia to Northern Australia & Polynesia, with a few species in Madagascar and tropical America (Chakrabarty and Gangopadhyay 1995; Hoffmann et al. 2006; Chakrabarty and Balakrishnan 2018). In India, it comprises of 22 species and 13 varieties (Balakrishnan et al. 2012; Chakrabarty and Balakrishnan 2018). A total of 16 species of the genus were reported by Kanjilal et al. (1940) in "Flora of Assam" from the erstwhile Assam.

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Department of Botany, Bodoland University, 783370 Kokrajhar, Assam, India The name Glochidion was first given by J. R. Forster and G. Forster in 'Characteres Generum Plantarum' (1776) and the generic word "Glochidion" came from the Greek word "Glochis" which signifies the extension of anther connectives of a female flower (Chakrabarty and Gangopadhyay 1995). Members of Glochidion are found predominantly in grasslands, evergreen forests, sal forests, deciduous forests, subtropical forests, hilly places, swampy areas along stream sides, and roadside areas. They are generally shrub or large trees, drooping branched having either glabrous or hairy plant bodies, inflorescences exhibit axillary to supra-axillary with lobed and unlobed capsules. The genus comprises of many species and varieties and has been popularly known for its medicinal value in India (Chakrabarty and Balakrishnan 2018).

While exploring the field survey, a voucher specimen of the member Glochidion was collected. The present communication reported the presence of Glochidion zeylanicum var. paucicarpum Chakrab. & N.P. Balakr. in Assam for the first time. The specimen was distributed in Andaman and Nicobar Island, India and it was reported as an endemic

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Figure 36. Front page of a published paper

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Comparative morphological and ethnobotanical assessment of certain taxa of genus Glochidion (Phyllanthaceae) from Assam, India

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Abstract: The genus Glochidion, a member of the family Phyllanthaceae, primarily comprises shrub or tree species. It stands out from other genera due to distinctive reproductive features, including prolonged styles in flowers and fruits, apiculate anthers, and lobed and unlobed capsules. This study aimed to compare the morphological characteristics of nine taxa in Assam to facilitate identification and assess ethnobotanical knowledge. Ethnobotanical information was gathered by interviewing the local community, and a taxonomic key was provided for accurate identification. Morphological data underwent principal component analysis (PCA) and cluster analysis using PAST for validation. The comparison revealed distinct characteristics in both vegetative and reproductive traits among Glochidion members. Reproductive features, such as inflorescence, style, ovary, and capsules, were key factors for differentiation and identification. PCA and cluster analysis demonstrated correlation and variation among the taxa, contributing significantly to their demartation. Ethnobotanical studies indicated the genus's potential medicinal properties, supported by both primary and secondary information.

Keywords: Angiosperms, cluster analysis, ethnobotany, PCA, Phyllanthoideae, taxonomy, UPGMA.

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Author details: Dr. S. Baruan is working as an assistant professor at the Department of Botany, Bodoland University, Kokrajhar, Assam. SB has 15 years of research and teaching experience on the plant taxonomy and conservation of threatened plants in north-east India. Pervanka Brawma is a bona fide Ph.D. research scholar in the Department of Botany at Bodoland University. PB is pursuing her PhD on the taxonomy and phytochemistry of Glochidlon in Assam.

Author contributions: PB has collected, done photography, identified, carried out the morphological analysis of the specimen, performed the PCA and cluster analysis and drafted the manuscript. SB contributed to the present study's design, supervised the work and revised the manuscript. The final manuscript was examined and approved by both the authors.

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Figure 37. Front page of a published paper





Foliar epidermal micromorphology of genus *Glochidion* J.R.Forst. & G.Forst. (Phyllanthaceae) by using light and electron microscopy

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ABSTRACT

The present study was conducted to compare both qualitative and quantitative characteristics of foliar epidermal micromorphology on some members of Chohidion in Assam. As taxonomic attributes, the foliar epidermal micromorphology study of nine taxa of both abaxial and adaxial surfaces was performed by using light microscopy (LM) and field emission scanning electron microscopy (FESEM). The result showed both amphistomatic and hypostomatic types of leaf surfaces. On the same surface of the leaf, multiple types of stomata were observed such as anomocytic, anisocytic, hemiparacytic, and paracytic types. Significant diversity and variations were observed in stomatal number, size, area, epidermal cell number, subsidiary cells, and trichomes. The stomatal index, stomatal shape, epidermal cell shape, length and width of the stomata, and trichomes showed variation among the studied taxa. Clauds were absent in all studied members. Papillae and epicuticular wax crystals were observed in some taxa. In addition, a taxonomic key was also provided based on foliar leaf epidermal characteristics using qualitative and quantitative data from LM and FESEM. Based on quantitative data for foliar leaf micromorphology, principal component analysis (PCA) and cluster analysis were carried out to authenticate the micromorphological data. These would aid in the identification of taxa as well as in taxonomic delimitation.

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KEYWORDS: Glochidium, Leaf epidermis, Light Microscopy, Field Emission Scanning Electron Microscopy, Principal Component Analysis, Trichome

INTRODUCTION

The genus Glochidion is a large member of the family Phyllanthaceae and the genus consists of c 320 species across the world and c 22 species and 8 varieties in India (Balakrishnan & Chakrabarty, 2007; Balakrishnan et al., 2012; Chakrabarty & Balakrishnan, 2018). A total of 16 species from erstwhile Assam were described in "Flora of Assam" by Kanjilal et al. (1940). Earlier the genus was placed under the family Euphorbiaceae (Bentham & Hooker, 1862-1863; Hooker, 1890; Kanjilal et al., 1940). According to Hoffmann et al. (2006), the genus was closely connected to the type of Phyllanthus as a result of the genus Glochidion, which includes Breynia J.R. & G.Forst., Flueggea Willd., and Margaritaria L.f., were classified under the family Phyllanthaceae and the tribe Phyllantheae. Additionally, a recent classification assigned the genus to the Phyllanthaceae family (The Angiosperm Phylogeny Group et al., 2016). The biovulate ovary and lack of latex distinguish the family Phyllanthaceae from Euphorbiaceae (Chakrabarty & Balakrishnan, 2018).

The members of Glochidion are mainly shrubs or small trees and large trees. They can be found primarily along roadsides, in damp or humid deciduous areas, tropical, primary, and secondary forests, sal woods, mountainous terrain, and occasionally swampy places. The genus can be identified by its drooping branches, axillary and supra-axillary inflorescence, arrangement of male and female flowers, and lobed and unlobed capsules. Taxonomic studies of Glochidion based on morphological characters were conducted in many regions of the world (Robinson, 1909; Beille, 1927; Backer & van den Brink, 1963; Shaw, 1981; Li, 1994; Chakrabarty & Gangopadhyay, 1995; Nguyen, 2007; Li & Gilbert, 2008; Chakrabarty & Balakrishnan, 2018; Yao et al., 2020). However, the micromorphological or leaf epidermal study is still lacking. Only a few species i.e., G. hohenckari and G. neilgherrense of the foliar epidermal study were covered by some workers (Thakur & Patil, 2011, 2014).

Micromorphological traits can often remain seen to be adequately varied to be used as a tool for taxonomy and species identification (Vislobokov et al., 2021). The Epidermal features

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Figure 38. Front page of a published paper





RESEARCH ARTICLE

Investigation of phytochemical constituents, GC-MS, DPPH free radical scavenging assay, and mineral contents of *Glochidion* sphaerogynum (Mull. Arg.) Kurz bark extract

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Abstract

The aim of the present study was to assess phytochemical constituents, chemical composition, DPPH free radical scavenging assay and mineral contents of Glochidion sphaerogynum (Mull.Arg.) Kurz bark extract. Standard procedures were used to test preliminary phytochemical constituents as well as quantitative analysis for total alkaloid, flavonoid, saponin, tannin, phenolic and terpenoid content. The extract was examined using gas chromatography-mass spectrometry (GC-MS) to know the biologically active compound. In-vitro antioxidant potential was investigated using DPPH free radical scavenging assay and the IC₅₀ value for the antioxidant activity of bark extract was 37.4479 µg/mL. The qualitative phytochemical investigation revealed the presence of important phytochemical constituents as well as considerable amounts of total alkaloid, flavonoid, saponin, tannin, phenolic and terpenoid content. GC-MS revealed the presence of biologically active compounds like 1,1,6-Trimethyl-3-methylene-2-(3,6,10,13,14-pentamethyl-3-ethenylpentadec-4-enyl)cyclohexane; Benzenepropanoic acid, 3,5-bis(1,1-dimethyl ethyl) -4-hydroxy-, methyl ester; Neophytadiene etc which would be important for medicinal industries. Mineral contents were determined by using Atomic Absorption Spectrometry (AAS). The result revealed the presence of a good concentration of Na (10.552±0.343 ppm) and Ca (8.973±0.310 ppm) elements followed by K, Fe, Mg and Mn and very less concentrations of heavy metals such as Cd, Cr and Pb indicated the species was devoid of harmful metals.

Keywords

Glochidion sphaerogynum, phytochemical, GC-MS, DPPH, mineral contents

Introduction

The Plant produces many phytochemical constituents that protect them against insects, pathogens, and herbivores (1). Many of these phytochemical constituents also have different biological activities and protect human beings against various diseases (2, 3). According to WHO (World Health Organization), 80% of people rely on indigenous plant-based medicines for curing different diseases (4). Plant produces antioxidants as secondary metabolites, which are substances that act in the cell to counteract free radicals and reactive oxygen species (ROS) and control oxidative damage in the body that become scientifically compelling compounds as a consequence of many pharmacological activities (5, 6). One of the reliable analytical methods for identifying the different chemical components in a plant sample is gas chromatography-mass spectrometry (GC-MS) which plays a significant role in the analysis of phytochemical compounds (7-9).

Glochidion sphaerogynum is a small tree belonging to the genus Glochidion of the family Phyllanthaceae which mainly grows in primary evergreen forests, secondary forests, hilly, or roadside areas, distributed in Bhutan, Nepal, Bangladesh, Myanmar, China, Vietnam, Thailand and states of India (10). This species can be identified by its glabrous plant body, long acuminate leaves, axillary inflorescence and distinguishable lobed capsules from other members of the genus Glochidion. The genus Glochidion contains various chemical constituents that include flavonoid, terpenoid, glycoside, steroid, phenolic compounds, triterpenoid, tannin, saponin, alkaloid, resin, anthraquinone, reducing sugar, phlobatannin and carbohydrate (11). This proves the importance of the genus as a source of medicinal value. The leaves and branches of G. sphaerogynum have been used mainly for the treatment of influenza, skin problems, common cold, fever, ulcers and inflammation in some countries (12-14). However, no detailed examination of the phytochemical composition, GC-MS analysis, antioxidant potential and assessment of mineral contents of this plant has been published so far. So, the present study desired to evaluate phytochemical analysis, identification of biologically active compounds using GC-MS, antioxidant potential using DPPH free radical scavenging assay and assessment of mineral contents in the bark extract of G. sphaerogynum based on their medicinal importance. Hence, the present study was also conducted to know the biologically active components, their mineral composition and the antioxidant capacity present in the bark extract.

Materials and Methods

Collection and identification of the plant material

The field survey was conducted from January to April 2021 and fresh plant specimens were collected in the Kokrajhar district of Assam. The plant material was identified following the literature i.e., Flora of Assam and a voucher specimen (008 BUBH) was deposited in the Bodoland University Botanical Herbarium (BUBH), Kokrajhar, Assam (15, 16).

Preparation of the plant extracts

After identification of the specimen fresh bark material was collected. The collected bark was properly washed with distilled water and dried in the shade for one and half months until it becomes completely moisture free. After drying, it was ground to a coarse powder and kept in an airtight container. Twenty gms of the powdered sample was taken and soaked in 200 mL of methanol for 72 hrs and filtered through Whatman filter paper no. 1. Then the filtrate extract was evaporated with a vacuum rotary evaporator (Ikon instruments) and set the temperature according to the boiling point of the solvent. After evaporation residue was taken and stored at 4 °C for future analysis of the sample (17, 18).

Phytochemical screening

Alkaloid, reducing sugar, steroid, phlobatannin, tannin, flavonoid, terpenoid, triterpenoid, saponin, glycoside and phenol were tested according to standard methods (18-21).

Quantitative estimation of phytochemical constituents

Total alkaloid content

2.50 g of powder bark sample was measured, and 200 mL of 10% acetic acid (CH $_3$ COOH) in ethanol (C $_2$ H $_5$ OH) was added, followed by 4 hrs of rest. The filtrate was then heated in a water bath after filtration. Until the precipitation was complete, concentrated ammonium hydroxide (NH $_4$ OH) was applied. It was then washed with 20 mL of 0.1M ammonium hydroxide (NH $_4$ OH) and filtered using Whatman filter paper no. 1. The residue was dried in an oven before being weighed on an electronic balance. The total alkaloid content can be calculated using the following formula-

Alkaloid (%) = Final weight of the residue/Initial weight of sample $\times 100$ (18, 19).

Total flavonoid content

2.50 g of powder bark sample was mixed with 50 mL of 80% aqueous methanol (CH $_3$ OH) in a 250 mL beaker, it was covered and kept at 20 °C for 24 hrs. The supernatant was discarded, and the residue was extracted 3 times more, each time with the same volume of ethanol (C_2H_5 OH). Then, using Whatman filter paper no. 1, the entire solution was filtered. The filtrate was dried and weighed in an electronic balance after being evaporated over a water bath. Total flavonoid content can be done as-

Flavonoid (%) = Final weight of the residue/Initial weight of sample $\times 100$ (18, 19).

Total saponin content

5 g of powdered bark material was put into a 250 mL conical flask, followed by 100 mL of 20% aqueous ethanol (C_2H_5OH), and heated in a hot water bath for 4 hrs at 55 °C. The procedure was carried out twice more. The mixed extract was then reduced to 40 mL in a water bath at 90 °C. The mixed extract was placed in a separating funnel and rapidly shaken with 20 mL diethyl ether $\{(C_2H_5)_2O\}$. The ether layer was then removed. The purifying process was repeated 2 times. After that, 60 mL of n-butanol ($C_4H_{10}O$) was added, followed by 2 times washing with 10 mL of 5% sodium chloride (NaCl). To obtain a final concentration of saponin, the sodium chloride layer was removed and the residual solution was heated in a water bath and dried at 50 °C in an oven. Total saponin content can be done as-

Saponin (%) = Final weight of the residue/Initial weight of sample ×100 (18, 19).

Total phenolic content

The total phenolic content of the methanolic bark extract was determined following the Folin Ciocalteu's (FC) method using UV-Vis Spectrophotometer (Shimadzu A 125358) with slight modification (22, 23). 0.01 g sample was weighed and made up to 1800 μ L of distilled water. Then 150 μ L of Folin Ciocalteu reagent and 1 mL of 10% sodium carbonate (Na₂CO₃) were added and made up to 3 mL of volume and incubated in the absence of light for 40 min. Absorbance was measured at 765 nm against a blank and the calibration curve was created using the standard gallic acid in the range of 10-50 μ g/mL. Based on the average of three experiments, the phenolic content of bark extract

was quantified as mg of gallic acid equivalents of dried extract (mg GAE/g dry extract).

