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APPENDIX-A

Table 9.1: Showing different nutrient compositions used in MS media

Macronutrients	Chemical formula	g/L (Concentration)
Ammonium nitrate	NH_4NO_3	16.5
Potassium nitrate	KNO_3	19
Calcium chloride	$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$	4.4
Magnesium sulphate	$\text{MnSO}_4 \cdot 4\text{H}_2\text{O}$	3.7
Potassium dihydrogen orthophosphate	KH_2PO_4	1.7
Micronutrients		
Manganese sulphate	$\text{MnSO}_4 \cdot 5\text{H}_2\text{O}$	2.23
Zinc sulphate	$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	0.86
Potassium iodide	KI	0.086
Cupric sulphate	$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	0.0026
Sodium molybdate	$\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$	0.025
Cobalt chloride	$\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$	0.0026
Boric acid	H_3BO_3	0.62
Vitamins		
Nicotinic acid	$\text{C}_6\text{H}_5\text{NO}_2$	0.05
Thiamine hydrochloride	$\text{C}_{12}\text{H}_{17}\text{ClN}_4\text{OS} \cdot \text{HCl}$	0.01
Pyridoxine hydrochloride	$\text{C}_8\text{H}_{12}\text{N}_2\text{O}_2 \cdot 2\text{HCl}$	0.05
Glycine	$\text{C}_2\text{H}_5\text{NO}_2$	0.2
Iron source		
Sodium EDTA	$\text{C}_{10}\text{H}_{14}\text{N}_2\text{O}_8\text{Na}_2\text{H}_2\text{O}$	2.78
Ferrous sulphate	$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	3.72
Myo-inositol		0.1g
Sucrose		30g
Phytigel		2g

APPENDIX-B

B: Chemicals Used

Acetic acid	Magnesium chloride
Acetone	Manganese sulphate
Agar agar	Mercuric chloride
Agarose	Methanol
Ammonium nitrate	MS media
Ascorbic acid	Myo-inositol
Bavistin	Nicotinic acid
Boric acid	Nucleotide
Calcium chloride	PCR master mix
Cobalt chloride	Phytigel
CTAB buffer	Potassium dihydrogen phosphate
De-ionized water	Potassium nitrate
Disodium molybdate	Primer
DPPH	Pyridoxine hydrochloride
Ethanol	Quercetin
Ethidium bromide	Sodium EDTA
FCR	Sucrose
Ferrous sulphate	Taq Polymerase
Gallic acid	Thiamine hydrochloride
Glycine	

C: Instruments used

Autoclave (Optics technology)
Laminar air flow (Optics technology)
Growth chamber (Bhanu Biotech)
Thermal cycler (Applied Biosystem)
HPLC (waters)
Magnetic stirrer (optics technology)
Lyophilizer (Telstar lyoquest freeze dryer)
Weighing balance
Incubator (Optics technology)
Hot air oven (Optics technology)
Centrifuge (RV/FM, super spin, plastocraft, India)
Deep freezer -80°C (Blue star, CRESCENT)
Water bath incubator shaker (Remi Model No. KWBS-2)
Refrigerator (Godrej)
Mili Q (Ellix)
Micropipette (Eppendrof)
pH meter (Elico pH meter LI617)

D: Abbreviations

MS: Murashige and Skoog

IAA: Indole-3-acetic acid

IBA: indole-3-butyric acid

NAA: 1-naphthaleneacetic acid

2,4-D: 2,4-dichlorophenoxyacetic acid

BAP: 6-benzylaminopurine

GA₃: gibberellic acid

SE: standard error

ANOVA: Analysis of variance

BM: Basal media

RAPD: Random amplified polymorphic DNA

GAE: Gallic acid equivalent

QE: Quercetin equivalent

AAE: ascorbic acid equivalent

HPLC: high pressure liquid chromatography

RP: Reverse phase

GC-MS: gas chromatography-mass spectroscopy

NMRC: nuclear magnetic resonance cryoporometry

PSD: pore size distribution

FCR: folin-ciocalteu reagent

DPPH: 2,2-diphenylpicrylhydrazyl

RT: retention time

TDZ: Thidiazuron

EtBr: Ethidium bromide

F: Research articles published in Scopus and Web of science

Research article I



RESEARCH ARTICLE

In vitro propagation and plant regeneration of *Torenia crustacea* (L.) Charm. & Schltld; an important ethnic medicinal plant

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Baro T, Das S. *In vitro* propagation and plant regeneration of *Torenia crustacea* (L.) Charm. & Schltld; an important ethnic medicinal plant. Plant Science Today. 2022; 9(sp2): 30-35. <https://doi.org/10.14719/pst.1708>