Total tannin content

The tannin content of the bark extract was quantified using the Folin Denis (FD) method in UV-Vis Spectrophotometer (Shimadzu A 125358) with slight modification (24-27). 0.01 g of the sample was weighed and made up of 1800 μ L of distilled water. After that, 150 mL FD reagent and 1 mL sodium carbonate (Na₂CO₃) were added to make a total volume of 3 mL, which was then incubated in the absence of light for 40 min. The calibration curve was built using tannic acid as a standard (10-50 g/mL) and calculated at the absorbance of 760 nm versus a blank. Based on the average of 3 experiments the total tannin content was quantified as mg of tannic acid equivalents of dried extract (mg TAE/g dry extract).

Total Terpenoid content

Total terpenoid content was determined as described by the given method (28, 29). For 24 hrs, 2 g of the material was dissolved with 50 ml of ethanol (C_2H_5OH). The mixture was then filtered using Whatman No. 1 filter paper. The filtrate was collected and extracted with petroleum ether (C_6H_{14}) before being dried at 65 °C in a water bath. Then the residue was collected and calculated the final volume. Total terpenoid content can be determined as-

Terpenoid (%)
$$= \frac{\text{Final weight of residue}}{\text{Initial weight of sample}} \times 100$$

GC-MS Analysis

GC-MS analysis of *G. sphaerogynum* (Mull.Arg.) Kurz bark extract was performed with a Perkin Elmer (USA) Clarus 680 GC and amp. Turbo Mass Ver. 6.1.2 instrument. The system was programmed using the software, and the peaks were examined with the NIST-2014 software. The stationary phase was 5% diphenyl 95% dimethyl polysiloxane, while the carrier gas was helium gas (99.99 %) at a flow rate of 1mL/min. A 2 μ L injection volume was used in splitless mode. The injector has a temperature of 280 °C, whereas the ion source has a temperature of 180 °C. Electron Impact positive (EI+) mode was used to obtain Mass Spectra at 70 eV. The mass spectrum of peaks and compounds was determined using NIST-2014 software and a library search (30).

DPPH free radical scavenging assay

The antioxidant activity of a methanolic extract of G. sphaerogynum bark was tested using the 2,2-Diphenyl-1-picryl-hydrazyl-hydrate (DPPH) assay with slight modification (31-34). In the absence of light, 1 mL methanolic extract solutions (10-60 μ g/mL) were dissolved in 3 mL DPPH solution and incubated. 3 mL methanol was used to make the blank, and 2 mL methanol and 1 mL DPPH solution were used to make the control. With a UV-Vis spectrophotometer, the absorbance of extracts was measured at 517 nm after 30 min and compared to that of normal ascorbic acid at equal amounts. The ability of the DPPH radical scavenging assay was calculated using the following formula-

% of inhibition = (Absorbance of control - Absorbance of the sample extract or standard)/Absorbance of control \times 100

The IC_{50} value was obtained using linear regression analysis. If the IC_{50} value is lower, the antioxidant activity is stronger.

Assessment of mineral contents

Atomic Absorption Spectrometry (AAS), Model: Shimadzu AA-7000, was used to determine Sodium (Na), Potassium (K), Calcium (Ca), Magnesium (Mg), Iron (Fe), Manganese (Mn), Cadmium (Cd), Chromium (Cr) and Lead (Pb) at CIF IASST, Guwahati, Assam. The material was digested with concentrated nitric acid (HNO₃) and hydrogen peroxide (H₂O₂) using the wet ashing process. 0.50 g of sample was mixed with 8 mL of HNO₃ and let to stand overnight. The following day, it was heated for 1 hr at 120 °C on a hot plate with 4 mL H₂O₂ until the digestion was colorless. At 80 °C, the residue was dried and diluted with hydrochloric acid (HCl) and filtered. The results of the AAS were determined as ppm concentration based on the average of 3 readings in AAS (35).

Statistical Analysis

All the tests were done in 3 replicates. The data was determined as an average of 3 experiments (n=3) and calculated mean \pm standard deviation (SD) using Microsoft Excel. The statistical analysis was performed by ANOVA single factor at $p \le 0.05$ level in Microsoft Excel.

Results

Phytochemical screening

The presence of significant secondary metabolites such as alkaloid, steroid, phlobatannin, tannin, flavonoid, terpenoid, triterpenoid, saponin, glycoside and phenol was confirmed in the methanolic bark extract of *G. sphaerogynum* (Table 1).

 $\textbf{Table 1.} \ \ \textbf{Preliminary phytochemical screening of methanolic bark extract of } \textit{G. sphaerogynum}$

Phytochemical constituents	Test/Reagent	Result
Alkaloid	Dragendorff's rea- gent	+
	Mayer's reagent	+
	Wagner's reagent	+
Reducing sugar	Fehling's test	-
Steroid	Salkowski test	+
Phlobatannin	HCl test	+
Tannin	FeCl₃test	+
Flavonoid	FeCl₃test	+
	H ₂ SO ₄ test	-
Terpenoid	Salkowski test	+
Triterpenoid	H ₂ SO ₄ test	+
Saponin	Foam test	+
	Keller-Killiani test	-
Glycoside	Borntrager's test	+
Phenol	FeCl₃test	+

Positive sign (+) = Present; Negative sign (-) = Absent.

Quantitative estimation of phytochemical constituents

According to the result of quantitative phytochemical analysis, total alkaloid and flavonoid contents obtained 3.73% (0.093±0.025) and 1.24% (0.031±0.019) yield respectively. The total saponin content exhibited a 12.94% (0.646±0.473) yield. Total tannin and phenolic contents were 6.92112 mg TAE/g dry extract (0.744±0.062) and 0.1666 mg GAE/g dry extract (0.328±0.045) respectively. The total terpenoid content obtained was 8.66% (0.173±0.066) yield (Table 2).

Table 2. The total alkaloid, flavonoid, saponin, phenolic, tannin and terpenoid contents in methanolic bark extract of *G. sphaerogynum*

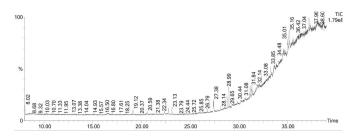
Parameters	Result
Total alkaloid content (% yield) Total flavonoid content (% yield) Total saponin content (% yield) Total phenolic content (mg GAE/g) Total tannin content (mg TAE/g) Total terpenoid content (% yield)	0.093±0.025 0.031±0.019 0.646±0.473 0.328±0.045 0.744±0.062 0.173±0.066

Sample was analyzed in three replicates and data were indicated as an average of three tests (n=3), mean \pm standard deviation (SD), and they were significantly different from each other at $p \le 0.05$ level (ANOVA single test).

GC-MS Analysis

The bioactive compounds present in methanolic bark extracts of the *G. sphaerogynum* were shown in Fig. 1. The retention time, molecular weight, molecular formula and % of peak area were identified using the NIST library and presented in Table 3. The important bioactive compound present in the methanolic bark extract were Hentriacontane, Xanthosine, Fluorene, Phenanthrene, Neophytadiene, Benzenepropanoic acid,3,5-bis(1,1-dimethyl ethyl)-4-hydroxymethyl ester, 2,6,10,14-Tetramethyl-7-(3-methylpent-4-enylidene)pentadecane, Z,Z-6,28-Heptatriactontadien-2-one, 1,1,6-Trimethyl-3-methylene-2-(3,6,10,13,14-pentamethyl-3-ethenyl-pentadec-4-enyl)cyclohexane, Methyl 2-hydroxyeicosanoate.

Fig. 1. GC-MS chromatogram of methanolic bark extract of G. sphaerogynum



DPPH free radical scavenging assay

The antioxidant potential of bark extract was evaluated using the DPPH assay. DPPH free radical scavenging assay of concentrations (10-60 μ g/mL) and percentage inhibition of bark extract and the standard ascorbic acid were presented in Table 4. Radical scavenging assay at the concentration of 60 μ g/mL, bark extract and ascorbic acid showed inhibition of 58.503±0.095% and 71.055±0.064% respectively. The IC₅₀ value of bark extract and standard ascorbic acid were 37.4479 μ g/mL and 20.9167 μ g/mL respectively that indicating a good amount of antioxidant potential.

Assessment of mineral contents

The result of the mineral analysis of *G. sphaerogynum* bark extract was enlisted (Table 5). The result showed that sodium (Na) had the highest mineral contents of 10.552±0.343 ppm followed by calcium (Ca) and potassium (K) contents of 8.972±0.310 ppm and 7.230±0.086 ppm respectively. Sodium, calcium and potassium are one of the most important minerals in the body. K and Na help to maintain ionic balance, which helps to prevent hypertension and improve blood pressure (36, 37). Ca helps to maintain teeth, bone, blood clotting and muscle to contract and regulate normal heart rhythms and nerve function (38).

Discussion

The results of preliminary qualitative phytochemical screening of the methanolic extract of G. sphaerogynum bark showed the presence of important phytochemical constituents such as alkaloid, flavonoid, steroid, phlobatannin, tannin, terpenoid, triterpenoid, saponin, glycoside and phenol that would be implied on the identification of new sources of plants as pharmaceutical applications. The quantitative phytochemical estimation revealed a high % yield of saponin content (12.94%) followed by terpenoid content (8.66%) and a good concentration of tannin content of 6.92112 mg TAE/g dry extract (0.744±0.062) and phenolic content of 0.1666 mg GAE/g dry extract (0.328±0.045) respectively. In the present study, a good amount of alkaloids and flavonoids also have been recorded. Important biological activities such as antioxidant, anti-inflammatory, antibacterial and antimicrobial activities are known to exist in terpenoids, phenols, flavonoids, saponins, tannins and alkaloids (39).

molecule 1,1,6-Trimethyl-3-methylene-2-(3,6,10,13,14-pentamethyl-3-ethenyl-pentamethyl-3ethenyl-pentadec-4-enyl) cyclohexane which is the primary compound identified as having pharmacological activities such as antimicrobial, anti-inflammatory, anticancer, antiarthritic, antiviral with the highest peak area of 2.263% followed by the compound Benzenepropanoic acid, 3,5-bis (1,1-dimethyl ethyl)-4-hydroxy-, methyl ester with the peak area of 1.091% that has antioxidant and antifungal properties found during the GC-MS analysis. From the GC-MS data, all the identified compounds possess important biologically active compounds such as antioxidants, antitumor, antimicrobial, anti-inflammatory, antifungal, cytotoxic and various pharmacological activities which creates a basis for defining the plant's potential benefits in medicinal industries (Table 3).

The DPPH free radical scavenging assay was used to examine antioxidant activity. The result showed that the concentrations of methanolic bark extracts had significant DPPH free radical scavenging effects. The highest % inhibition (59.512±0.557) showed in 50 µg/mL followed by 58.503±0.095 % inhibition in 60 µg/mL with a potent antioxidant amount of IC50 value. Further assays are required to know the significance of the antioxidant capacities of the extract.

Table 3. Identified chemical composition of methanolic bark extract of G. sphaerogynum using GC-MS and their biological activity

Sl. No.	Retention time	Compound name	Peak area %	Molecular weight	Molecular for- mula	Biological activity
1	19.123	Hentriacontane	0.249	436	C ₃₁ H ₆₄	Anti-inflammatory, antitumor and antimi- crobial activities (44)
2	20.588	Xanthosine	0.247	284	$C_{10}H_{12}N_4O_6$	Therapeutic and pharmacological property (45)
3	23.139	Fluorene	0.473	166	$C_{13}H_{10}$	Drug design (46)
4	27.381	Phenanthrene	0.608	178	$C_{14}H_{10}$	Analgesic, antitussive, antimalarial, cyto- toxic, anti-constipation, antioxidant, anti- inflammatory activity (47)
5	28.141	Neophytadiene	0.229	530	$C_{20}H_{38}$	Anti-inflammatory, antipyretic, analgesic, antioxidant, antimicrobial, antifungal, antibacterial activity (48-52)
6	28.987	Benzenepropanoic acid, 3,5-bis (1,1-dimethyl ethyl)-4-hydroxy-, methyl ester	1.091	292	$C_{18}H_{28}O_3$	Antioxidant, antifungal (53-56)
7	31.848	2,6,10,14-Tetramethyl-7-(3- methylpent-4-enylidene) penta- decane	0.468	348	C ₂₅ H ₄₈	Anti-inflammatory, Antioxidant (57)
8	33.849	Z,Z-6,28-Heptatriactontadien-2- one	0.751	530	C ₃₇ H ₇₀ O	Vasodilatory, carcinogenic, antioxidant activity (58, 59)
9	35.174	1,1,6-Trimethyl-3-methylene-2- (3,6,10,13,14-pentamethyl-3- ethenyl-pentadec-4-enyl) cyclohexane	2.263	442	C ₃₂ H ₅₈	Pharmacological activity, antimicrobial, anticancer, antiarthritic, anti- inflammatory and antiviral properties (60, 61)
10	37.955	Methyl 2-hydroxy-eicosanoate	0.275	342	$C_{21}H_{42}O_3$	Pharmaceutical property, antioxidant, anti-inflammatory (62, 63)

Table 4. DPPH assay of methanolic bark extract of G. sphaerogynum

Concentration (µg/mL)	% inhibition				
	Bark extract	Ascorbic acid			
10	38.272±1.165	41.767±0.641			
20	44.080±0.586	59.143±0.109			
30	42.013±1.190 56.091±0.461				
40	51.070±0.794 62.072±0.346				
50	59.512±0.557	65.985±0.031			
60	58.503±0.095	71.055±0.064			
$IC_{50}(\mu g/mL)$	37.4479	20.9167			

The sample was analyzed in three replicates and data were indicated as an average of three tests (n=3), mean \pm standard deviation (SD) and they were significantly different from each other at $p \le 0.05$ level (ANOVA single test)

In order to be healthy, the human body requires a variety of minerals found in different varieties of plants. Apart from this, plants possess minerals that are beneficial for humans and some toxic metals which are harmful to human health. The presence of heavy metal toxicity in plants may create severe problems. The determination of mineral composition showed the highest concentration in sodium (Na) elements followed by calcium (Ca), potassium (K), iron (Fe), magnesium (Mg), manganese (Mn), and less concentration of chromium (Cr), lead (Pb) and cadmium (Cd). Na maintains the osmotic pressure in the body and activates nerves and muscle functions (40). K and Na maintain the ionic equilibrium that helps to prevent hypertension and enhances blood pressure (36, 37). Mg is important for enzyme activation and bone formation and Ca plays an important role as a constituent of bones and teeth formations, regulation of nerves and muscles functions in living cells and helps in membrane formulating (40). Fe is

Table 5. Mineral assessment of bark of G. sphaerogynum

Minerals	Result (ppm)
Sodium (Na) Potassium (K) Calcium (Ca) Magnesium (Mg) Iron (Fe) Manganese (Mn) Chromium (Cr) Lead (Pb) Cadmium (Cd)	10.552±0.343 7.230±0.086 8.973±0.310 3.379±0.167 4.026±0.076 3.367±0.179 0.162±0.011 0.162±0.016 0.009±0.002

The sample was analyzed in three replicates and data were indicated as an average of three tests (n=3), mean \pm standard deviation (SD), and they were significantly different from each other at $p \le 0.05$ level (ANOVA single test)

linked to haemoglobin and oxygen transmission from the lungs to tissue cells. Mn activates enzymes and is involved in urea formation, haemoglobin formation, helping in the nervous system and normal bone growth (37, 41, 42). Cd, Pb and Cr are non-essential elements in both plants and humans if it consists of high concentration. They are responsible for toxic substances (43). Plants are a good source of phytochemicals and minerals that can be useful for the nutraceutical industry. Several investigations have found elemental content in plant extracts that we consume as herbal health supplements or medicine and biochemical processes in the human body are affected by macro and trace elements (36, 37, 41). Therefore, it was evident from the results that the plants would be free of harmful materials because of the very low concentration of Cr, Pb, and Cd and could be important mineral consuming plants due to the presence of the important mineral elements.

Conclusion

The study revealed that plant has a potential source of phytochemical constituents that would be beneficial to humankind. The GC-MS analysis justifies the presence of important biological and pharmacological properties such as antifungal, antibacterial, antioxidant, antimicrobial, anti-inflammatory, antimalarial, analgesic and cytotoxic activities). This study can serve as a basis for additional research on the biological activities of the plant. The DPPH assay of bark extract exhibited a good amount of antioxidant compound which could be useful in the pharmaceutical and medicinal industry. Mineral determination indicated the presence of a significant number of mineral factors such as sodium (Na), calcium (Ca), potassium (K), iron (Fe), magnesium (Mg), manganese (Mn) and the plant could be free from toxic substances due to less concentration of heavy metal such as Cadmium (Cd), Chromium (Cr) and Lead (Pb). This is the first report citing mineral contents and detailed phytochemical evaluation, GC-MS and antioxidant analysis of the G. sphaerogynum. Further studies are needed to isolate biologically active compounds as important in pharmacological industries.

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Authors contributions

PB collected and identified the specimen, analyzed the data, performed the statistical analysis and drafted the manuscript. SB participated in the design of the study, supervised the work and revised the manuscript. Both authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interest to declare.

Ethical issues: None.

References

- Taiz L, Zeiger E. Plant Physiology. 3rd ed. Sunderland: Sinauer Associates; 2002.
- Ghasemzadeh A, Jaafar HZ, Rahmat A. Phytochemical constituents and biological activities of different extracts of Strobilanthes crispus (L.) Bremek leaves grown in different locations of Malaysia. BMC Complement Altern Med. 2015; 15 (1):422_https://doi.org/10.1186/s12906-015-0873-3
- 3. Khan IH, Javaid A. Anticancer, antimicrobial and antioxidant compounds of quinoa inflorescence. Adv Life Sci. 2020; 8 (1):68-72.
- WHO, IUCN, WWF. Guidelines on the conservation of medicinal plants. Switzerland; 1993.
- 5. Zehiroglu C, Ozturk Sarikaya SB. The importance of antioxidants and place in today's scientific and technological studies. J Food Sci Technol. 2019; 56(11):4757-74. https://doi.org/10.1007/s13197-019-03952-x
- Khan IH, Javaid A. Antifungal, antibacterial and antioxidant components of ethyl acetate extract of quinoa stem. Plant Prot. 2019; 3(3):125-30. http://dx.doi.org/10.33804/ pp.003.03.3109
- Khan IH, Javaid A, Ahmed D, Khan U. Identification of volatile constituents of ethyl acetate fraction of *Chenopodium quinoa* roots extract by GC-MS. Int J Biol Biotechnol. 2020; 17(1):17-21.
- 8. Olivia NU, Goodness UC, Obinna OM. Phytochemical profiling and GC-MS analysis of aqueous methanol fraction of *Hibiscus asper* leaves. Future J Pharm Sci. 2021;**7**:59. https://doi.org/10.1186/s43094-021-00208-4
- Ferdosi MFH, Khan IH, Javaid A, Fardosi MFA. GC-MS examination of methanolic extract of *Cirsium arvense* flower. Pak J Weed Sci Res. 2021;27(2):173-80. https://doi.org/10.28941/pjwsr.v27i2.946
- Chakrabarty T, Balakrishnan NP. Indo-Burmese Phyllanthaceae: A Taxonomic Revision. Dehradun, India; 2018.
- Sandhya S, Chaintanya RSNAKK, Vinod KR, Rao KNV, David B, Sudhakar K, Swetha R. An updated review on the Genus Glochidion Plant. Arch Appl Sci Res. 2010;2(2):309-22.
- 12. Bajpai O, Kumar A, Srivastava AK, Kuhwaha AK, Pandey J, Chaudhary LB. Tree species of Himalayan Terai region of Uttar Pradesh, India: a checklist. Check List. 2015;11(4):1-15. https://doi.org/10.15560/11.4.1718
- 13. Lai XZ, Yang YB, Shan XL. The investigation of Euphorbiaceous medicinal plants in Southern China. Econ Bot. 2004; 58:S307-S320. https://doi.org/10.1663/0013-0001(2004)58 [S307:TIOEMP]2.0.CO;2
- 14. Lalrinkimi, Lallianthanga RK. Documentation of tree species within Mizoram Science Centre, Berawtlang, Aizawl, India with notes on their ethnomedicinal values. Sci Vision. 2019;19(3):63-78. http://dx.doi.org/10.33493/scivis.19.03.01
- 15. Kanjilal UN, Kanjilal PC, Dey RN, Das A. Flora of Assam. vol-4. Calcutta; 1940.
- 16. Jain SK, Rao RR. A Handbook of Field and Herbarium Methods. New Delhi: India; 1977.
- 17. Khan IH, Javaid A. Antifungal activity of leaf extract of *Cannabis* sativa against Aspergillus flavipes. Pak J Weed Sci Res 2020; 26 (4):447-53. http://dx.doi.org/10.28941/pjwsr.v26i4.883
- Harborne JB. Phytochemical Methods, A Guide to Modern Techniques of Plant Analysis. London; 1973. https://doi.org/10.1007/978-94-009-5921-7_1
- Ogunjobi KM, Abdulwahab SO, Gakenou OF, Thompson OE, Olorunfemi O. Qualitative and quantitative evaluation of the phytochemical constituents of three wood species in Ogun state, Nigeria. Trop Plant Res. 2020; 7(3):627-33. http:// dx.doi.org/10.22271/tpr.2020.v7.i3.078
- Raaman N. Phytochemical Techniques. New Delhi: India; 2006.
- 21. Tripathi IP, Mishra C. Phytochemical screening of some medicinal plants of Chitrakoot region. Indian J Appl Res. 2015;5 (12):56-60. https://doi.org/10.3126/sw.v12i12.13598
- 22. Chethana KR, Sasidhar BS, Naika M, Keri RS. Phytochemical

- composition of *Caesalpinia crista* extract as potential source for inhibiting cholinesterase and **β**-amyloid aggregation: Significance to Alzheimer's disease. Asian Pac J Trop Biomed. 2018; 8(10):500-12. https://doi.org/4103/2221-1691.244159
- Jamuna S, Subramaniam P, Krishnamoorthy K. Phytochemical analysis and evaluation of leaf and root parts of the medicinal herb, *Hypochaeris radicata* L. for *in vitro* antioxidant activities. Asian Pac J Trop Biomed. 2014; 4(Suppl1):S359-S367. https://doi.org/10.12980/apjtb.4.2014c1030
- 24. Padma R, Parvathy NG, Renjith V, Kalpana PR. Quantitative estimation of tannins, phenols and antioxidant activity of methanolic extract of *Imperata cylindrica*. Int J Res Pharm Sci. 2013;4(1):73-77.
- Polshettiwar SA, Ganjiwale RO. Spectrophotometric estimation of total tannins in some ayurvedic eye drops. Indian J Pharm Sci. 2007; 69(4):574-76. http://dx.doi.org/10.4103/0250-474X.36949
- Saxena V, Mishra G, Saxena A, Kamlesh KR, Vishwakarma. A comparative study on quantitative estimation of tannins in *Terminalia chebula*, *Terminalia belerica*, *Terminalia arjuna* and *Saraca indica* using spectrophotometer. Asian J Pharm Clin Res. 2013; 6(7):148-49.
- 27. Soni V, Jha AK, Dwivedi J, Soni P. Qualitative and quantitative determination of phytoconstituents in some antifertility herbs. Indian J Pharm Sci. 2018; 80(1):79-84. http://dx.doi.org/10.4172/pharmaceutical-sciences.1000332
- 28. Ferguson NM. A textbook of pharmacognosy. New York: Max Millam Company; 1956.
- Kong HS, Musa KH, Mohd-Kasim Z, Abdullah Sani N. Qualitative and quantitative phytochemical analysis and antioxidant properties of leaves and stems of *Clinacanthus nutans* (Burm. f.) Lindau from two herbal farms of Negeri Sembilan, Malaysia. ASM Sc. J. 2019;12:1-13. http://dx.doi.org/10.32802/asmscj.2019.87
- 30. Casuga FP, Castillo AL, Corpuz MJAT. GC-MS analysis of bioactive compounds in different extracts of an endemic plant *Broussonetia luzonica* (Blanco) (Moraceae) leaves. Asian Pac J Trop Biomed. 2016;6(11):957-61. https://doi.org/10.1016/ j.apjtb.2016.08.015
- 31. Islary A, Sarmah J, Basumatary S. Proximate composition, mineral content, phytochemical analysis and *in vitro* antioxidant activities of a wild edible fruit (*Grewia sapida* Roxb. ex DC.) found in Assam of North-East India. J Investig Biochem. 2016;5(1):21-31. http://dx.doi.org/10.5455/jib.20160422015354
- 32. Islary A, Sarmah J, Basumatary S. Nutritional properties, phytochemicals and *in vitro* antioxidant assessment of two wild edible fruits from Assam of North-East India. J Pharm Nutr Sci. 2017; 7(2):55-63. http://dx.doi.org/10.6000/1927-5951.2017.07.02.4
- 33. Narzary H, Islary A, Basumatary S. Phytochemicals and antioxidant properties of eleven wild edible plants from Assam, India. Mediterr J Nutr Metab. 2016; 9(3):191-201. http://dx.doi.org/10.3233/MNM-16116
- 34. Shukla RK, Painuly D, Porval A, Shukla A. Proximate analysis, nutritional value, phytochemical evaluation, and biological activity of *Litchi chinensis* Sonn. leaves. J Herbs Spices Med Plants. 2014; 20:196-208, http://dx.doi.org/10.1080/10496475.2013.848830
- 35. Pequerul A, Perez C, Madero P, Val J, Monge E. A rapid wet digestion method for plant analysis. 1993. http://dx.doi.org/10.1007/978-94-017-2496-8_1
- 36. Radha, Kumar M, Puri S, Pundir A, Bangar SP, Changan S et al. Evaluation of nutritional, phytochemical and mineral composition of selected medicinal plants for therapeutic uses from cold desert of Western Himalaya. Plants (Basel). 2021;10(7):1429. https://doi.org/10.3390/plants10071429
- 37. Tapan Seal. Determination of nutritive value, mineral contents and antioxidant activity of some wild edible plants from Meghalaya State, India. Asian J Applied Sci. 2011;4(3):238-46. https://dx.doi.org/10.3923/ajaps.2011.238.246
- 38. Thomas RA, Krishnakumari S. Proximate analysis and mineral composition of *Myristica fragrans* seeds. J Pharmacogn Phytochem. 2015;3(6):39-42.
- 39. Kabir S, Zahan R, Chowdhury AMS, Rashid MA, Haque MR,

- Hasan CM. Evaluation of antioxidant, antitumor, analgesic and anti-inflammatory activities of *Glochidion lanceolarium* (Roxb.) Voigt. J Sci Found. 2020;18(2):72-80. https://doi.org/10.3329/jsf.v18i2.52781
- 40. Paul SH, Usman AA, Gana IN, Manase A, Adeniyi OD, Olutoye MA. Comparative study of mineral and nutritional composition of a multifunctional flora composite formulated from seven medicinal plants and their applications to human health. Eng Technol Open Acc. 2018; 1(5):555572. https://doi.org/10.19080/etoaj.2018.01.555572
- 41. Saupi N, Zakaria MH, Bujang JS. Analytic chemical composition and mineral content of yellow velvet leaf (*Limnocharis flava* L. Buchenau)'s edible parts. J Appl Sci. 2009;9:2969-74. https://dx.doi.org/10.3923/jas.2009.2969.2974
- 42. Liang P, Sang H, Sun Z. Cloud point extraction and graphite furnace atomic absorption spectrometry determination of manganese (II) and iron (III) in water samples. J Colloid Interface Sci. 2006;304:486-90. https://doi.org/10.1016/j.jcis.2006.09.006
- 43. Behera B, Bhattacharya S. The importance of assessing heavy metals in medicinal herbs: A quantitative study. Tang [Humanitas Medicine]. 2016;6(1):3.1-3.4. https://doi.org/10.5667/tang.2015.0029
- 44. Khajuria V, Gupta S, Sharma N, Kumar A, Lone NA, Khullar M et al. Anti-inflammatory potential of hentriacontane in LPS stimulated RAW 264.7 cells and mice model. Biomed Pharmacother. 2017;92:175-86. https://doi.org/10.1016/j.biopha.2017.05.063
- 45. Kulikowska E, Kierdaszuk B, Shugar D. Xanthine, xanthosine and its nucleotides: solution structures of neutral and ionic forms and relevance to substrate properties in various enzyme systems and metabolic pathways. Acta Biochim Pol. 2004; 51(2):493-531. http://dx.doi.org/10.18388/abp.2004_3587
- Gillis EP, Eastman KJ, Hill MD, Donnelly DJ, Meanwell NA. Applications of Fluorine in Medicinal Chemistry. J Med Chem. 2015; 58 (21):8315-59. https://doi.org/10.1021/acs.jmedchem.5b00258
- 47. Kwofie MA, Gupta M. Phenanthrene: A versatile molecule; A review. Plant Archives. 2021; 21(1). http://dx.doi.org/10.51470/PLANTARCHIVES.2021.v21.no1.051
- 48. Adnan M, Nazim Uddin Chy M, Mostafa Kamal ATM, Azad MOK, Paul A, Uddin SB et al. Investigation of the biological activities and characterization of bioactive constituents of *Ophiorrhiza rugosa* var. *prostrata* (D.Don) & Mondal leaves through *in vivo*, *in vitro* and *in silico* approaches. Molecules. 2019; 24(7):1367. https://doi.org/10.3390/molecules24071367
- Ceyhan-Güvensen N, Keskin D. Chemical content and antimicrobial properties of three different extracts of *Mentha pulegium* leaves from Mugla Region, Turkey. J Environ Biol. 2016;37(6):1341-46.
- Singh R, Dar SA, Sharma P. Antibacterial activity and toxicological evaluation of semi purified hexane extract of *Urtica dioica* leaves. Res J Med Plant. 2012;6:123-35. https://doi.org/10.3923/RJMP.2012.123.135
- 51. Venkata Raman B, La S, Saradhi PM, Rao N, Krishna NV, Sudhakar M, Tm R. Antibacterial, antioxidant activity and GC-MS analysis of *Eupatorium odoratum*. Asian J Pharm Clin Res. 2012;5(2):99-106.
- 52. Ozie Akbar Pratama, Woro Anindito Sri Tunjung, Sutikno Sutikno, Budi Setiadi Daryono. Bioactive compound profile of melon leaf extract (*Cucumis melo* L. 'Hikapel') infected by downy mildew. Biodiversitas. 2019;20(11):3448-53. https://doi.org/10.13057/biodiv/d201143
- Gogoi D, Bora G, Borgohain R, G Handique J. Antioxidant capacity and GC-MS analysis of hexane, ethylacetate and methanol extracts of *Ficus bhotanica* A potential folklore medicinal plant. Int J Pharmacogn Phytochem Res. 2018;10 (5):201-12. https://doi.org/10.25258/phyto.10.5.5
- 54. Akpuaka A, Ekwenchi MM, Dashak DA, Ahmed D. Biological activities of characterized isolates of n-hexane extract of *Azadirachta indica* A. Juss (Neem) leaves. N Y Sci J. 2013;6 (6):119-24. http://www.sciencepub.net/newyork
- 55. Kumar V, Sharma A, Thukral A, Bhardwaj R. Phytochemical profiling of methanolic extracts of medicinal plants using GC-