Abstract

Torenia crustacea (L.) Cham. & Schltld is an important medicinal herb used in India, Indonesia and Malaysia. This herb is used by Bodo tribes in the Bodoland Territorial Region, Assam to treat various diseases like diabetes. The present experiment developed an efficient protocol for *in vitro* mass propagation technique for *Torenia crustacea* using its nodal explants. For tissue culture, the rapidly growing nodal explants of *Torenia crustacea* were used. Effective explant surface sterilisation was found at 15 min. of treatment with 2% sodium hypochlorite resulting in a maximum explant survival rate and a lower rate of explant contamination after 21 days of explant culture. Media containing full strength MS (Murashige and Skoog's) + BAP (6-Benzyl amino purine) (1 mg/l) were found most effective for the establishment of the explant. The highest shoot proliferation and multiplication were observed in the media containing full strength MS + BAP (1 mg/l) + NAA (Naphthalene acetic acid) (0.2 mg/l); in this combination, an average of 16 shoots formed per nodal explant. The rooting of explants was observed highest in MS media along with 0.4 mg/l concentration of NAA and 1 mg/l BA. The *in vitro* multiplied shoots were grown in plastic pots containing vermicompost fertiliser and soil mixture and successfully grown in the open field condition.

Keywords

Explant culture, MS media, shoot multiplication, surface sterilisation, tissue culture

Introduction

Numerous traditional medicinal plants exist in nature in different regions of India. The North-Eastern states of India are one of the richest areas of medicinal and aromatic plant resources in the world (1) and most of the medicinal plants are unexplored. Due to the high demands of crude drugs, the medicinal plants are being overexploited, resulting in the threat for many rare species and many important plants are disappearing from nature (2). Because of various anthropogenic activities, many species are on the way to extinction without being properly explored or identified which could have immense medicinal property. Hence, identifying and preserving these plants are very important as they are widely used by people. Plant tissue culture is one of the fundamental tools for plant science and is continuously employed in mass propagation, plant improvement and conservation of plant species (3). Plant tissue culture or micropropagation is one of those promising techniques for the rapid growth of the plant tissue and plant organ under sterile conditions in a culture medium. The propagation is uni-

Research article II



RESEARCH ARTICLE

In vitro propagation and somaclonal variation study of *Phlogacanthus thyriformis* Nees an ethnic medicinal plant

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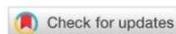
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Abstract

Bodoland Territorial Region is very rich in natural bioresources, and the Bodo tribes of the region use numerous ethnic medicinal plants to treat different kinds of diseases. *Phlogacanthus thyriformis* Nees is an important medicinal shrub species used in the region to treat different kinds of diseases. The objectives of the present study were to develop an efficient *in vitro* mass propagation technique of the species using nodal explant and study genetic stability in the genome of *in vitro* propagated plantlets by different RAPD markers. Effective explant surface sterilisation resulted at 2 min of treatment with 0.1% mercuric chloride. Explant responses were found most effective in the full strength MS + 1mg/l BAP (6- benzyl amino purine), and explant highest shoot proliferation multiplication and rooting were found in the media MS + 1 mg/l BAP + 0.250 mg/l NAA. *Ex vitro* rooting of micro propagated plants was most effective when the explants were dipped in 1 mg/l IBA for an hr. RAPD assays were conducted using eight sets of random primers (OPC02, OPC05, OPC07, OPC08, OPC09, OPX06, DK2 and OPA01). All the primers except OPC07 and OPA01 formed monomorphic DNA bands in gel electrophoresis and polymorphism was detected by OPC07 and OPA01 primer.

Keywords

explants culture, molecular marker, MS media, plant regeneration, tissue culture, RAPD

Introduction

The global population mostly relies on ethnic medicine (about 80% of the total population) for primary healthcare, most of which is taken from plant extracts (1). Traditional medicines play a crucial role in the health care and treatment of diseases in most developing countries (2). *Phlogacanthus thyriformis* Nees (Barsukha) is a shrub species of the Acanthaceae family. It is 3-7 feet in height. It is used as folk medicine in Assam and its distribution ranges from Bhutan to the North-Eastern states of India in the Himalayas and Indo-China, southern China and Sulawesi (3). The shrub is used to treat allergies, cough, cold, chronic bronchitis, rheumatoid arthritis, asthma etc. also the leaf extract is used as anti-bacterial and also effective in inhibiting the growth of HeLa cells (3). From the survey, it was found that the flowers of the plant are also consumed by local peoples considering medicinal importances like- anti gastric, anti- diabetes, anti-cough to treat cold fever.

Because of the presence of bioactive potential in medicinal plants, their demands are being higher day after day. Further, due to deforestation,

G: Paper presentation in international and national seminar

Paper presented in National seminar entitled “*in vitro* propagation and study of genetic stability of *Phlogacanthus thyrsoformis*; an important medicinal plant species’



Paper presented in international conference entitled “ *In vitro* propagation and plant regeneration of medicinal herb *Enydra fluctuans*”



The certificate features a decorative border with green leaves on the left and right sides. At the top left is the Department of Biotechnology logo, and at the top right is the NEHU logo. The central text is in bold, black, uppercase letters. The theme and dates are in a smaller font. The main body of the certificate is a paragraph certifying participation, with handwritten details in blue ink. At the bottom, there are four signatures and their corresponding titles.

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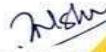
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