- MS. Phytochemistry. 2016;5(3):2153-58.
- Bashir A, Khan I, Bashir S, Azam S. Chemical composition and antifungal, phytotoxic, brine shrimp cytotoxicity, insecticidal and antibacterial activities of the essential oils of *Acacia* modesta. J Med Plants Res. 2012;6(31):4653-59. http:// dx.doi.org/10.5897/JMPR12.016
- 57. Marchioni I, Najar B, Ruffoni B, Copetta A, Pistelli L, Pistelli L. Bioactive compounds and aroma profile of some Lamiaceae edible flowers. Plants (Basel). 2020;9(6):691. https://doi.org/10.3390/plants9060691
- 58. Mallikadevi T, Paulsamy S, Jamuna S, Karthika K. Analysis for phytoceuticals and bioinformatics approach for the evaluation of therapeutic properties of whole plant methanolic extract of Mukia made- raspatana—A traditional medicinal plant in western districts of Tamil Nadu. India. Asian J Pharm Clin Res. 2012;5(4):163-68.
- 59. Ralte L, Khiangte L, Thangjam NM, Kumar A, Singh YT. GC–MS and molecular docking analyses of phytochemicals from the underutilized plant, *Parkia timoriana* revealed candidate anti-cancerous and anti-inflammatory agents. Sci Rep. 2022;12:3395. https://doi.org/10.1038/s41598-022-07320-2

- 60. Painuli S, Rai N, Kumar N. GC-MS analysis of methanolic extract of leaves of *Rhododendron arboreum*. Asian J Pharm Clin Res. 2016;9(1):101-04.
- 61. Rawat P, Bachheti RK, Kumar N, Rai N. Phytochemical analysis and evaluation of *in vitro* immunomodulatory activity of *Rhododendron arboreum* leaves. Asian J Pharm Clin Res. 2018;11(8):123-28. https://doi.org/10.22159/ajpcr.2018.v11i8.25372
- 62. Ponnudurai G, Peter Paul J. GC-MS analysis of methanolic extract of *Colpomenia sinuosa* (Mertens ex Roth) Derb. Et Sol. from Manapad in the South East Coast of Tamil Nadu, India. Asian J Pharm Res Dev. 2020;8(4):41-43. https://doi.org/10.22270/ajprd.v8i4.761
- 63. Das M, Malipeddi H. Phytochemical screening, GC-MS analysis and biological activity of *Ipomoea eriocarpa* leaf extracts. Int J Pharm Pharm Sci. 2014;6(4):592-94.

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Comparative morphological and ethnobotanical assessment of certain taxa of genus *Glochidion* (Phyllanthaceae) from Assam, India

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Abstract: The genus *Glochidion*, a member of the family Phyllanthaceae, primarily comprises shrub or tree species. It stands out from other genera due to distinctive reproductive features, including prolonged styles in flowers and fruits, apiculate anthers, and lobed and unlobed capsules. This study aimed to compare the morphological characteristics of nine taxa in Assam to facilitate identification and assess ethnobotanical knowledge. Ethnobotanical information was gathered by interviewing the local community, and a taxonomic key was provided for accurate identification. Morphological data underwent principal component analysis (PCA) and cluster analysis using PAST for validation. The comparison revealed distinct characteristics in both vegetative and reproductive traits among *Glochidion* members. Reproductive features, such as inflorescence, style, ovary, and capsules, were key factors for differentiation and identification. PCA and cluster analysis demonstrated correlation and variation among the taxa, contributing significantly to their demarcation. Ethnobotanical studies indicated the genus's potential medicinal properties, supported by both primary and secondary information.

Keywords: Angiosperms, cluster analysis, ethnobotany, PCA, Phyllanthoideae, taxonomy, UPGMA.

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Author contributions: PB has collected, done photography, identified, carried out the morphological analysis of the specimen, performed the PCA and cluster analysis and drafted the manuscript. SB contributed to the present study's design, supervised the work and revised the manuscript. The final manuscript was examined and approved by both the authors.

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INTRODUCTION

Northeastern India, including Assam, has a wide variety of vegetation due to its topographic and climatic diversity. It is one of the most diverse regions in the world (Dutta & Dutta 2005; Mao & Roy 2016; Bhattacharyya et al. 2020). The people of Assam have employed a number of plants to treat a wide range of ailments since ancient times (Kanjilal et al. 1940; Bhattacharya et al. 1991). There are many medicinal plants in the area that are well-known to ethnic communities, and Assam is regarded as one of the ecological hot spots in the world (Myers et al. 2000; Asati & Yadav 2004; Saikia et al. 2006).

The species of *Glochidion* J.R.Forst. & G.Forst. have been used by local people in different places in the world and have immense value in the field of medicine (Lai et al. 2004; Xiao et al. 2008; Bajpai et al. 2016; Chakrabarty & Balakrishnan 2018). Some important biological and pharmacological activities, including the anticancer, antioxidant, and antimicrobial activities of a few members of *Glochidion* have been reported by many workers (Azam et al. 2012; Rathod & Rajurkar 2017). The people of India mainly rely on medicinal plants and are well-known for ethnobotanical knowledge (Maikhuri & Gangwar 1993; Prakash et al. 2008). Therefore, it was felt worth exploring the genus *Glochidion* in Assam for its current taxonomy and to assess its ethnomedicinal uses.

The genus Glochidion J.R.Forst. & G.Forst. is a member of the family Phyllanthaceae which is native to northern Australia, Polynesia, southern Asia, and tropical Asia (Chakrabarty & Balakrishnan 2018). The members of the genus are either shrubs or trees, monoecious, pubescent, or glabrous with drooping branches. They are mostly found in evergreen, moist deciduous, tropical, primary and secondary forests, sal forests, hilly areas, and some swampy areas. There are over 320 species worldwide; about 22 species, and eight varieties in India (Balakrishnan & Chakrabarty 2007; Balakrishnan et al. 2012; Chakrabarty & Balakrishnan 2018; Brahma & Baruah 2023). Kanjilal et al. (1940) designated 16 species from erstwhile Assam in 'Flora of Assam'. At present 12 species and four varieties of the genus are found in Assam (Chakrabarty & Balakrishnan 2018). Traditionally, Glochidion was placed in Euphorbiaceae (Bentham & Hooker 1862–1883; Hutchinson 1973). Later, Hoffmann et al. (2006) discovered that the genus Glochidion sensu lato includes *Breynia* J.R.Frost & G.Forst., *Flueggea* Willd. and Margaritaria L.f., which are all allied to Phyllanthus as members of the tribe Phyllantheae and, therefore, belong to the segregate family Phyllanthaceae, and this was later ascertained by Chase et al. (2016) on the molecular basis. The absence of latex and the biovulate ovary distinguish the family Phyllanthaceae from Euphorbiaceae (Chakrabarty & Balakrishnan 2018).

Earlier in some floras, *G. ellipticum* used to be referred to as *G. assamicum*, a synonym of *G. ellipticum*; *G. velutinum*, i.e., synonym of *G. heyneanum*; *G. hirsutum* or *G. tomentosum*, i.e., synonym of *G. zeylanicum* var. tomentosum; and *G. arborescens*, i.e., synonym of *G. zeylanicum* var. arborescens (Hooker 1890; Kanjilal et al. 1940). According to recent literature and taxonomy data, the taxa *G. ellipticum*, *G. heyneanum*, *G. zeylanicum* var. tomentosum, *G. zeylanicum* var. arborescens are the accepted names (Chakrabarty & Balakrishnan 2018; WFO 2023).

The present study aimed to resolve the taxonomic identity of certain members of the genus *Glochidion* based on their morphology. The principal component analysis (PCA) and cluster analysis were carried out to authenticate the morphological evaluation of the taxa studied. All the relevant ethnobotanical information about the *Glochidion* taxa collected from Assam was documented based on primary sources that could yield potential information in the field of medical research.

MATERIALS AND METHODS

Sample collection, Identification, and Ethnobotanical assessment

Field surveys were conducted in the diverse forest areas in Assam from December 2019 to January 2023. Before conducting the field survey, approval was taken from Assam State Biodiversity Board (ASBB) and PCCF Wildlife Warden, Panjabari, Assam. Glochidion specimens were collected randomly from various locations in Assam, India (Table 1). During the field, collected specimens were taken in an airtight poly bag for further morphological examination and photographs of the specimens were taken using a Realme XT 64 MP mobile camera phone. Garmin GPS etrex 10 was used to record and identify precise latitudes and longitudes of the area of the specimen. In the lab, both vegetative and reproductive characteristics of freshly collected specimens were examined carefully under a Biocraft 20X simple microscope and a Leica EZ4W stereo microscope.

After a critical analysis of the character, identification was made with the help of some authentic literature (Hooker 1890; Kanjilal et al. 1940; Borthakur et al. 2018; Chakrabarty & Balakrishnan 2018), online taxonomic databases (e-Floras 2008; The Plant List 2013; GBIF



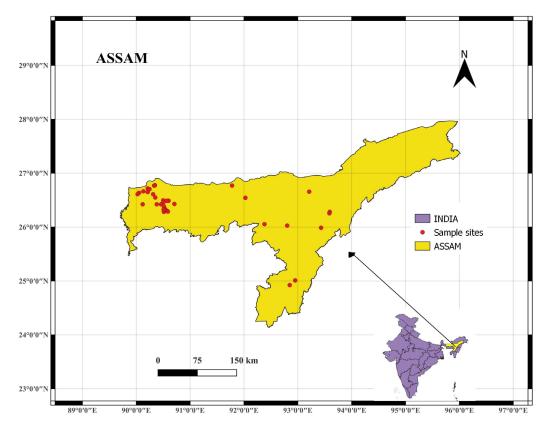


Figure 1. Distribution map of collected taxa from different places in Assam.

2021; POWO 2023) and also with the help of India herbaria (ASSAM, ARUN, and CAL) and digital herbaria (A, MO, NY) (acronyms following Thiers 2018). After reviewing pertinent literature, the threat status of the collected plant taxa was also determined (IUCN 2022). During identification, accepted scientific names and the synonyms of the collected taxa were also checked and confirmed through online databases such as IPNI, POWO, and The WFO Plant List (IPNI 2023; POWO 2023; WFO 2023). The dominant characters that played a key role in the identification of the specimen were their reproductive characters. The list of the collected specimens with their locality, accession number, GPS coordinates, and distribution map were procured (Table 1; Figure 1). The distribution map was created with QGIS 3.26.3 version software.

Ethnobotanical information of all collected taxa was made by the scrutiny of literature as well as communication with some local people and traditional healers in the study area. In addition to documenting the traditional uses and parts utilized for the specimen, we recorded their vernacular names, mode of preparation, application, and route of administration, as outlined in Table 5.

Herbarium preparation and deposition

Herbarium preparations adhered to the established techniques outlined by Jain & Rao (1977), while poisoning procedures followed the methods specified by Clark (1986). Authenticated and verified herbarium specimens for each collected taxon were deposited at the Botanical Survey of India (BSI) in Shillong, Meghalaya.

PCA and Cluster analysis

Fifteen morphological characters (Table 3) were analyzed based on using principal component analysis (PCA) and cluster analysis (Hammer et al. 2001). Multivariate PCA and hierarchical cluster analysis were assessed using the software PAST 4.06b version.

RESULTS

Glochidion: Morphological diagnosis

Monoecious and rarely dioecious; primarily of shrubs or trees; pubescent or glabrous; droopingly branched. Leaves simple, alternate, usually asymmetrical at the base, entire, petiolate, stipulate. The inflorescence is usually axillary, supra-axillary, or pedunculate, with few to many flowers. Staminate flowers are mostly long



Table 1. List of recorded taxa in the studied area with their locality, accession number, and GPS coordinates.

Таха	Locality	Accession No.	GPS coordinates
Glochidion ellipticum Wight	Kokrajhar District, Assam	98605	26.4947°N, 90.4319°E
G. heyneanum (Wight & Arn.) Wight	Kokrajhar District, Assam	98606	26.6236°N, 90.4061°E
G. lanceolarium (Roxb.) Voigt	Chakrashila Wildlife sanctuary, Kokrajhar District, Assam	98608	26.4236°N, 90.4963°E
G. multiloculare (Rottler ex Willd.) Voigt	Kokrajhar District, Assam	98604	26.7338°N, 90.4308°E
G. multiloculare var. pubescens Chakrab. & M.Gangop.	Orang National Park, Udalguri District, Assam	98610	26.7858°N, 92.3305°E
G. sphaerogynum (Mull.Arg.) Kurz	Chakrashila Wildlife Sanctuary, Kokrajhar District, Assam	98609	26.2902°N, 90.3747°E
G. zeylanicum var. arborescens (Blume) Chakrab. & M.Gangop.	Ultapani Forest Range, Kokrajhar District, Assam	98603	26.8002°N, 90.3466°E
G. zeylanicum var. tomentosum Trimen.	Ultapani Forest Range, Kokrajhar District, Assam	98607	26.7722°N, 90.4158°E
G. zeylanicum (Gaertn.) A.Juss	Nokpakghat, Karbi Anglong District, Assam	98611	26.3838°N, 93.2061°E

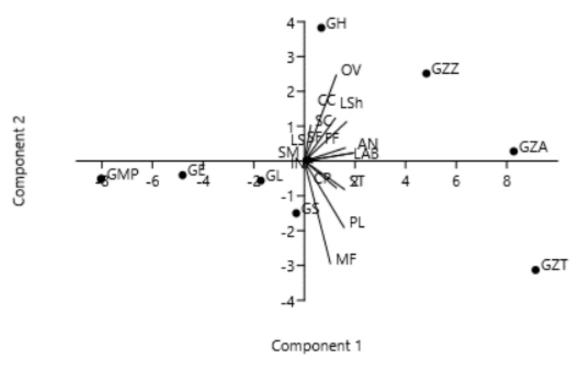


Figure 2. Multivariate Principal Component Analysis (PCA) of different members of *Glochidion* based on morphological characteristics.

pedicellate, with sepals 3–6, free, but no petals or disc. Anthers are present, 3–12, connective, pistillode absent. Pistillate flowers shortly pedicellate or sessile; sepals 3–6, free to connate; petals or disc absent; staminodes absent. Ovary 3–14 locular, biovulate locules; styles usually connate into a column, conical, or globose. Capsular pedicellate or sessile with a style column at the apex, depressed, subglobose, unlobed, and deeply or conspicuously lobed; pubescent or glabrous, green, white, or creamy to reddish. Seeds are usually 3–14, compressed, hemispherical with an arillate coat.

Review on the ethnobotanical knowledge of members of *Glochidion* in India

Some ethnobotanical uses of members of the genus *Glochidion* were mentioned by earlier workers. These are given below-

The paste of *Glochidion tomentosum* Dalz. is used externally in wounds by the tribes of Eastern Ghat, India (Reddy et al. 2006). The Chiru tribe of Manipur, India, consumed young leaves of *G. multiloculare* (Rottler ex. Willd.) Voigt and cooked them as an enjoyable curry and used them against stomach disorders (Rajkumari et

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Table 2. Comparative morphological characters of certain species of genus Glochidion collected from different localities of Assam.

Characters	G. ellipticum	G. heyneanum	G. lanceolarium	G. multiloculare var. multiloculare	G. multiloculare var. pubescens	G. sphaerogynum	G. zeylanicum var. arborescens	G. zeylanicum var. tomentosum	G. zeylanicum var. zeylanicum
Leaf shape	Elliptic to lanceolate, oblong to obovate	Ovate to elliptic, obovate	Lanceolate to oblanceolate, elliptic	Oblong to lanceolate, elliptic to oblanceolate	Oblong to lanceolate, elliptic to oblanceolate	Oblong to elliptic, falcate	Ovate to elliptic	Ovate to elliptic, cordate	Ovate to elliptic, cordate
Leaf apex & and base	Apiculate, caudate, acuminate at apex, obtuse at base	Acute, apiculate at apex, obtuse or rounded at base	Apiculate, acuminate, or acute at apex, obtuse or rounded at base	Acute, apiculate, or retuse at apex, obtuse or rounded at base	Acute, apiculate, or retuse at apex, obtuse or rounded at base	Acuminate at apex, attenuate at base	Acute, acuminate at apex, obtuse or rounded at base	Obcordate, acute at apex, obtuse, truncate, asymmetric at base	Acute, apiculate at apex, cordate, asymmetric, truncate at base
Leaf surface	Glabrous on both surfaces	Pubescent on both surfaces and densely pubescent beneath	Glabrous on both surfaces	Glabrous on both surfaces at mature and pubescent at young	Pubescent on both surfaces and densely pubescent beneath	Glabrous on both surfaces	Densely pubescent on both surfaces	Densely pubescent on both surfaces	Glabrous on both surfaces
Petiole length	0.4–1 cm long	0.1–0.5 cm long	0.6–1 cm long	0.1–0.5 cm long	0.1–0.5 cm long	0.9-1 cm long	0.3–0.5 cm long	0.5-0.7 cm long	0.1–0.8 cm long
Inflorescence	Axillary	Axillary	Axillary	Axillary	Axillary	Axillary	Supra-axillary, pedunculate, rarely axillary	Supra-axillary, pedunculate, rarely axillary	Supra-axillary, pedunculate, rarely axillary
Male flower	Pedicellate, 0.5– 1.7 cm long	Pedicellate, 0.5–1 cm long	Pedicillate, 0.9–2 cm long	Pedicellate, 0.5–1 cm long	Pedicellate, 0.5–1 cm long	Pedicellate, 0.5– 1.8 cm long	Pedicellate, 0.5– 0.7 cm long	Pedicellate, 0.7– 1.8 cm long	Pedicellate, 0.5–1 cm long
Sepal	9	9	9	6	6	9	9	9	9
Anther	4–5	3–4	4–6	5–12	5–12	3–5	5–7	5-8	3–8
Female flower	Pedicellate, 0.1– 0.5 cm long	Pedicellate, 0.1– 0.6 cm long	Sessile, 0.06–0.09 cm long	Pedicellate, 0.3– 0.5 cm long	Pedicellate, 0.3– 0.5 cm long	Pedicellate, 0.3– 0.5 cm long	Pedicellate, 0.4– 0.6 cm long	Pedicellate, 0.4–1 cm long	Pedicellate, 0.1– 0.9 cm long
Sepal	9	9	9	6–12	6–12	9	9	9	9
Style	Columnar, conical	Columnar	Columnar	Conical, subglobose	Conical, subglobose	Discoid	Columnar, persistent	Columnar, subconical	Free
Ovary	Subglobose, 3–6 locular	Depressed, subglobose, 4–5 locular	Depressed, subglobose, 5–8 locular	Depressed, 5-12 locular	Depressed, 5-12 locular	Depressed, 4–12 locular	Subglobose, 4–6 locular	Depressed, Subglobose, 5–8 locular	Depressed, Subglobose, 4–8 locular
Capsule	Pubescent, pedicellate, shallowly lobed	Pubescent, pedicellate, conspicuously lobed	Sparsely pubescent, sessile, shallowly lobed to deeply lobed	Pubescent, pedicellate, conspicuously lobed	Pubescent, pedicellate, conspicuously lobed	Glabrous, pedicellate, ambiguously lobed	Densely pubescent, pedicellate, obviously unlobed	Densely pubescent, pedicellate, ambiguously lobed	Glabrous, pedicellate, ambiguously lobed
Capsule color	Light green to creamy, whitish	Green to yellow- green	Light green, creamy to reddish	Green	Green	Green	Light green, yellow green to reddish	Light green, yellow green to reddish	Light green, yellow green to reddish
Seed color	Yellow green to red	Yellow green to orange	Yellow green to red	Green to red	Green to red	Green to orange	Yellow green to red	Yellow green to red	Yellow green to red



Table 2. Character state	of was walled a sign I also we at a vietic.	of different meaniness of	Glochidion used in cluster analysis.
lable 3. Character state	ot morphological characteristics	s ot aimerent members of	<i>Giochigion</i> used in cluster analysis.

Таха	LSh	LAB	LS	PL	INF	MF	SM	AN	FF	SF	ST	ov	СР	сс	sc
Е	1	1	2	1	0	1	0	1	1	1	1	1	1	1	1
Н	4	4	1	0	0	0	0	4	3	1	2	4	2	3	3
L	2	2	2	2	0	2	0	2	2	1	2	2	2	2	1
М	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MP	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
S	3	3	2	3	0	3	0	3	0	1	3	3	3	0	2
ZA	6	6	1	5	1	4	0	6	5	1	5	6	4	4	1
ZT	5	7	1	6	1	5	0	7	6	1	6	2	5	4	1
ZZ	5	5	2	4	1	0	0	5	4	1	4	5	3	4	1

E—G. ellipticum | H—G. heyneanum | L—G. lanceolarium | M—G. multiloculare var. multiloculare | MP—G. multiloculare var. pubescens | S—G. sphaerogynum | ZA—G. zeylanicum var. arborescens | ZT—G. zeylanicum var. tomentosum | ZZ—G. zeylanicum var. zeylanicum | LSh—Leaf shape | LAB—Leaf apex & base | LS—Leaf surface | PL—Petiole length | INF— Inflorescense | MF—Male flower | SM—Sepal of male flower | AN—Anther | FF—Female flower | SF—Sepal of female flower | ST—Style | OV—Ovary | CP—Capsule | CC—Capsule color | SC—Seed color

Character states: Leaf Shape: Oblong to lanceolate, elliptic to oblanceolate = 0; Elliptic to lanceolate, oblong to obovate = 1; Lanceolate to oblanceolate, elliptic = 2; Oblong to elliptic, falcate = 3; Ovate to elliptic, obovate = 4; Ovate to elliptic, cordate = 5; Ovate to elliptic = 6; Leaf apex & base: Acute, apiculate or retuse at apex, obtuse or rounded at base = 0; Apiculate, caudate, acuminate at apex, obtuse at base = 1; Apiculate, acuminate or acute at apex, obtuse or rounded at base = 2; Acuminate at apex, attenuate at base = 3; Acute, apiculate at apex, obtuse or rounded at base = 4; Acute, apiculate at apex, cordate, asymmetric, truncate at base = 5; Acute, acuminate at apex, obtuse or rounded at base = 6; Obcordate, acute at apex, obtuse, truncate, asymmetric at base = 7; Leaf surface: Glabrous on both surfaces at mature and pubescent at young = 0; Pubescent on both surfaces and densely pubescent beneath = 1; Glabrous on both surfaces = 2; Petiole length: 0.1-0.5 cm long = 0; 0.4-1 cm long = 1; 0.6-1 cm long = 2; 0.9-1 cm long = 3; 0.1-0.8 cm long = 4; 0.3-0.5 cm long = 5; 0.5-0.7 cm long = 6; Inflorescence: Axillary = 0; Supra-axillary, pedunculate, rarely axillary = 1; Male flower: Pedicellate, 0.5-1 cm long = 0; Pedicellate, 0.5-1.7 cm long = 1; Pedicillate, 0.9-2 cm long = 2; Pedicellate, 0.5-1.8 cm long 3; Pedicellate, 0.5–0.7 cm long = 4; Pedicellate, 0.7–1.8 cm long = 5; Sepal of male flower: 6 = 0; Anther: 5–12 = 0; 4–5 = 1; 4–6 = 2; 3–5 = 3; 3–4 = 4; 3–8 = 5; 5–7 = 6; 5-8 = 7; Female flower: Pedicellate, 0.3-0.5 cm long = 0; Pedicellate, 0.1-0.5 cm long = 1; Sessile, 0.06-0.09 cm long = 2; Pedicellate, 0.1-0.6 cm long = 3; Pedicellate, 0.1-0.9 cm long = 4; Pedicellate, 0.4-0.6 cm long = 5; Pedicellate, 0.4-1 cm long = 6; Sepal of female flower: 6-12 = 0; 6 = 1; Style: Conical, subglobose = 0; Columnar, conical =1; Columnar = 2; Discoid = 3; Free = 4; Columnar, persistent = 5; Columnar, subconical = 6; Ovary: Depressed, 5-12 locular = 0; Subglobose, 3-6 locular = 1; Depressed, subglobose, 5-8 locular = 2; Depressed, 4-12 locular = 3; Depressed, subglobose, 4-5 locular = 4; Depressed, Subglobose, 4-8 locular = 5; Subglobose, 4-6 locular = 6; Capsule: Pubescent, pedicellate, conspicuously lobed = 0; Pubescent, pedicellate, shallowly lobed = 1; Sparsely pubescent, sessile, shallowly lobed to deeply lobed = 2; Glabrous, pedicellate, ambiguously lobed =3; Densely pubescent, pedicellate, obviously unlobed =4; Densely pubescent, pedicellate, ambiguously lobed; Capsule color: Green = 0; Light green to creamy, whitish = 1; Light green, creamy to reddish = 2; Green to yellow green = 3; Light green, yellow green to reddish = 4; Seed color: Green to red = 0; Yellow green to red = 1; Green to orange = 2; Yellow green to orange = 3

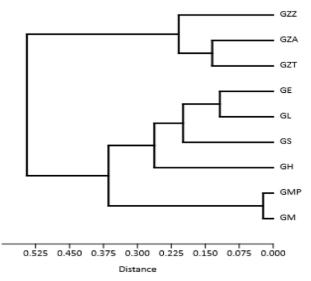


Figure 3. Paired group (UPGMA) dendrogram using hierarchical cluster analysis of different members of genus *Glochidion* based on their morphological characteristics.

al. 2013). The bark of *G. multiloculare* is used for skin diseases and wounds (Bajpai et al. 2016). Roots of *G.*

Table 4. Principal component analysis (PCA) based on morphological characteristics of different members of *Glochidion*.

PC	Eigenvalue	% variance
1	41.3263	83.254
2	4.25921	8.5804
3	2.02297	4.0754

multiloculare are used in snake bites (Brahma et al. 2002). The fruit and stem of *G. heyneanum* (Wight & Arn.) Wight is used in diabetes, fever, and bone fracture (Kumar et al. 2019). Roots of *G. heyneanum* are used in snake bites (Bajpai et al. 2016). Barks and leaves of *G. zeylanicum* (Gaertn.) A.Juss. are used in snake bites and stomach ulcers, and tender shoots are applied to itches (Das et al. 2013; Chakrabarty & Balakrishnan 2018; Kumar et al. 2019). Branches and leaves of *G. sphaerogynum* (Mull. Arg.) Kurz. are used in influenza and eczema (Lalrinkimi & Lallianthanga 2019). Fruits of *G. daltonii* (Mull. Arg.) Kurz. is used in cough and dysentery, and the bark of *G. ellipticum* Wight is used in inflammation (Bajpai et al.

(25)

Table 5. Enumeration of ethnobotanical knowledge of Glochidion in Assam.

Botanical name	Vernacular names	Parts used	Mode of preparation	Application	Route of administration
Glochidion multiloculare (Rottler ex Willd.) Voigt	Thakha Biphang or thakha mala (Bodo), Gorumora, Dolpoduli (Assamese)	Leaves, barks, Roots	 - A small number of leaves are ground into a paste. - A small amount of bark is taken and ground into a paste. - 3-4 roots are taken to make a paste. 	- Fracture and body swelling - Skin diseases and wounds - Snake bite	- External - External - External
<i>G. ellipticum</i> Wight	Thakha Biphang or thakha mala (Bodo), Panimadhuri (Assamese), Latimaowa (Nepali)	Bark, stem branches, and roots	- A small amount of bark is removed and ground into a paste, which is then administered to the diseased area A paste is made by crushing the stem and applying it to the swelling area Roots are ground into a paste.	- Body swelling, Skin problem - Body swelling - Snake bite	- External - External - External
G. sphaerogynum (Mull.Arg.) Kurz.	Thakha Biphang or thakha mala (Bodo), Panimadhuri (Assamese), Boljakru (Garo)	Young branches and leaves	Young branches and leaves are ground into a paste and blended with a small amount of water.	- Skin diseases - Branches are used as firewood also.	- External

2016). The paste made from the seeds of *G. ellipticum* is used as an antiallergic (Babu 1995). The bark of *G. lanceolarium* (Roxb.) Voigt is used in stomach diseases and is used as an anti-itch drug, oil made from seeds is also used as a source of light (Chanda et al. 2007; Bajpai et al. 2016; Chakrabarty & Balakrishnan 2018). The seeds of *G. calocarpum* Kuna are applied externally for skin diseases, and leaves are used orally to cure fever (Elanchezhian et al. 2007). According to Lalfakzuala et al. (2007), fruits of *G. arborescens* are used as wild edible fruits that are consumed by the local people of Mizoram. The fruits of *G. khasicum* (Mull.Arg.) Hook.f. are also edible and consumed by the tribal people of the Khasi hills in Meghalaya (Chakrabarty & Balakrishnan 2018).

DISCUSSION

The comparative morphological characteristics of certain species of the genus Glochidion showed many similarities and distinctive characteristics, which can be helpful for the identification and classification of the taxa (Table 2). Some of the major distinctive characteristics were leaf morphology, petiole length, and reproductive structures, i.e., inflorescences, male and female flowers, anthers, style, ovary, and capsule. The presence or absence of hairs on stems, leaves, inflorescence, and capsules also significantly differentiates the taxa. G. multiloculare var. pubescens an endemic variety of Assam showed nearly identical habit, vegetative and reproductive characteristics, with the exception of a glabrous plant body in G. multiloculare var. multiloculare (Chakrabarty & Balakrishnan 2018). The other major distinctive characteristics of both the taxa are that

solitary or individual flowers and fruits occur in each axil in *G. multiloculare* var. *pubescens* while multiple flowers and fruits in each axil of the plant body have been observed in *G. multiloculare* var. *multiloculare*. The variety *G. zeylanicum* var. *tomentosum* presented almost the same character as *G. zeylanicum* var. *zeylanicum*, with the major difference being its hairy or tomentose character. *G. heyneanum* showed puberulous habits on the stems and leaves. In some taxa, leaves were asymmetric or symmetric at the base.

The majority of taxa exhibited axillary inflorescence while G. zeylanicum var. zeylanicum, G. zeylanicum var. tomentosum and G. zeylanicum var. arborescens showed supra-axillary or pedunculate and rarely axillary inflorescence. Male flowers of the taxa revealed remarkably similar traits, but the number of anthers separated them. Female flowers presented different characteristics from male flowers. The peduncles of all the female flowers were shorter than the male flowers. The number of locules in the ovary varied by taxon, and style characters also played a key role. The shape, size, color, locules, and hairy habit of the capsule were distinguished among taxa, which showed taxonomic significance. Some taxa like G. zeylanicum var. zeylanicum and their varieties, were easily identified with their unlobed and ambiguously lobed capsule. G. multiloculare and G. sphaerogynum exhibited deeply or conspicuously lobed capsules while G. ellipticum presented a superficially lobed capsule. The capsule of G. lanceolarium was sessile, i.e., the fruit without the stalk or it lacked a pedicel, which distinguished it from other taxa. When dried, most of the leaves of the members were curled at the margin. While Glochidion and Epicephala moths were mutualists (Kato et al.



2003), most *Glochidion* leaves and drooping branches were found in insect-damaging conditions. As a result, some easily detectable characters for taxa identification could exist.

Based on morphological data, both PCA and cluster analysis were analyzed (Table 3-4 & Figure 2-3). The first PCA variance was 83.254% with an eigenvalue of 41.3263 followed by the second PCA variance of 8.5804% with an eigenvalue of 4.25921. The line connected to PC1 and PC2 makes up 91.8344% of the total variance and is a good sign of the variability of the initial data. PC1 represented the variation of the taxa based on the characters such as leaf shape (LSh), leaf apex and base (LAB), anther (AN), female flower (FF), sepal of the female flower (SF), ovary (OV), capsule color (CC), seed color (SC) and PC2 represented the characters such as leaf surface (LS), petiole length (PL), inflorescence (INF), male flower (MF), sepal of male flower (SM), style (ST), capsule (CP). In PC1, five taxa were observed i.e., G. ellipticum (GE), G. lanceolarium (GL), G. multiloculare var. pubescens (GMP), G. sphaerogynum (GS), and G. zeylanicum var. tomentosum (GZT). PC2 denoted a total of four taxa, viz., G. heyneanum (GH), G. multiloculare var. multiloculare (GM), G. zeylanicum var. arborescens (GZA), and G. zeylanicum var. zeylanicum (GZZ). From cluster analysis the tree revealed that G. multiloculare var. multiloculare (GM) and G. multiloculare var. pubescens (GMP) as cluster 1, G. heyneanum (GH) as cluster 2, G. sphaerogynum (GS) as cluster 3, G. ellipticum (GE) and G. lanceolarium (GL) as cluster 4, G. zeylanicum var. arborescens (GZA), and G. zeylanicum

var. tomentosum (GZT) as cluster 5, G. zeylanicum var. zeylanicum (GZZ) as cluster 6. Taxa present in the same cluster specified more correlation than the taxa present in the different clusters.

The ethnobotanical study revealed that some members of the genus *Glochidion* were traditionally used to cure different diseases (Table 5) in Assam. Among the uses, the most frequent are skin diseases, fractures, body swelling, and snake bites.

CONCLUSION

The study revealed that there are similarities and differences among the members of the genus Glochidion which are more reliable for grouping and classifying the taxa. Documentation of ethnobotanical evidence signifies the importance of the genus. Both primary and secondary sources of the ethnobotanical knowledge showed the members have medicinally important properties and almost all parts, i.e., leaves, bark, and roots, have been used by the local people for the treatment of various diseases in India including Assam. This study summarized that both taxonomical study and conservation of ethnobotanical knowledge are of great significance, with the ability to stimulate subsequent biological investigation. Moreover, PCA and cluster analysis also validated the data on comparative morphological traits that showed correlation and variation among the analyzed species.

Key to the species and varieties based on vegetative and reproductive characters

Inflorescence axillary Inflorescence axillary to supra-axillary	
Capsules shortly pedicellate Capsules sessile	3 G. lanceolarium
3. Leaves curl upwards with a margin when dry 3. Leaves do not curl upwards with a margin when dry	
4. Plant part glabrous except the reproductive organs 4. Plant parts all pubescent	
5. Capsules 3–6 locular; green to white creamy	
6. Fruits obscurely lobed	
7. Plants entirely glabrous	



 $Image \ 1. \ Morphological \ characters: A-Glochidion \ multiloculare \ var. \ multiloculare \ | \ B-G. \ ellipticum \ | \ C-G. \ heyneanum \ | \ D-G. \ lance olarium \ | \ D-G. \ l$ $| E-G. sphaerogynum | F-G. multiloculare \ var. \ pubescens | G-G. zeylanicum \ var. \ zeylanicum | H-G. zeylanicum \ var. \ arborescens | I-G. zeylanicum \ var. \ tomentosum. \\ @ Priyanka Brahma.$



REFERENCES

- **Asati, B.S. & D.S. Yadav (2004).** Diversity of horticultural crops in north eastern region. *ENVIS Bulletin: Himalayan Ecology* 12: 1–11.
- Azam, A.T.M.Z., A.H. Abdullah, G.U. Mohammad, M.M. Mohammad & M.H. Choudhury (2012). Antimicrobial, Antioxidant and Cytotoxic Activities of *Glochidion multiloculare* (Roxb. Ex Willd.) Mull. Arg. (Euphorbiaceae). *Dhaka University Journal of Pharmaceutical Sciences* 11(2): 117–120. http://doi.org/10.3329/dujps.v11i2.14560
- Babu, P.S.P. (1995). Euphorbiaceae of Andhra Pradesh, India. Ph.D. Thesis. Department of Botany, Sri Krishnadevaraya University, Andhra Pradesh, India.
- Bajpai, O., A. Kumar, A.K. Srivastava, A.K. Kuhwaha, J. Pandey & L.B. Chaudhary (2015). Tree species of Himalayan Terai region of Uttar Pradesh, India: a checklist. Checklist 11(4): 1–15. https://doi.org/10.15560/11.4.1718
- Balakrishnan, N.P. & T. Chakrabarty (2007). The Family Euphorbiaceae in India: A synopsis of its Profile, Taxonomy and Bibliography. M/s Bishen Singh Mahendra Pal Singh, Dehradun, India, 342 pp.
- Balakrishnan, N.P., T. Chakrabarty, M. Sanjappa, P. Lakshminarsimhan & P. Singh (eds.) (2012). Flora of India. Vol. 23. Botanical Survey of India, New Delhi, India, 414 pp.
- Bentham, G. & J.D. Hooker (1862–1883). *Genera Plantarum*. Vol. 3. L. Reeve & Co., 6 Henrietta Street, Covent Garden, London, England, 279 pp.
- Bhattacharya, P., R. Muzumder & G. Sarmah (1991). Rare medicinal plants of Assam. *Ancient Science of Life* 10(4): 234–8.
- Bhattacharyya, R., K.K. Medhi, S.K. Borthakur & S. Borkataki (2020). An ethnobotanical study of medicinal plants used against jaundice by tea tribes of Morigaon District, Assam (India). *Journal of Natural Remedies* 20(1): 16–28. https://doi.org/10.18311/jnr/2020/23879
- Borthakur, S.K., A. Bawri, D. Baro & A. Boro (2018). Flora of BTAD (Bodoland Territorial Area Districts, Assam). Vol. 3. EBH publishers, India, 74 pp.
- Brahma, B.K., B. Patere & H. Basumatary (2002). Boroni Muli Biphang Laiphang. Bodo Publication Board, Bodo Sahitya Sabha, Kokrajhar, Guwahati, Assam, 93 pp.
- Brahma, P. & S. Baruah (2023). Extended distribution of an endemic variety *Glochidion zeylanicum* var. *paucicarpum* Chakrab. & N.P.Balakr. (Phyllanthaceae) from Assam, India. *Vegetos*. https://doi.org/10.1007/s42535-023-00650-0
- Chakrabarty, T. & N.P. Balakrishnan (2018). Indo-Burmese Phyllanthaceae: A Taxonomic Revision. M/s Bishen Singh Mahendra Pal Singh, Dehradun, India, 194 pp.
- Chanda, R., J. Mohanty, N.R. Bhuyan, P.K. Kar & L.K. Nath (2007). Medicinal plants used against gastrointestinal tract disorders by the traditional healers of Sikkim Himalayas. *Indian Journal of Traditional Knowledge* 6(4): 606–610.
- Chase, M.W., M.J.M. Christenhusz, M.F. Fay, J.W. Byng, W.S. Judd, D.E. Soltis, D.J. Mabberley, A.N. Sennikov, P.S. Soltis & P.F. Stevens (2016). An update of the angiosperm phylogeny group classifications for the orders and families of flowering plants: APG IV. Botanical Journal of the Linnaean Society 181(1): 1–20. https://doi.org/10.1111/boj.12385
- Clark, S.H. (1986). Preservation of Herbarium Specimens: An Archive Conservator's Approach. *Taxon* 35(4): 675–682.
- Das, A.K., N. Stalin, C. Muthuperumal & P.S. Swamy (2013). Wild plants used by Muthuvan and Kattunaikkan tribal communities of Palakkayam settlement in Nilambur of Malappuram district, Kerala. Medicinal Plants International Journal of Phytomedicines and Related Industries 5(2): 82–89. http://doi.org/10.5958/j.0975-6892.5.2.013
- Dutta, B.K. & P.K. Dutta (2005). Potential of ethnobotanical studies in North East India: An overview. *Indian Journal of Traditional Knowledge* 4(1): 7–14.
- eFloras (2008). Missouri Botanical Garden, St. Louis, MO & Harvard University Herbaria, Cambridge, MA. http://www.efloras.org. Electronic version accessed 18 May 2021.

- Elanchezhian, R., R.S. Kumar, S. Beena & M.A. Suryanarayana (2007). Ethnobotany of Shompens - a primitive tribe of Great Nicobar Island. *Indian Journal of Traditional Knowledge* 6(2): 342–345.
- GBIF (2021). GBIF- Global Biodiversity Information Facility. https://www.gbif.org Electronic version accessed 24 May 2021.
- Hammer, O., D. Harper & P. Ryan (2001). PAST: Paleontological Statistics Software Package for Education and Data Analysis. Palaeontologia Electronica 4: 1–9.
- Hoffmann, P., H. Kathriarachchi & K.J. Wurdack (2006). A phylogenetic classification of Phyllanthaceae (Malpighiales; Euphorbiaceae sensu lato). Kew Bulletin 61: 37–53. http://www.jstor.org/stable/20443245
- **Hooker, J.D. (1890).** *The Flora of British India.* Vol. 5. L. Reeve & Co., Henrietta street, Covent Garden, London, 456 pp.
- **Hutchinson, J. (1973).** The Families of Flowering Plants. 3rd Edition. The Clarendon Press, Oxford, 519 pp.
- IPNI (2023). International Plant Name Index. Published on the Internet http://www.ipni.org. The Royal Botanic Gardens, Kew, Harvard University Herbaria & Libraries and Australian National Herbarium. Electronic version accessed 25 July 2023.
- IUCN (2022). The IUCN Red List of Threatened Species. Version 2021-3. https://www.iucnredlist.org Electronic version accessed 8 May 2022.
- Jain, S.K. & R.R. Rao (1977). A Handbook of Field & Herbarium Methods. Today & Tomorrow Printers & Publishers, New Delhi, India, 22 pp.
- Kanjilal, U.N., P.C. Kanjilal, R.N. Dey & A. Das (1940). Flora of Assam. Vol. 4. Prabasi Press, Upper Circular Road, Calcutta, 179 pp.
- Kato, M., A. Takimura & A. Kawakita (2003). An obligate pollination mutualism and reciprocal diversification in the tree genus Glochidion (Euphorbiaceae). Proceedings of the National Academy of Sciences, USA 100(9): 5264–5267. https://doi.org/10.1073/pnas.0837153100
- Kumar, J.U.S., M.J.K. Chaitanya, A.J. Semotiuk & V. Krishna (2019).
 Indigenous knowledge on medicinal plants used by ethnic communities of South India. Ethnobotany Research and Applications
 18: 1–112. https://ethnobotanyjournal.org/index.php/era/article/view/1291
- Lai, X.Z., Y.B. Yang & X.L. Shan (2004). The investigation of Euphorbiaceous medicinal plants in Southern China. *Economic Botany* 58: S307–S320. https://doi.org/10.1663/0013-0001(2004)58[S307:TIOEMP]2.0.CO;2
- Lalfakzuala, R., H. Lalramnghinglova & H. Kayang (2007). Ethnobotanical usage of plants in western Mizoram. *Indian Journal of Traditional Knowledge* 6(3): 486–493.
- Lalrinkimi & R.K. Lallianthanga (2019). Documentation of tree species within Mizoram Science Centre, Berawtlang, Aizawl, India with notes on their ethnomedicinal values. *Science Vision* 19(3): 63–78. https://doi.org/10.33493/scivis.19.03.01
- Maikhuri, R.K. & A.K. Gangwar (1993). Ethnobiological notes on the Khasi and Garo tribes of Meghalaya, Northeast India. *Economic Botany* 47: 345–357. https://doi.org/10.1007/BF02907348
- Mao, A.A. & D.K. Roy (2004). Ethnobotanical studies in northeast India: a review,) pp. 99–112. In: Jain, A.K. (ed.). Indian Ethnobotany: Emerging Trends. Scientific Publisher, India.
- Myers, N., R.A. Mittermeier, C.G. Mittermeier, G.A.B. da Fonseca & J. Kent (2000). Biodiversity hotspots for conservation priorities. Nature 403(6772): 853–858. https://doi.org/10.1038/35002501
- **POWO (2023).** Plants of the Word Online. Facilitated by the Royal Botanical Gardens, Kew. http://www.plantsoftheworldonline.org/ Electronic version accessed 1 August 2023.
- Prakash, J.W., R.D.A. Raja, N.A. Anderson, C. Williams, G.S. Regini, K. Bensar, R. Rajeev, S. Kiruba, S. Jeeva & S.M. Das (2008). Ethnomedicinal plants used by Kani tribes of Agasthiyarmalai Biosphere Reserve, southern Western Ghats. *Indian Journal of Traditional Knowledge* 7(3): 410–413.
- Rajkumari, R., P.K. Singh, A.K. Das & B.K. Dutta (2013). Ethnobotanical investigation of wild edible and medicinal plants used by the Chiru Tribe of Manipur, India. *Pleione*7(1): 167–174.
- Rathod, V. & N.S. Rajurkar (2017). Phytochemical screening and antioxidant activity of Glochidion ellipticum. Journal of Applicable



- Chemistry 6(2): 219–226.
- Reddy, S.R., K.N. Reddy, C. Pattanaik, V.S. Raju & J. Autonagar (2006). Ethnobotanical observations on some endemic plants of Eastern Ghats, India. *Ethnobotanical Leaflets* 10: 82–91.
- Saikia, A.P., V.K. Ryakala, P. Sharma, P. Goswami & U. Bora (2006). Ethnobotany of medicinal plants used by Assamese people for various skin ailments and cosmetics. *Journal of Ethnopharmacology* 106(2): 149–157. https://doi.org/10.1016/j.jep.2005.11.033
- The Plant List (2013). Version 1.1. http://www.theplantlist.org/. Accessed on 20 April 2021.
- **The World's Herbaria (2018).** A Summary Report Based on Data from Index Herbariorum Issue 3.0, published January 10, 2019, Barbara M. Thiers Editor, Index Herbariorum.
- WFO (2023). World Flora Online. http://www.worldfloraonline.org. Accessed on 22 June 2023.
- Xiao, H.T., H.P. He, J. Peng, Y.H. Wang, X.W. Yang, X.J. Hu, X.Y. Hao & X.J. Hao (2008). Two new norbisabolane sesquiterpinoid glycosides from *Glochidion coccineum. Journal of Asian Natural Product Research* 10(1-2): 1–5. https://doi.org/10.1080/10286020701189393



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Articles

Patterns of livestock depredation by carnivores: Leopard *Panthera pardus* (Linnaeus, 1758) and Grey Wolf *Canis lupus* (Linnaeus, 1758) in and around Mahuadanr Wolf Sanctuary, Jharkhand, India

- Shahzada Iqbal & Orus Ilyas, Pp. 24291-24298

Wetland biodiversity of Ramaroshan Lake complex: a need for conservation

Ram Devi Tachamo-Shah, Deep Narayan Shah, Subodh Sharma,
 Lila Sharma, Jagan Nath Adhikari & Deepak Rijal, Pp. 24299–24320

Diversity of wintering avifauna throughout the heterogeneous aquatic habitats of Bankura District, West Bengal, India

– Biplob Kumar Modak, Subha Shankar Mukherjee, Susobhan Mondal, Mainak Sarkar & Asif Hossain, Pp. 24321–24330

Assessing and understanding diversity and foraging guilds of bird community structure in Gautam Buddha Wildlife Sanctuary, Bihar and Jharkhand, India

Umar Saeed, Mujahid Ahamad, Vivek Ranjan, Syed Ainul Hussain
 Ruchi Badola, Pp. 24331–24344

Communications

Identifying potential habitats of Himalayan Red Panda *Ailurus* fulgens (Cuvier, 1825) (Mammalia: Carnivora: Ailuridae) in Neora Valley National Park, West Bengal, India

– Sangay W. Bhutia, Asim Giri, Pranita Gupta & Basavaraj S. Holeyachi, Pp. 24345–24351

Recent record of Eurasian Otter *Lutra lutra* (Linnaeus, 1758 (Mammalia: Carnivora: Mustellidae) from Kerala part of the Western Ghats, India and an insight into the behaviour and habitat preferences

– Sreehari K. Mohan, Lathish R. Nath, K.S. Subin, Sreekumar K. Govindankutty & P.O. Nameer, Pp. 24352–24356

A review of Baya Weaver *Ploceus philippinus* (Linnaeus, 1766) (Aves: Passeriformes: Ploceidae): ecological and conservation status

– Yusufkhan Pathan & Arvindgiri Goswami, Pp. 24357–24367

An updated checklist of non-marine molluscs of the western Himalaya

Hilal Ahmed, Imtiaz Ahmed & Neelavar Ananthram Aravind,
 Pp. 24368–24395

Nonessential elements (AI, As, Cd, & Pb) in shrimps and mussels from southeastern Brazil

 Ana Paula Madeira Di Beneditto, Inácio Abreu Pestana, Dayvison Felismindo Lima & Roberto Weider de Assis Franco, Pp. 24396– 24401

Three new additions to the flora of Himachal Pradesh, India from Khokhan Wildlife Sanctuary, Kullu District

Ashutosh Sharma, S. Noorunnisa Begum, G.S. Goraya, Gopal S.
 Rawat & Vaneet Jishtu, Pp. 24402–24408

Comparative morphological and ethnobotanical assessment of certain taxa of genus *Glochidion* (Phyllanthaceae) from Assam, India

- Priyanka Brahma & Sanjib Baruah, Pp. 24409-24419

Notes on *Discospermum sphaerocarpum* Dalzell ex Hook.f., a rare species of Rubiaceae (Ixoroideae: Coffeeae) from southern India

– C. Pramod, V.V. Drisya, A.K. Pradeep & K.T. Chandramohanan,Pp. 24420–24426

Legumes (Fabaceae) from Satmala hills, Maharashtra, India – Swapnil D. Wagh & Manoj T. Patil, Pp. 24427–24436

Report of new myristica swamp ecosystems from the Western Ghats at Pathanapuram, Kerala, India

- Niji Joseph, R. Sreejai & M. Ajayakumar, Pp. 24437-24442

Short Communications

First confirmed record of Arabian Gazelle *Gazella arabica*Lichtenstein, 1827 (Mammalia: Artiodactyla: Bovidae) on
Masirah Island, off the coast of eastern Oman in the Arabian Sea

– Taimur Al Said, Haitham Al Rawahi, Maha Al Ansari, Al Mutasim

– Taimur Al Said, Haitham Al Rawahi, Maha Al Ansari, Al Mutasim Al Hinai, Ahmed Al Amri, Ahmed Al Wahaibi, Ghasi Al Farsi, Saud Al Wihibi & Salman Al Farsi, Pp. 24443–24446

First report of the longhorn beetle, *Rosalia* (*Eurybatus*) *formosa* (Saunders, 1839) (Insecta: Coleoptera: Cerambycidae) from Mizoram, India

- Amit Rana & Lobeno Mozhui, Pp. 24447-24450

Publisher & Host



Threatened Taxa

APPENDIX 2

SPECIMENS DEPOSITED IN BSI, SHILLONG (ASSAM)



भारत सरकारं/GOVERNMENT OF INDIA

पर्यावरण वल एवंजलवायु परिवर्तन मंत्रालय/MINISTRY OF ENVIRONMENT, FOREST & CLIMATE CHANGE भारतीय वलस्पति सर्वेक्षण/BOTANICAL SURVEY OF INDIA प्रभारी वैज्ञानिक का कार्यालय/OFFICE OF THE SCIENTIST IN-CHARGE पूर्वी क्षेत्रीय केंद्र/EASTERN REGIONAL CENTRE



शिलांग-793003/SHILLONG - 793003

दूरभाष/Telephone: 0364- 2223971, 2223618

ई-मेल/e-mail- <u>bsibsishll@yahoo.co.in</u>

Telefax: 0364- 2224119

संख्या/No.: BSI/ERC/Tech/2022-23/ 2/90

दिनाक/Dated: 19.07.2022

सेवा मे/То.

Miss Priyanka Brahma Ph.D Research Scholar Department of Botany Bodoland University Kokrajhar, Assam- 783 370

विषय/Sub.: Accession of plant specimen and incorporation at ASSAM herbarium. reg.

Dear Miss Brahma,

With reference to your letter No. nil, dated 06/07/2022 regarding the subject cited above, I am to inform you that your plant specimen have been incorporated in ASSAM with accession number as below.

SI.	Coll. No.	Species	Family	Accession no.	
1.	001	Glochidion zeylanicum var. arborescens (Blume) Chakrab. & M.Gangop.	Phyllanthaceae	98603	
2.	002	2 Glochidion multiloculare (Rottler ex Willd.) Phyllanthac Voigt var. multiloculare		98604	
3.	003	Glochidion ellipticum Wight Phyllanthaceae 98605		98605	
4.	004	Glochidion heyneanum (Wight & Arn.) Wight Phyllanthaceae 98606 var. heyneanum		98606	
5.	005	en		98607	
6.	006	Glochidion zeylanicum (Gaertn.) A.Juss. var. Phyllanthaceae 98611 zeylanicum		98611	
7.	007	Glochidion lanceolarium (Roxb.) Voigt Phyllanthaceae 98608		98608	
8.	008	Glochidion sphaerogynum (Mull.Arg.) Kurz Phyllanthaceae 98609		98609	
9.	009	Glochidion multiloculare var. pubescens Phyllanthaceae 98610 Chakrab. & M.Gangop.		98610	

This is for your kind information and necessary office record.

Thank you / सधन्यवाद

(Dr. N. Odyuo)

वैज्ञानिक- E एवं कार्यालय प्रमुख/ Scientist-E & HoO

Figure 39. Deposited voucher specimens with accession number

SIRE TRAIT/GOVERNMENT OF INDIA

पर्यावरण वन एवजलवायु परिवर्तन अत्रालय/MINISTRY OF ENVIRONMENT, FOREST & CLIMATE CHANGE भारतीय वनस्पति सर्वेक्षण/BOTANICAL SURVEY OF INDIA



प्रभारी वैज्ञानिक का कार्यालय/OFFICE OF THE SCIENTIST IN CHARGE पुर्वी क्षेत्रीय केद्र/EASTERN REGIONAL CENTRE

द्राभाष Telephone: 0364- 2223971, 2223618

\$-ਸੋਰ e-mail- bsibsishll@yahoo co in

Telefax 0364-2224119

ист /No.: BSI FRC/ Tech/2023-24/1953

(4-us) /Dated, 11.08, 2023

सेवा मे/To, Priyanka Brahma Ph.D. Research Scholar C O-Dr. Sanjib Baruah Department of Botany Bodoland University Kokrajhar-783370, Assam

বিষয়/Sub.: Request for Accession Number of plant specimen-reg.

Dear Sir Madam.

With reference to your Ref. No. Nil dated 18th July 2023 regarding the subject cited above. I am to inform you that your plant specimens have been identified and confirmed as below with respective accession number.

Field No.	Name of the S	pecimen		Name of the Family	Accession Number	
010	Glochidion paucicarpum N.P.Balakr.	zeylanicum Chakrab.	var. &	Phyllanthaceae	099973	

This is for your kind information and office record. Thanking You,

भवदीय /Yours sincerely

(Dr. N. Odyuo)

वैज्ञानिक- ई एवं कार्यालय प्रमुख/ Scientist-E & H.o.O

वैज्ञानिक - ई एवं कार्यालय अध्यक्ष्य Scientist-E & H.O.O. भारतीय वनस्पति सर्वेक्कण Botanical Survey of India पु.क्षे के./Eastern Regional Centre शिलांग /Shillono-793003

Figure 40. Deposited voucher specimens with accession number

SEMINAR PRESENTATION

- 1. Assam Botany Congress (ABC-02) & International Conference on Plant Science organized by Botanical Society of Assam, Guwahati and Department of Botany, Cachar College, Silchar, Assam (2021) (**Figure 41**)
- National Seminar on Ethnobotany and Resource Management of the Bodos' organized by the Centre for Bodo Studies, Bodoland University, Kokrajhar, Assam (2021) (Figure 42)
- 3. National Seminar on Plant Taxonomy and Traditional Knowledge in the Himalayas and North-East India & Annual Conference of East Himalayan Society for Spermatophyte Taxonomy (EHSST) jointly organized by Department of Botany, Rajiv Gandhi University and East Himalayan Society for Spermatophyte Taxonomy, Botanical Survey of India (2022) (**Figure 43**)
- 4. International Seminar on Advances in Entrepreneurial Botany: Entrepreneurship Opportunities from Plant Resources for Sustainable Development organized by Department of Botany, University of Science & Technology Meghalaya in collaboration with CSIR-National Botanical Research Institute, Lucknow (2022) (Figure 44)
- 5. Bodoland International Knowledge Festival, 2023, India organized by Bodoland Territorial Region (BTR), Government & Bodoland University, Kokrajhar, Assam (2023) (**Figure 45**)
- 6. 2nd National Conference on Natural Products/AYUSH System of Medicine organized by Experimental Pharmacology Laboratory (EPL), Department of Pharmacology, PGIMER, Chandigarh under the aegis of Tridev Aushdiya Podh Utpadak Farmer Society, Rohal, IUPHAR-NP in collaboration with RCFC (NR-1) of National Medicinal Plant Board Ministry of AYUSH, Govt. of India, Joginder Nagar, Mandi, H.P (2024) (Figure 46)
- 7. National Seminar on Biodiversity for Sustainable Future with special focus on Plant Taxonomy and Annual Conference of East Himalayan Society for Spermatophyte Taxonomy (EHSST) organized by Department of Botany, Gauhati University, Assam (2024) (Figure 47)



Figure 41. Certificate of oral presentation in Assam Botany Congress (ABC-02) & International Conference on Plant Science



Figure 42. Certificate of oral presentation in the National Seminar on Ethnobotany & Resource Management of the Bodos'



Figure 43. Certificate of oral presentation in the National Seminar & Annual Conference

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intrepreneurial Botany: Ent Plant Resources for Sustain	repreneurship able Development"
red by: Department of Bota	ny 🥟
SCIENCE & TECHNOLOG	Y MEGHALAYA ited 'A' Grade by NAAC
CSIR-National Botanical Research Ins	titute Lucknow
ificate of Participa	tion PAA BESITE TOUCH
iyanka. Buahma	
	has participated in the International
al Botany: Entrepreneurship Opportunitie	
of Botany, USTM, in Collaboration with	•
Forst lo Gifforst. (Phyllanthaceae)	in driam.
We wish him/her all success in life.	0
(I)AD	NI Nava
113	/ ON3WC
Dr. Mautushi Das Convener	Prof S.K. Barik Director, CSIR-NBRI Lucknow
	Plant Resources for Sustainanted by: Department of Bota Science & Technolog Accredity CSIR-National Botanical Research Instificate of Participal Signaka. Breakman (Stamman Botany, Sakmajhan (Stamman Botany, USTM, in Collaboration with and presented a paper (Oral/Poster) entitled Forst to Griffinst (Phyllanthaseae) We wish him/her all success in life.

Figure 44. Certificate of Oral Presentation in the International Seminar



Figure 45. Certificate of 3rd prize of the poster presentation in BIKF, 2023, India



Figure 46. Certificate of Oral Presentation in the 2nd National Conference

	NATIONAL SEMI	NAR ON	
BIO	DIVERSITY FOR SUST	TAINABLE FUTURE	
WITH	SPECIAL FOCUS ON	PLANT TAXONOM	Y
	AND ANNUAL CONF	ERENCE OF	
EAST HIMALAYA	IN SOCIETY FOR SPERM	ATOPHYTE TAXONO	MY (EHSST)
	27th & 28th Septem Department of Botany, Ga		
	•		
C	ERTIFICATE OF PA	ARTICIPATION	
This is to certify that, Mr./Ms	/Prof./Dr. Priyanka Break	ma	0
Doct of Botany Bodolano	1 University part	icinated in the National Sen	ningr on Biodiversity fo
	ial focus on Plant Taxonomy as		
	hytochemical profiling in vita		
			•
Glochidion sphaenogynum (MWE NUG) NWCZ		••••••
40	^		
AMANDAY	of s	W	Meas
Prof. A.K. Pandey	Prof. P.P. Baruah	Prof. N. Devi	Dr. N. Nath
	Convener	President	Organizing Secretary

Figure 47. Certificate of Oral Presentation in the National Seminar and Annual Conference

WORKSHOP AND TRAINING

- One-Day Workshop on Research Manuscript Writing and Publication in Reputed Journal organized by the Centre for Bodo Studies, Bodoland University, Kokrajhar, Assam (2021) (Figure 48)
- Two Days National Workshop (Online) on Plant Identification and Herbarium Methodology jointly organized by Botanical Survey of India, Central National herbarium, Howrah and Bareilly College, Bareilly (U.P.) NSS Unit-I (Boys) (2021) (Figure 49)
- 3. One day Hands-on training on Plants Identification and Documentation organized by the Department of Botany, Bodoland University, Kokrajhar, Assam (2021) (Figure 50)
- 4. Two days National workshop on Green Biotechnology and Therapeutic Potential of Medicinal Plants jointly organized by Department of Botany and Zoology, S. S. Khanna Girl's Degree, Prayagraj (2022) (**Figure 51**)
- 5. Hands-on Workshop on Advanced Techniques in Natural Products & Medicinal Chemistry organized by CSIR-Indian Institute of Integrative Medicine, Jammu (2022) (Figure 52)
- 6. Awareness workshop on Access & Benefit Sharing (ABS) and Peoples' Biodiversity Register jointly organized by Assam State Biodiversity Board, Guwahati and Department of Zoology, Bodoland University, Kokrajhar, Assam (2022) (Figure 53)
- 7. Training Course on Classical & Modern Methods in Plant Taxonomy & Biosystematics organized by CSIR-National Botanical Research Institute, Lucknow (2024) (**Figure 54**)

Certificate No. くからいしょう	Date 23/10/21
	बर' फरायसं मिरु
	CENTRE FOR BODO STUDIES
ROGO	land University, Kokrajhar
	CERTIFICATE
	CERTIFICATE
This is to certify that Pr	of./Dr./Mr./Ms. Priyanka Brahma
	Scholan, B.U has participated in the one-day workshop
	SCRIPT WRITING AND PUBLICATION IN REPUTED
JOURNAL held on 7th O	ctober, 2021, organised by the Centre for Bodo Studies, Bodoland
University	
200	
Ladudingl	B. Probled Director
Vice-Chancellor	Director

Figure 48. Certificate of Worksop on Research manuscript writing



Figure 49. Certificate of Workshop on Plant Identification & Herbarium Methodology



Figure 50. Certificate of Hands-on training on Plant Identification & Documentation



Figure 51. Certificate of Workshop on Green Biotechnology and Therapeutic

Potential of Medicinal Plants



Figure 52. Certificate of High- End Workshop (KARYASHALA)

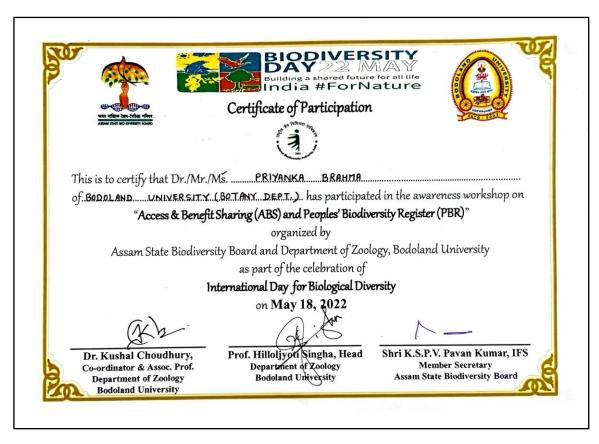


Figure 53. Certificate of Workshop on ABS & PBR



Figure 54. Certificate of Training Course on Classical & Modern Methods in Plant Taxonomy & Biosystematics

PERMISSION

For field surveys and collection of the specimens in different protected areas of Assam permission was taken from the Assam State Biodiversity Board (ASBB) and Principal Chief Conservator of Forests (PCCF), Chief Wildlife Warden, Panjabari, Guwahati, Assam (Figure 55 to Figure 58).

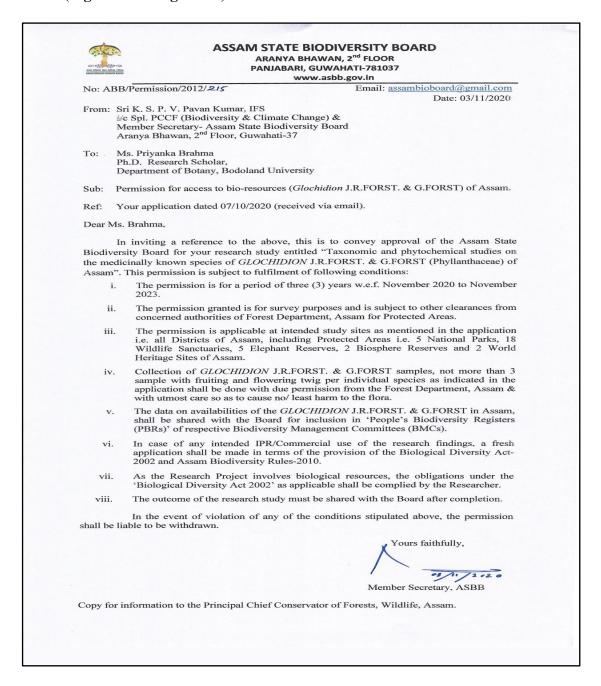


Figure 55. Permission copy from ASBB

GOVERNMENT OF ASSAM OFFICE OF THE PRINCIPAL CHIEF CONSERVATOR OF FORESTS (WILDLIFE) AND CHIEF WILDLIFE WARDEN, ASSAM: PANJABARI: GUWAHATI-37 No. WL/FG/31/Research .T.C. / 32rd T.C. 2022.

To,

Dr. Sanjib Baruah. Asst. Prof. (Stage II). Dept. of Botany, Bodoland University, Kokrajhar.

Taxonomic and Phytochemical studies on the medicinally known species of Glochidion J.R. Forest. & G. Forest. (Phyllanthaceae) of Assam.

With reference to the above mentioned subject, permission is hereby granted to you for a period of 1 (one) year from the date of issue of permission under Section 28 of the Wildlife (Protection) Act, 1972 for scientific research and other relevant provisions for entry. flora and fauna etc. subject to the terms and conditions as given below:

1. All the provisions relating to the National Parks and Sanctuaries under the Wildlife (Protection) Act, 1972

shall be strictly adhered to.

Entry into NP/WLS after sun-set and before sunrise is prohibited.

No boundary mark of the Protected Areas (PA) shall be damaged, altered, destroyed, moved or defaced.

No other wild animal shall be teased, molested or disturbed.

- No other wild animal shall be teased, molested or disturbed. There should not be any damage to any flora or fauna, and no floral or faunal sample should be collected, unless specified otherwise.

 The ground of the Protected Areas shall not be littered.
 A Research Monitoring Officer (RMO) authorised by the PA authorities shall supervise the activities to ensure the adherence of all the conditions stipulated herein.

 The park authorities shall not take responsibility for arrangement of food, lodging and conveyance.
 A copy of interim progress report, both hard copy and soft copy, shall be submitted by you to this office and also to the DFO/PA Authority for the extension of the project, and three copies of final report along with a soft copy shall be furnished to the CWLW, Assam for office record immediately after completion of the work.
- of the work.

 10. In case of research period more than one year, the researcher shall provide interim progress report for the progress of research work and make a Presentation before Park Manager once in six months failing which the permit issued shall be cancelled.

 11. An amount of Rs. 10,000/- will have to be deposited in the form of a "fixed deposit" pledged in favour of the Chief Wildlife Warden, Assam, Panjabari, Guwahati-37 as a security deposit which will be released immediately after fulfilling the clause 9 and also on receipt of the NOC from P.A. authority about satisfactory compliance of all the above stipulations and also an amount of Rs.1,000/- as Govt. Chalan under head of accounts:0406-02-800 will be deposited by the applicant which is non-refundable.
- 12. The Park Authority reserves the right to cancel/terminate this permission at any time, wherever it is considered that the activities resulting from this permission are affecting the flora and fauna adversely or the permit-holder is not abiding by the stipulations contained herein, with the prior sanction of the Chief Wildlife Worden.
- 13. Entry to the Protected Areas would be as per the convenience of the local forest authority and a Research register has to be maintained by the PA Managers at Range level for entering in to the PA with authentication of the local forest authority, mentioning the entry and exit timing along with signature of
- authentication of the local forest authority, mentioning the entry and exit timing along with arguments or researcher at both the timing.

 14. Movements of the researcher and his assistants in the PA shall be recorded in a log book duly countersigned by the local forest authority and has to be maintained by him. The log will be submitted to the Park Manager every month.

 15. Collection of parts of plants for herbarium, anatomical study and phytochemical screening etc. may be allowed as per provision of Section 29 of the Wildlife (Protection) Act. 1972.

 16. The applicant shall have to deposit the herbarium collection, to Assam State Zoo Division, Guwahati.

 17. An MoU have to be executed between this office and the applicant.

If agreed to all the above stipulations and on furnishing the required documents, MoU and security deposit the permission will be granted. An undertaking will have to be signed by the applicant before obtaining the permission for entering into the Protected Area.

Principal Chief Conservator of Forests, Wildlife & Chief Wildlife Warden, Assam

- Copy for information and necessary action to:
 1. The Field Director, Kaziranga National Park & Tiger reserve, Bokakhat
 2. The Divisional Forest Officer, Western Assam Wildlife Division, Tezpur.
 3. The Divisional Forest Officer, Mangaldoi Wildlife Division, Mangaldoi.
 4. The Divisional Forest Officer, Assam State Zoo Division, Guwahati-05.
 5. Ms. Priyanka Brahma, Research Scholar, Bodoland Univ Ms. Priyanka Brahma, priyabrahma659@gmail.com University Kokrajhar, Email:

Principal Chief Conservator of Forests, Wildlife & Chief Wildlife Warden, Assam

UNDERTAKING

I do hereby undertake that I shall abide by all the stipulations contained in this permission and I shall enter into the PA at my own risk and in case of any violation of any of the stipulations, I shall be liable to be prosecuted under the relevant provisions of law.

Priyan Ka Brahma Signature of the applicant

Figure 56. Permission copy from PCCF



GOVERNMENT OF ASSAM OFFICE OF THE DIVISIONAL FOREST OFFICER: EASTERN ASSAM WILDLIFE DIVISION, KAZIRANGA NATIONAL PARK, BOKAKHAT

Letter No. B/W/2022/ Research & Permission/ 767/-77

The Range Officers-

- Kaziranga Range, Kohora
 Western Range, Bagori
- Burapahar Range, Ghorakati
 Eastern Range, Agoratoli
- 5. Bokakhat Range, Bokakhat

Sub: Permission to carry out Ph.D. work on "Taxonomic and Phytochemical studies on the medicinally known species of Glochidion J.R. Forest & G. Forest (Phyllanthaceae) of

The PCCF (Wildlife) & CWLW, Assam Order no. 258 Ref:

Dated.02.09.2022

Please find herewith a photocopy of letter received from the Principal Chief Conservator of Forests, Wildlife & Chief Wildlife Warden, Assam, Aranya Bhawan Panjabari Guwahati-37 which is self explanatory for your information and needful action.

You are requested to provide necessary support and at the same time ensure that existing rules and regulations in force are followed.

Enclo: As stated above.

Divisional Forest Officer Eastern Assam Wildlife Division **Bokakhat**

<u>Copy to</u>:

1.Shri Rabindra Sarmah, Wildlife Research Officer, EAWL Division, Bokakhat for his

2. Miss Priyanka Brahma, Research Scholar, Botany Department, Bodoland University, Kokrajhar, Assam for her information and necessary action. He is requested to submit contact no and address to the respective Range Officers before entry in to the Park.

> Divisional Forest Officer Eastern Assam Wildlife Division Bokakhat

Memo No: A/WLG/ Research & Permission /2022/ 3 8 0 5 Date: / 3 .09.2022 Copy to the Field Director, Kaziranga National Park & Tiger Reserve, Bokakhat for favour of his kind information.

Divisional Forest Officer Eastern Assam Wildlife Division Bokakhat

Figure 57. Permission copy from PCCF



Email ID: cwlw.assam@gmail.com

Office Order No. 258

Dated Guwahati the . 02/09/2022.

The Permission to carry out Ph.D. work on "Taxonomic and Phytochemical studies on the medicinally known species of Glochidion J.R. Forest. & G. Forest. (Phyllanthaceae) of Assam" to be conducted in Kaziranga National Park, Orang National Park, Nameri National Park and Borail Wildlife Sanctuary in Assam is hereby accorded to Miss. Priyanka Brahma, Research Scholar, Botany Department, Bodoland University, Kokrajhar, Assam for a period of one year from the date of issue of the order under Section 28 of the Wildlife (Protection) Act, 1972 (as amended up to date) on submission of the undertaking to abide by all the stipulations laid down as communicated vide this office letter No. WL/FG/31/Research .T.C/32nd T.C.2022, dtd. 30/07/2022. The petitioner deposited an amount of Rs. 10,000/-(Rupees Ten Thousand) only as security deposit vide Account No. 5276166545, dated 17/08/2022, Central Bank of India, Kokrajhar, Assam, India pledged in favour of the Chief Wildlife Warden, Assam which is refundable and also deposited a non-refundable amount of Rs. 1000/- (Rupees One Thousand) vide Treasury Challan No.22/08/1611, dated 17/08/2022, SBI Bank, Kokrajhar only as revenue. The permission may be extended for the period as requested in the initial application subject to submission of progress report to this office.

(M.K. Yadava, IFS) Chief Wildlife Warden, Assam

Memo No. WL/FG/31/Research .T.C/32nd T.C.2022

Dated Guwahati the. 02 /09 /2022.

Copy for information and necessary action to:-

- 1. The Field Director, Kaziranga National Park & Tiger Reserve, Bokakhat.
- 2. The Divisional Forest Officer, Western Assam Wildlife Division, Tezpur.
- 3. The Divisional Forest Officer, Mangaldoi Wildlife Division, Mangaldoi.
- 4. The Divisional Forest Officer, Assam State Zoo Division, Guwahati.
- Dr. Sanjib Baruah, Asst. Prof. (Stage II), Department of Botany, Bodoland University, Korajhar.
- Miss. Priyanka Brahma, Research Scholar, Department of Botany, Bodoland University Korajhar, Email: priyabrahma659@gmail.com

7. Office Order Book.

Chief Wildlife Warden, Assam

Figure 58. Permission copy from PCCF